1 Administration Routes for SSTR- / PSMA- and FAP-directed

- 2 Theranostic Radioligands in Mice
- 3 Running title: Biodistribution of ⁶⁸Ga-ligands
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ABSTRACT

- 27 Introduction: The NETTER-1, VISION, and TheraP trials prove efficacy of repeat intravenous (i.v.)
- application of small radioligands. Application by subcutaneous (s.c.), intraperitoneal (i.p.), or oral
- 29 (p.o.) access are important alternatives and may yield comparable or favorable organ and tumor
- 30 radioligand uptake. Here, we assess organ and tumor biodistribution for various radioligand
- 31 application routes in healthy mice and models of somatostatin receptor (SSTR)-, prostate-specific
- membrane antigen (PSMA)-, and fibroblast activation protein (FAP)- expressing cancer.
- 33 Methods: Healthy and tumor-bearing male C57BL/6 or NOD SCID Gamma mice, respectively,
- were applied with a mean of 6.0±0.5 MBq ⁶⁸Ga-DOTATOC (RM1-SSTR allograft), 5.3±0.3 MBq
- 35 ⁶⁸Ga-PSMA11 (RM1-PSMA allograft) or 4.8±0.2 MBq ⁶⁸Ga-FAPI46 (HT1080-FAP xenograft) i.v.,
- 36 i.p., s.c. or p.o.. *In vivo* positron emission tomography and *ex vivo* biodistribution in tumor, organs,
- and at the injection site were assessed up to 5h post injection (p.i.). Healthy mice were monitored
- for up to 7 days after the last scan for signs of stress or adverse reactions.
- 39 Results: After i.v., i.p. and s.c. radioligand administration, average residual activity at the injection
- site was <17%IA/g (1h p.i.), <10%IA/g (2h p.i.) and ≤4%IA/g (4h p.i.) for all radioligands. Following
- 41 oral administration ≥50%IA/g remained within the intestines until 4h p.i.. Biodistribution in organs
- of healthy mice was nearly equivalent following i.v., i.p., and s.c. application at 1h p.i. and all
- subsequent timepoints (≤1%IA/g for liver, blood and bone marrow; 11.2±1.4%IA/g for kidneys). In
- 44 models for SSTR-, PSMA- and FAP-expressing cancer, tumor uptake was higher or equivalent for
- 45 i.p./s.c. versus i.v. injection at 5h p.i. (ex vivo): SSTR: 7.2±1.0%IA/g (p=0.0197) / 6.5±1.3%IA/g
- 46 (p=0.0827) versus 2.9±0.3%IA/g; PSMA: 3.4±0.8%IA/g (p=0.9954) / 3.9±0.8%IA/g (p=0.8343)
- 47 versus 3.3±0.7%IA/g and FAP: 1.1±0.1%IA/g (p=0.9805) / 1.1±0.1%IA/g (p=0.7446) versus
- 48 1.0±0.2%IA/g.
- 49 Conclusion: In healthy mice, biodistribution of small theranostic ligands following i.p. or s.c.
- application is nearly equivalent compared to i.v. injection. S.c. administration resulted in highest
- 51 absolute SSTR tumor and tumor-to-organ uptake as compared to the i.v. route, warranting further
- 52 clinical assessment.

53 **Keywords:**

- radioligand, biodistribution, small animal PET, theranostic, intravenous, subcutaneous,
- 55 intraperitoneal, PSMA, SSTR, FAP, alternative application routes

INTRODUCTION

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NETTER-1 (1), and the more recent clinical trials TheraP (2), and VISION (3), establish somatostatin receptor (SSTR)- and prostate specific membrane antigen (PSMA)-directed small ligand radiotheranostics as efficacious cancer therapy with favorable safety profiles. Recently, fibroblast activation protein (FAP)-targeting small ligands have emerged for positron emission tomography (PET) and therapy of cancers (4). Intravenous (i.v.) application is the standard route for radioligand applications. However, oral (p.o.), intraperitoneal (i.p.), and subcutaneous (s.c.) administrations are faster and require a lower level of training when compared to i.v., both in the preclinical and clinical settings. The volume of preclinical and clinical radioligand applications is growing rapidly and thus, there is an urgent unmet need to assess alternative application routes to address the increasing demand. In addition, novel FAP-directed therapies are in a dynamic and evolving process, highlighting the emerging need for an optimization of administration routes of these novel radioligands for ongoing preclinical and clinical assessment. In this intent, assessment of biodistribution and administration routes of ⁶⁸Ga-FAPI46 (fibroblast activation protein inhibitor) was requested by the German Federal Institute for Drugs and Medical Devices (BfArM) for recent approval of a prospective clinical trial on ⁶⁸Ga-FAPI46 PET/CT for various types of cancer (NCT04571086).

