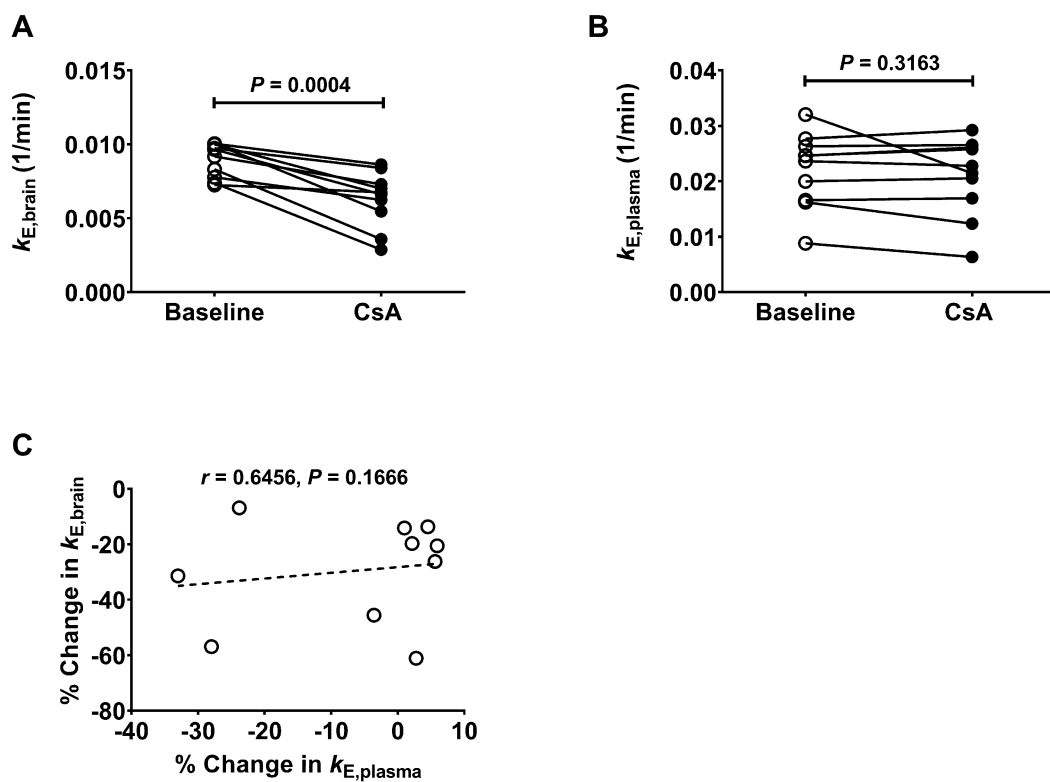


## Determination of Cyclosporine A Concentrations in Blood

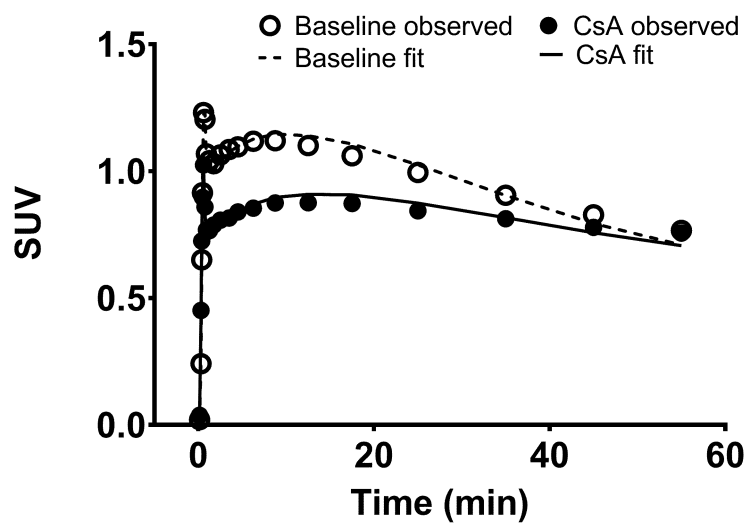
The concentration of cyclosporine A in whole blood was determined by high-performance liquid chromatography (HPLC) with minor modifications as described previously (*1*). Briefly, after hemolysis of 1 mL whole blood by the addition of a mixture of zinc sulfate/methanol (65:35, w/v; 2 mL), 50  $\mu$ L (1.0  $\mu$ g) of cyclosporine D (Cayman Chemical, Ann Arbor, MI, USA) was added as the internal standard. After centrifugation (5 min, 3,000 x g), the supernatant was passed through an Oasis HLB 1 cc SPE cartridge (30 mg; Waters Corporation, Milford, MA, USA) which had been equilibrated with 2 mL of methanol and water (pH 3.0), respectively. The column was washed with methanol in water (50%, v/v; 2.3 mL) and heptane (0.5 mL), and cyclosporine A was eluted with ethanol (100%, 300  $\mu$ L). The eluate was mixed with 100  $\mu$ L of water (pH 3.0) and 1 mL of heptane and centrifuged for 5 min at 3,000 x g. An aliquot (80  $\mu$ L) of the aqueous phase was injected onto the HPLC column. HPLC was performed using a Dionex “UltiMate 3000” system (Dionex Corp., Sunnyvale, CA) with UV detection at 205 nm. Chromatographic separation was carried out at 75°C on a Hypersil BDS-C18 column (5  $\mu$ m, 250 x 4.6 mm, Thermo Fisher Scientific, Inc, Waltham, MA), preceded by a Hypersil BDS-C18 pre-column (5  $\mu$ m, 10 x 4.6 mm). The mobile phase consisted of a continuous gradient, mixed from solvent A (acetonitrile in water [35:75, v/v]) and solvent B (acetonitrile in water [85:15, v/v]). The column was equilibrated with 45% solvent B at time 0; after injection of the sample (80  $\mu$ L), the content of solvent B was linearly increased to 93% at 19 min. Subsequently, the percentage of solvent B was decreased to 45% within 2 min, to equilibrate the column for 8 min before application of the next sample. Linearity was tested by assaying drug-free whole blood spiked with 0.1, 0.5, 2.5 and 10  $\mu$ g cyclosporine A. The calibration curve for cyclosporine A in blood was linear over the tested concentration range (correlation coefficient: 0.985). Quantification of cyclosporine A was based on the comparison of cyclosporine A/cyclosporine D ratios.