We hypothesize that i.p. and s.c. application will yield near equivalent organ and tumor biodistribution compared to the routine i.v. injection. We further hypothesized that organ and tumor uptake will be significantly lower following p.o. application of radioligands. Here, we compare tumor and organ biodistribution following i.v., i.p., s.c and p.o. application of small radioligands in healthy mice and mouse models of SSTR-, PSMA-, and FAP-expressing cancer.

METHODS

Cell Culture

RM1 cells, virally stably transduced with SFG-Egfp/Luc (RM1-PGLS) or pMSCV-IRES-YFP II-hSSTR (RM1-SSTR) to express high levels of cell surface human PSMA or SSTR2 (*5*), were obtained from Johannes Czernin (University of California, Los Angeles). HT1080-FAP cells were a gift from Uwe Haberkorn (University Hospital of Heidelberg). HT1080 cells were stably transfected with the plasmid pcDNAI/neo-FAP (expressing the untagged full-length cDNA of human FAP) followed by neomycin selection (*6*). RM1-PSMA and RM1-SSTR were maintained in Roswell Park Memorial Institute 1640 medium (GIBCO) and HT1080-FAP in Dulbecco's Modified Eagle Medium (GIBCO), both with 10% fetal bovine serum (Thermo Fisher Scientific) and 0.5% penicillin/streptomycin (GIBCO), at 37°C with 5% CO₂. Cells were thawed 2 weeks or passaged 3 times before inoculation. Cells were routinely assessed for mycoplasma contamination using the VenorGeM OneStep kit (Minerva Biolabs).

Radiosynthesis

68Ga-DOTATOC, 68Ga-PSMA11 and 68Ga-FAPI46 were obtained from the radiopharmacy of our clinic. Clinical-grade radiolabeling of precursors (DOTATOC, PSMA11, FAPI46) was performed using the Modular-Lab eazy for DOTATOC and PSMA or Scintomics GRP 3V for FAPI using commercially available reagent kits. The final solution had <5μg/mL for 68Ga-DOTATOC, <3μg/mL for 68Ga-PSMA, and ~50μg/mL for 68Ga-FAPI with 100μl injected volume per mouse. Radiochemical purity was determined with radio-high-performance liquid chromatography. FAPI: Chromolith Performance RP18e column from Merck (100 x 3mm) gradient: 0-20% MeCN+0.1% TFA in 5 min run time 15 min; PSMA: 5-40% MeCN+0.1% TFA in 10 min run time 15 min; DOTATOC: 24% MeCN+0.1% TFA for 8 min, then 24-60% in 1 min, run time 15 min; and thin-layer chromatography (iTLC-SG, ammonium acetate (77g/L), methanol R (50:50 v/v)). The radiochemical purity exceeded 98% for all radioligands.

Mice and Tumor Models

Male C57BL/6 and NOD SCID Gamma mice were purchased from Charles River Laboratories (6-8 weeks old) and housed under specific pathogen-free conditions with food and

water available *ad libitum*. Health status monitoring of mice was performed by assessing a summarized score twice a week (healthy animals) or daily (tumor-bearing animals). The study was approved by the North Rhine-Westphalia State Agency for Nature, Environment and Consumer Protection (LANUV), Germany (permit number: AZ.81-02.04.2018.A090).

For subcutaneous tumors, mice were injected with 0.1×10^6 RM1-SSTR cells or RM1-PSMA (C57BL/6) or 1.0×10^6 HT1080-FAP (NOD SCID Gamma) in matrigel/PBS (50:50 ratio) into the shoulder region. Tumor volume (V) was calculated by measuring the length (L) and width (W) of tumors by caliper and using the formula $V = 1/2(L \times W^2)$ (T). PET scans were acquired 7-10 days after tumor inoculation, as described previously (5.8). Mean+SEM tumor volumes were 0.39 ± 0.09 cm³ (interquartile range 0.07-0.66cm³) for RM1-SSTR tumors, 0.05 ± 0.01 cm³ (interquartile range 0.02-0.08cm³) for RM1-PSMA tumors, and 0.22 ± 0.03 cm³ (interquartile range 0.06-0.25cm³) for HT1080-FAP tumors.

Radioligand Application and Small Animal Positron Emission Tomography/Computed Tomography (PET/CT)

Healthy or tumor-bearing anesthetized mice (1.5-2% isoflurane) received (mean±SEM) 6.0 ± 0.5 MBq 68 Ga-DOTATOC, 5.3 ± 0.3 MBq 68 Ga-PSMA11 or 4.8 ± 0.2 MBq 68 Ga-FAPI46 i.v. (tail vein), i.p., s.c. or p.o. (p.o. HT1080-FAP tumor-bearing mice only) (differences between injected activities, p=n.s.). Each healthy mouse received i.v., i.p., s.c. and p.o. administration with 1 week interval between PET/CT scans (Supplemental Figure 1A). Each tumor-bearing mouse was scanned twice, at 1h and 4h p.i., following either i.v., i.p., or s.c. application and was sacrificed ~5h p.i for *ex vivo* analysis (Supplemental Figure 1B). Imaging was performed with a β -CUBE (PET) and X-CUBE (CT) (Molecubes) in temperature-controlled beds with monitoring of breathing frequency. PET/CT was acquired (PET, 15 minutes; CT, 5 minutes) in list mode with frames for 5, 10 and 15 minutes (dynamic scans, maximum delay between injection and scan start 5 minutes) and static scans 1h, 2h and 4h p.i. for healthy mice and 1h and 4h p.i. for tumor groups.

Image Reconstruction and Processing

Images were reconstructed using an iterative reconstruction algorithm (ISRA, 30 iterations) with attenuation correction of the corresponding CT image. PET data were reconstructed into a 192x192 transverse matrix, producing a 400 µm isometric voxel size. PET images were evaluated with PMOD software (PMOD Technologies LLC). Decay-corrected mean percent injected activity

per gram (%IA/g) of the tumor and organs of interest was derived from DICOM images. Volumes of interest (VOIs) were defined as spheres of 5 mm (lung, liver, spleen, intestines, heart, brain, kidneys) and 2.5 mm (bone marrow, thigh muscle, blood pool, injection site, tumor) diameter in tissues of interest. %IA/g was calculated from the average pixel values reported in Bq/mL within these VOIs corrected for radioactive decay and mouse body weight.

Ex Vivo Analysis

Approximately 5h p.i., animals were sacrificed and organs of interest were extracted, dabbed dry, weighed, and radioactivity was measured in an automated gamma counter (Perkin-Elmer Gamma Counter 2480 Wizard²). Organ and tumor uptake was calculated from radioactive counts, decay-corrected and expressed as %IA/g.

Data and Statistical Analysis

Data are presented as mean±SEM unless indicated otherwise. All statistical analyses were performed using GraphPad Prism (version 9.1.0; GraphPad Software). Tumor-to-organ uptake ratios were calculated for blood, kidney, liver and bone marrow (femur) using %IA/g at 1h and 4h in *in vivo* VOIs and at 5h for *ex vivo* gamma counter measurements (%IA/g tumor / %IA/g organ). Statistical significance was assessed using Brown-Forsythe and Welch ANOVA test with Dunnett's T3 multiple comparisons test or Tukey's multiple comparisons test. p-values below 0.05 were considered statistically significant. Statistically significant data are indicated by asterisks (*p<0.05; **p<0.01; ***p<0.001; ****p<0.0001).

RESULTS

Local and Systemic Activity

To assess biodistribution of radioligands applied via different routes, we measured the activity retained at the injection site versus the overall systemic activity distribution excluding the application site. Activity at the injection site decreased over time following i.v., i.p. and s.c. administration in healthy mice (Figure 1, Supplemental Figure 1). Residual activity at the injection site 4h p.i. was (mean±SEM) for i.v.: 1.0±0.3%IA/g, i.p.: 4.4±2.1%IA/g and s.c.: 2.1±0.5%IA/g) for all radioligands; this correlated inversely with increased systemic availability of radioligands. Oral administration resulted in significant and prolonged retention of radioligands in the stomach and proximal small bowel as well as a low systemic distribution (Figure 1A-C). Following p.o. administration, average systemic uptake was highest for ⁶⁸Ga-FAPI46 (Figure 1C). Therefore, p.o. application was further explored in HT1080-FAP tumor-bearing mice.