### **Determination of Parent $^{11}\text{C}$ -metoclopramide in Plasma**

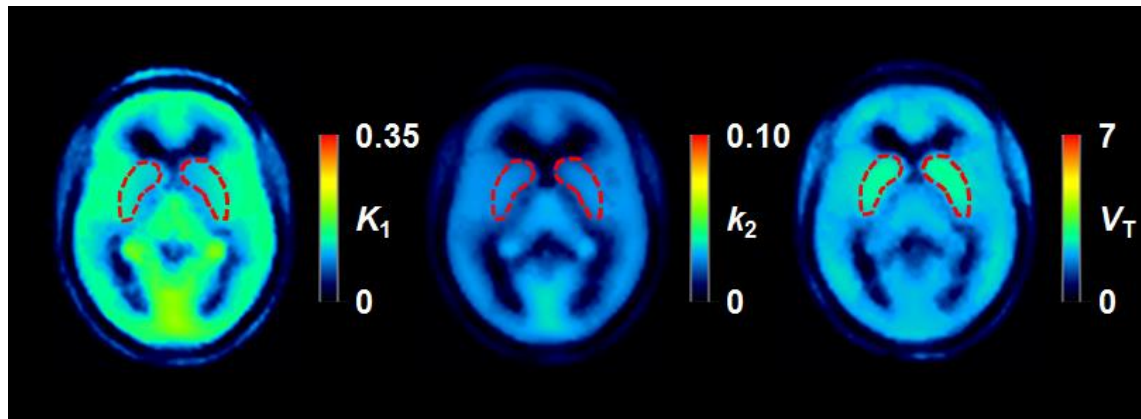
Plasma samples (830  $\mu\text{L}$ ) collected at 5, 10, 20, 30 and 40 min after  $^{11}\text{C}$ -metoclopramide injection were mixed with acetonitrile (600  $\mu\text{L}$ ) and vortexed to precipitate plasma proteins. After addition of water (600  $\mu\text{L}$ ) and phosphate-buffered saline (10-fold concentrate, pH 7.4, 100  $\mu\text{L}$ ) samples were centrifuged (4 min, 15,000  $\times$  g, 4°C) and the protein pellet and supernatant were separately counted in a gamma counter to determine the recovery of radioactivity, which was for the 10 min time point  $87 \pm 2\%$  for the baseline scan and  $88 \pm 2\%$  for the CsA scan ( $n = 9$ ). The supernatant (1.5 mL) was then injected into the HPLC system. An Atlantis T3 OBD HPLC column (250  $\times$  10 mm, 10  $\mu\text{m}$ , Waters, Austria) equipped with a pre-column (Atlantis T3 Prep Guard Cartridge, 10  $\times$  10 mm, 10  $\mu\text{m}$ ) was eluted with a mixture of 25 mM aqueous ammonium acetate (solvent A) and acetonitrile (solvent B). A linear gradient from 20% to 30% of solvent B over 5.5 min (total run length: 10 min) was applied to the column at a flow rate of 5 mL/min. On this HPLC system  $^{11}\text{C}$ -metoclopramide and its radiolabeled metabolites eluted with retention times of approximately 8.5 and 4 min, respectively. HPLC eluates were collected in 1-min fractions, which were counted for radioactivity in a gamma counter. The measured fractions were corrected for radioactive decay to determine the percentage of unmetabolized  $^{11}\text{C}$ -metoclopramide in plasma at different time points.