Near Equivalent Organ Biodistribution of Radioligands Following i.p., s.c., and i.v. Application in Healthy Mice

In healthy mice, i.p., s.c., and i.v. injection of radioligands resulted in near equivalent organ biodistribution *in vivo* (Figure 2-4, Supplemental Figures 2-3). Radioligand retention in blood and kidney is listed in Supplemental Table 1. Blood retention in healthy mice was significantly higher following i.p. or s.c. versus i.v. application of ⁶⁸Ga-PSMA11: i.p., 1h p=0.0226, 2h p=0.0463, and 4h p=0.0394; s.c., 1h p=0.0880, 2h p=0.0021, and 4h p=0.065. For ⁶⁸Ga-DOTATOC and ⁶⁸Ga-FAPI46, blood and kidney distribution after i.p. and s.c. application were comparable to those following i.v. injection (Figure 2-4). In further organs, including liver, bone marrow, lung, heart spleen, intestines, brain and muscle, i.p., s.c., and i.v. application routes exhibited comparable physiological biodistribution (Supplemental Figure 2). Moreover, in healthy mice, no short-term and longer-term adverse effects of radioligand application and PET/CT procedures were noted during the study duration (5 weeks).

Increased or Comparable Tumor Uptake Following i.p. or s.c. Versus i.v. Injection of Radioligands

To evaluate the impact of the application route on tumor uptake of ⁶⁸Ga-DOTATOC, ⁶⁸Ga-PSMA11, or ⁶⁸Ga-FAPI46, we assessed *in vivo* and *ex vivo* tumor and organ uptake in SSTR-, PSMA- and FAP-expressing tumor models (Table 1, Figure 5-7, Supplemental Figures 4-5).

In mice bearing SSTR tumors, i.p./s.c. application resulted in significantly higher tumor uptake (mean±SEM) when compared to i.v.: p=0.0124 / p=0.0377 at 1h; p=0.0301 / p=0.0411 at 4h; and p=0.0197 / p=0.0827 at 5h (*ex vivo*) (Table 1; Supplemental Figure 4). Tumor uptake of ⁶⁸Ga-PSMA11 or ⁶⁸Ga-FAPI46 following i.p./s.c. injection of mice bearing PSMA- or FAP-expressing tumors was comparable to the uptake observed after i.v. injection (Table 1).

Oral administration in mice bearing FAP-expressing tumors did not result in notable tumor uptake (Table 1, Supplemental Figure 4). Oral application of ⁶⁸Ga-FAPI46 in tumor-bearing mice yielded comparable biodistribution characteristics as seen in healthy mice (Supplemental Figure 4) with high gastrointestinal retention of the radioligand and low systemic distribution.

Tumor-to-organ uptake ratios of organs relevant for dosimetry (*9,10*) for i.p./s.c. versus i.v. application are depicted in Figure 5-7. l.p./s.c. application resulted in higher or equivalent tumor-to-liver ratios at 5h p.i. when compared to i.v. (5h p.i. mean ratio±SEM): (i) ⁶⁸Ga-DOTATOC: 27.4±2.2-fold (p=0.0138) / 25.3±5.6-fold (p=0.2756) versus 13.9±2.9-fold; (ii) ⁶⁸Ga-PSMA11: 28.2±7.4-fold (p=0.4504) / 39.4±5.7-fold (p=0.0259) versus 16.9±2.8-fold; and (iii) ⁶⁸Ga-FAPI46: 6.1±1.6-fold (p=0.4198) / 12.0±1.1-fold (p=0.0005) versus 3.7±0.4-fold (Figure 5-7). Tumor-to-bone marrow ratios were higher for i.p. compared with i.v. application in mice bearing SSTR-expressing tumors: 50.7±4.3 versus 25.7±4.9 (p=0.0096) (Figure 5). S.c. application resulted in higher tumor-to-blood ratios when compared with i.v. application in mice bearing PSMA-expressing tumors: 24.5±4.2-fold versus 6.0±0.9-fold (p=0.0186). For other tumor-to-organ uptake ratios no significant difference was observed (Figure 6). Oral application of ⁶⁸Ga-FAPI46 resulted in negligible uptake in organs and tumors (Table 1, Supplemental Figure 4).

DISCUSSION

The current delivery method for radioligands for nuclear imaging or therapy is i.v. injection. However, comparing different application routes is important for the translation of novel FAP ligands and optimization of current clinical protocols for PSMA or SSTR ligands.

The current study aimed at comparing the biodistribution of SSTR-, PSMA-, and FAP-directed small radioligands administered i.p., s.c., or p.o. with the standard i.v. application. Alternative application routes may alter systemic distribution and tumor uptake (11-13), for instance by slowing absorption due to a reduced rate of molecular transport via the lymphatics and blood flow to the organs of interest/tumor (14).

Administration of small radioligands i.v., i.p., and s.c. was feasible and well tolerated as assessed by a scoring system including behavior and overall physical appearance of mice. Small radioligand systemic availability and biodistribution was comparable for i.p. and s.c. versus i.v. application (Figure 1-4). In addition, i.p. and s.c. administration in mice resulted in significantly higher ⁶⁸Ga-DOTATOC tumor uptake (Table 1), tumor-to-liver and tumor-to-bone marrow ratio in SSTR-expressing tumors when compared with i.v. injection (Figure 5).

These findings have implications for preclinical and clinical radioligand administration, since they could offer advantages for both fields. In mice, i.v. injection requires highly trained personnel, and is more error-prone (e.g., paravenous injection) and time consuming. I.p. and s.c. administration may serve as simple alternative application routes for imaging at later timepoints after injection or therapy, allowing a higher throughput in mouse studies, with lower dropout rates and high reproducibility. In mice, i.p. administration did not compromise radioligand tumor accumulation despite a high initial absorbed dose in the intestines (15). However, due to slower systemic bioavailability following i.p. or s.c. injection, i.v. application is recommended for early dynamic imaging.

In clinical routine, usage of alternative application routes to i.v. may improve outpatient care and benefit potential new therapy schemes allowing repeat radioligand application at short interval.

In patients, i.p. application is limited due to a higher likelihood of infection or abdominal organ damage. However, s.c. application is already well established as a standard route for outpatient injectable medications and has an emerging role in delivery of biotherapeutics or monoclonal antibodies (16,17). Indeed, in patients with accidental paravenous infusion of ¹⁷⁷Lu-DOTATOC absorption from the paravenous injection site occurs with a half-life of less than 4h (Supplemental Figure 6); this is in line with a short drainage observed following s.c. injection in mice. We therefore expect that s.c. application in patients would be feasible.

Still, an increased radiation dose to organs such as kidneys, bone marrow, blood, lungs or liver, may limit benefit from i.p./s.c. injection. However, if radioligand therapy regimens would be changed to a weekly or biweekly schedule using s.c. application, activities for each administration could probably be reduced in favor of these more frequent treatments. Weekly or biweekly i.p./s.c. application could be realized by outpatient care, reducing the patient's time in hospital, personnel capacities and thus, reduced costs.

In this study, the uptake in non-target tissues did not exceed critical values or radiation dose as suggested from measured uptake in %IA/g (Figure 2-4). Therefore, we assume that a detrimental radiation burden to organs at risk (mainly kidneys) after s.c. and i.p. application when compared to the standard i.v. route is unlikely. Notably, preclinical and clinical studies for DOTATOC- and PSMA-targeting radiotherapies demonstrated that after i.v. absorbed doses in organs of risk are not likely to cause relevant radiotoxicity (9,10,18,19). However, to precisely estimate the additional absorbed dose to the adjacent tissue (by i.p. and s.c.) following radioligand therapy, further studies with Lu-177 labeled ligands and quantitative preclinical SPECT imaging should be performed. Yet, if we assume a half-life of 2.3 hours for the change in local activity over time at the injection site, as recently published by Tylski (20), we would not expect to detect a change in dosimetry between one Lu-177 administration and, e.g., 2-3 administrations spaced by 48 h.

To date, the entire theranostics routine is based on rather conservative application schemes with few possibilities of patient-specific modification. Our observation that s.c. application showed similar tumor uptake as compared to i.v. may open up new opportunities for alternative application schemes in the clinical routine – e.g., weekly or biweekly applications, which are less feasible if using repeat i.v. injections. Also, s.c. application is faster and easier than i.v., and could thus be realized in outpatient care by medical laboratory assistants in a time-efficient manner for both, patient and clinic personnel. Furthermore, it would be interesting to investigate the influence of i.v. application rate (applied dose per time) on tumor uptake. This could be realized in a clinical study or observational trial on patients with poor vein status.