**SUPPLEMENTAL FIGURE 1.** Individual  $k_{E,brain}$  values in whole brain grey matter (A) and  $k_{E,plasma}$  values (B) for scans without (baseline) and with CsA infusion. Shown  $P$  values are from two sided, paired t-test. Correlation of percentage change in  $k_{E,brain}$  and  $k_{E,plasma}$  in response to CsA. (C) ( $r$  = Pearson correlation coefficient).



**SUPPLEMENTAL FIGURE 2.** Time-activity curves in whole brain grey matter for scan without (baseline) and with CsA infusion and fits obtained with the 1T2K model for one representative subject.



**SUPPLEMENTAL FIGURE 3.** Mean parametric  $K_1$ ,  $k_2$  and  $V_T$  images ( $n = 10$ ) in Talairach space for baseline scans without CsA infusion. The basal ganglia, a dopamine D<sub>2</sub> receptor rich structure containing the ROIs caudate nucleus and putamen, are marked with a red broken line.

# SUPPLEMENTAL TABLE 1

Descriptive Pharmacokinetic Parameters for Different Brain Regions for Scans without  
(Baseline) and with CsA Infusion

Brain region	Condition	C <sub>max</sub> (SUV)	T <sub>max</sub> (min)	C <sub>55 min</sub> (SUV)	AUC (SUV.min)	k <sub>E,brain</sub> (1/min)
Whole brain grey matter	Baseline	1.06 ± 0.19	7.0 ± 1.2	0.67 ± 0.10	47 ± 8	0.009 ± 0.001
	CsA	0.97 ± 0.26	10.5 ± 3.2*	0.75 ± 0.16*	47 ± 12	0.006 ± 0.002*
White matter	Baseline	0.61 ± 0.16	14.8 ± 6.4	0.50 ± 0.19	33 ± 5	0.005 ± 0.004
	CsA	0.62 ± 0.15	26.8 ± 12.3*	0.59 ± 0.12	32 ± 8	0.001 ± 0.002*
Frontal lobe mid gyrus	Baseline	1.07 ± 0.19	6.7 ± 1.6	0.67 ± 0.11	48 ± 8	0.009 ± 0.001
	CsA	0.99 ± 0.26	13.3 ± 5.3*	0.76 ± 0.16*	48 ± 11	0.006 ± 0.002*
Putamen	Baseline	1.21 ± 0.24	9.5 ± 2.5	0.83 ± 0.18	60 ± 11	0.010 ± 0.002
	CsA	1.15 ± 0.33	23.3 ± 18.5*	0.94 ± 0.24	57 ± 16	0.006 ± 0.003*
Caudate nucleus	Baseline	0.97 ± 0.17	11.6 ± 3.1	0.70 ± 0.16	46 ± 9	0.007 ± 0.001
	CsA	0.90 ± 0.19	21.3 ± 14.6	0.78 ± 0.17	45 ± 10	0.004 ± 0.002*
Occipital lobe medial part	Baseline	1.08 ± 0.18	6.3 ± 1.6	0.69 ± 0.10	49 ± 8	0.009 ± 0.001
	CsA	1.02 ± 0.27	9.3 ± 3.5*	0.78 ± 0.16*	49 ± 11	0.006 ± 0.002*
Cerebellum	Baseline	1.15 ± 0.21	4.4 ± 1.5	0.61 ± 0.09	47 ± 8	0.011 ± 0.001
	CsA	1.04 ± 0.30	6.5 ± 2.5*	0.69 ± 0.15*	47 ± 12	0.008 ± 0.002*
Pituitary gland	Baseline	4.06 ± 0.64	2.0 ± 0.9	2.20 ± 0.63	160 ± 270	0.009 ± 0.007
	CsA	3.61 ± 0.50	2.5 ± 1.5	2.33 ± 0.57	149 ± 28	0.007 ± 0.010

Values are reported as mean ± standard deviation

C<sub>max</sub>, maximum concentration; T<sub>max</sub>, time of C<sub>max</sub>; AUC, area under the time-activity curve; k<sub>E,brain</sub>, elimination slope for radioactivity washout from the brain

\* *P* < 0.05 for comparison of CsA scan with baseline scan using a two sided, paired t-test

## SUPPLEMENTAL TABLE 2

Outcome Parameters of the Reversible 1T2K Model in Different Brain Regions for Scans  
without (Baseline) and with CsA Infusion