This study has some limitations. This study assessed ⁶⁸Ga-ligands for PET imaging and did not examine therapeutic ¹⁷⁷Lu-labelled ligands. Furthermore, in-bed injection with concurrent dynamic PET acquisition was not performed and due to the short ⁶⁸Ga half-life, timepoints beyond 5h p.i. were not feasible.

CONCLUSION

In mice, PET imaging after i.v., i.p., or s.c. injection of SSTR-. PSMA-, or FAP-directed small radioligands is feasible. I.p. and s.c. administration of SSTR-ligands resulted in higher absolute tumor and relative tumor-to-organ uptake compared to i.v., which may translate into improved tumor irradiation in the setting of radioligand therapies and warrants further translational assessment.

DISCLOSURE

WPF was a consultant for Janssen and Calyx, and he received fees from Bayer and Parexel outside of the submitted work. KH reports personal fees from Bayer, personal fees and other from Sofie Biosciences, personal fees from SIRTEX, non-financial support from ABX, personal fees from Adacap, personal fees from Curium, personal fees from Endocyte, grants and personal fees from BTG, personal fees from IPSEN, personal fees from Siemens Healthineers, personal fees from GE Healthcare, personal fees from Amgen, personal fees from Novartis, personal fees from ymabs, personal fees from Aktis Oncology, personal fees from Theragnostics, personal fees from Pharma15, outside the submitted work. KL reports paid consulting activities for Sofie Biosciences/iTheranostics, and funding from AMGEN outside of the submitted work. No other potential conflict of interest relevant to this article was reported.

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KEY POINTS

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QUESTION: Are there alternatives to intravenous injection of SSTR-, PSMA-, or FAP-directed 297 298 radioligands? 299 PERTINENT FINDINGS: In healthy mice, i.p. and s.c. application of small radiotheranostic ligands resulted in near equivalent systemic availability and organ biodistribution at early (1h) and late 300 (4h) timepoints p.i. when compared to i.v. injection. I.p./s.c. administration significantly increased 301 absolute tumor and relative tumor-to-organ uptake in SSTR tumors (68Ga-DOTATOC) compared 302 303 to i.v. route. 304 IMPLICATIONS FOR PATIENT CARE: I.p. and s.c. application is feasible in animal models of 305 small radioligand imaging or therapy. Tumor uptake and tolerability of s.c. application warrants 306 assessment in clinical studies.

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Table 1. I.p. or s.c. application led to higher or equivalent tumor uptake compared to i.v injection. Mice with subcutaneous RM1-SSTR, RM1-PSMA or HT1080-FAP tumors were injected with 68 Ga-DOTATOC, 68 Ga-PSMA11 or 68 Ga-FAPI46. Absolute tumor uptake (%IA/g) at 1h and 4h p.i (*in vivo* PET), and 5h p.i. (*ex vivo* gamma counter) is given. Data are represented as mean %IA/g±SEM of n=6 mice/group. *p<0.05; **p<0.01.

RM1-SSTR (68Ga-DOTATOC)										
	i.v.	i.p.	s.c.	p-valı i.v. vs.	ue .	p-value i.v. vs. s.c.				
in vivo 1 h	5.3 ± 0.6	9.9 ± 1.0	10.8 ± 1.6	p=0.0124*		p=0.0377*				
in vivo 4 h	4.4 ± 0.7	8.6 ± 1.1	11.1 ± 2.0	p=0.0301*		p=0.0411*				
ex vivo 5 h	2.9 ± 0.3	7.2 ± 1.1	6.5 ± 1.3	p=0.0197*		p=0.0827				
	RM1-PSMA (68Ga-PSMA11)									
	i.v.	i.p.	s.c.	p-value i.v. vs. i.p.		p-value i.v. vs. s.c.				
in vivo 1 h	2.9 ± 0.2	3.0 ± 0.6	2.6 ± 0.4	p=0.9837		p=0.8297				
in vivo 4 h	2.6 ± 0.2	2.6 ± 0.7	2.9 ± 0.5	p=0.9996		p=0.8289				
ex vivo 5 h	3.3 ± 0.7	3.4 ± 0.8	3.9 ± 0.8	p=0.9954		p=0.8343				
	HT1080-FAP (⁶⁸ Ga-FAPI46)									
	i.v.	i.p.	s.c.	p.o.	p-value i.v. vs. s.c	p-value i.v. vs. s.c	p-value i.v. vs. p.o.			
in vivo 1 h	1.2 ± 0.2	2.0 ± 0.4	2.2 ± 1.1	0.1±0.03	p=0.3024	p=0.6732	p=0.0032**			
in vivo 4 h	1.0 ± 0.2	1.5 ± 0.3	1.1 ± 0.6	0.1±0.04	p=0.4559	p=0.9911	p=0.0087**			
ex vivo 5 h	1.0 ± 0.2	1.1 ± 0.1	1.4 ± 0.4	0.02±0.01	p=0.9805	p=0.7446	p=0.0058**			