Brain region	Condition	$K_1$ (mL/(cm <sup>3</sup> .min))	$k_2$ (1/min)	$V_T$ (mL/cm <sup>3</sup> )	$V_b$ (mL)	$V_T / f_P$
Whole brain grey matter	Baseline	0.092±0.014 (2±1)	0.044±0.005 (4±1)	2.1±0.3 (3±1)	0.043±0.008 (4±1)	10.4±2.7
	CsA	0.099±0.012 (2±0)	0.037±0.004 (4±1)*	2.7±0.4 (3±1)*	0.042±0.006 (4±1)	11.1±3.8
White matter	Baseline	0.027±0.004 (11±2) <sup>†</sup>	0.092±0.015 (21±3) <sup>†</sup>	0.3±0.0 (15±2) <sup>†</sup>	0.026±0.004 (8±1)	1.5±0.5 <sup>†</sup>
	CsA	0.051±0.005 (3±1)*	0.022±0.003 (8±3)* <sup>‡</sup>	2.4±0.4 (6±2)*	0.023±0.003 (6±2)*	9.7±3.4*
Frontal lobe mid gyrus	Baseline	0.087±0.010 (2±1)	0.043±0.005 (4±3)	2.0±0.3 (3±2)	0.032±0.004 (5±3)	10.1±3.1
	CsA	0.097±0.009 (3±1)*	0.036±0.005 (5±3)*	2.8±0.5 (3±2)*	0.032±0.004 (6±4)	11.3±3.3*
Putamen	Baseline	0.091±0.015 (2±1)	0.035±0.003 (5±2)	2.6±0.4 (4±2)	0.037±0.007 (3±1)	12.9±3.2
	CsA	0.106 ± 0.019 (3±1)*	0.029±0.005 (5±3)*	3.7±0.7 (5±3)*	0.031±0.005 (3±1)*	14.9±4.2*
Caudate nucleus	Baseline	0.069±0.009 (2±1)	0.032±0.005 (6±3) <sup>†</sup>	2.2±0.5 (4±2)	0.029±0.007 (3±1)	10.9±3.3
	CsA	0.079 ± 0.011 (2±1)*	0.026± 0.005 (5±2)*	3.1±0.5 (4±2)*	0.025±0.005 (3±1)	12.6±3.5*
Occipital lobe medial part	Baseline	0.089±0.009 (2±0)	0.043±0.004 (4±2)	2.1±0.3 (4±1)	0.052±0.007 (4±1)	10.4±3.3
	CsA	0.104±0.009 (3±1)*	0.037±0.005 (6±3)*	2.8±0.5 (4±2)*	0.040±0.012 (3±1)	11.6±4.1*
Cerebellum	Baseline	0.098±0.010 (2±1)	0.053±0.006 (4±1)	1.9±0.3 (3±1)	0.045±0.011 (4±1)	9.4±3.0
	CsA	0.116±0.016* (3±1)	0.047±0.008 (5±2)*	2.5±0.4 (4±2%)*	0.042±0.006 (5±1)*	10.3±3.7
Pituitary gland	Baseline	0.384±0.055 (5±2) <sup>†</sup>	0.056±0.014 (12±3) <sup>†</sup>	7.3±2.0 (9±2) <sup>†</sup>	0.144±0.056 (13±7)	37.3±15.2 <sup>†</sup>
	CsA	0.454±0.126 (7±3) <sup>‡</sup>	0.056±0.023 (15±6) <sup>‡</sup>	9.0±3.0 (11±5) <sup>‡</sup>	0.148±0.103 (12±6)	36.9±15.6 <sup>‡</sup>

Values are reported as mean ± standard deviation. The value in parentheses represents the precision of parameter estimates (expressed as their coefficient of variation in percent).

$K_1$  (mL/(cm<sup>3</sup>.min)), rate constant for radioactivity transfer from plasma into brain;  $k_2$  (1/min), rate constant for radioactivity transfer from brain into plasma;  $V_T$  (mL/cm<sup>3</sup>), total volume of distribution;  $V_T / f_P$ ,  $V_T$  over the free fraction of <sup>11</sup>C-metoclopramide in plasma;  $V_b$ , fractional arterial blood volume in the brain

\*  $P < 0.05$  for comparison of CsA scan with baseline scan using a two sided, paired t-test

<sup>†</sup>  $P < 0.05$  for comparison with whole brain grey matter region in baseline scans using one-way ANOVA followed by a Tukey's multiple comparison test

<sup>‡</sup>  $P < 0.05$  for comparison with whole brain grey matter region in CsA scans using one-way ANOVA followed by a Tukey's multiple comparison test



## **SUPPLEMENTAL REFERENCES**

1. Hamwi A, Salomon A, Steinbrugger R, Fritzer-Szekeres M, Jager W, Szekeres T. Cyclosporine metabolism in patients after kidney, bone marrow, heart-lung, and liver transplantation in the early and late posttransplant periods. *Am J Clin Pathol*. 2000;114:536-543.