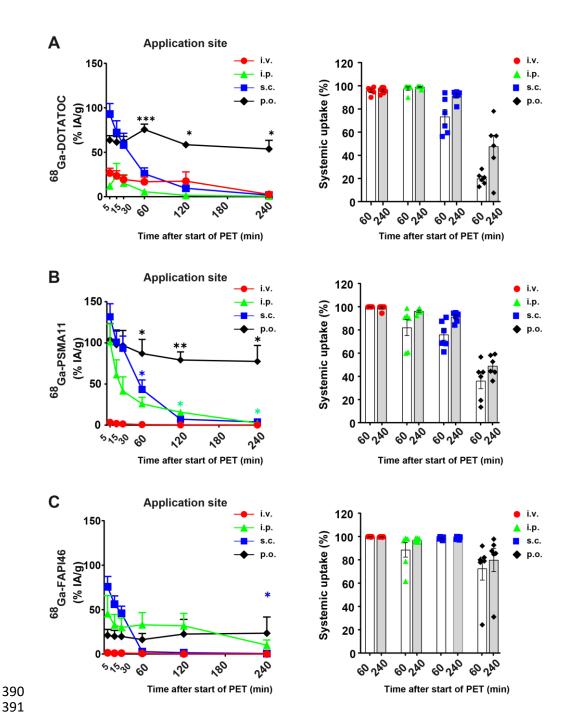


Figure 1. Activity at the application site and systemic availability over time in healthy mice. Retention of (A) ⁶⁸Ga-DOTATOC, (B) ⁶⁸Ga-PSMA11, and (C) ⁶⁸Ga-FAPI46 in healthy mice (n=6/group) at the application site. *Left panel*: Time activity curves illustrate radioligand dynamics at the application site for i.v., i.p., s.c., and p.o. application. *Right panel*: Relative systemic uptake of whole body VOI excluding application site VOI displayed in % of total body uptake. Each dot represents a mouse. Data are shown as mean+SEM. %IA/g: percent of the injected activity per gram. Asterisks indicate significance compared to i.v. application. *p<0.05; **p<0.01; ***p<0.001.



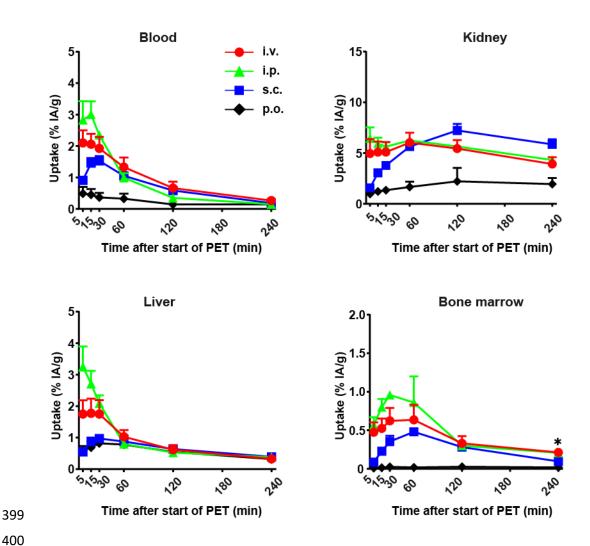


Figure 2. In healthy mice, organ biodistribution at ≥1h p.i following i.p. and s.c. radioligand application is nearly equivalent to i.v. injection. Healthy mice (*n*=6/group) underwent PET scans following i.v., i.p., s.c., and p.o. radioligand application, respectively, at minute 0-30 after start of PET and after 1h, 2h and 4h with subsequent sacrifice of animals. Time-activity curves illustrate *in vivo* PET biodistribution of ⁶⁸Ga-DOTATOC dynamics in VOIs at indicated times for i.v., i.p., s.c., and p.o. application. Data are shown as mean+SEM. %IA/g: percent injected activity per gram. Asterisks indicate significance compared to i.v. injection.*p<0.05.



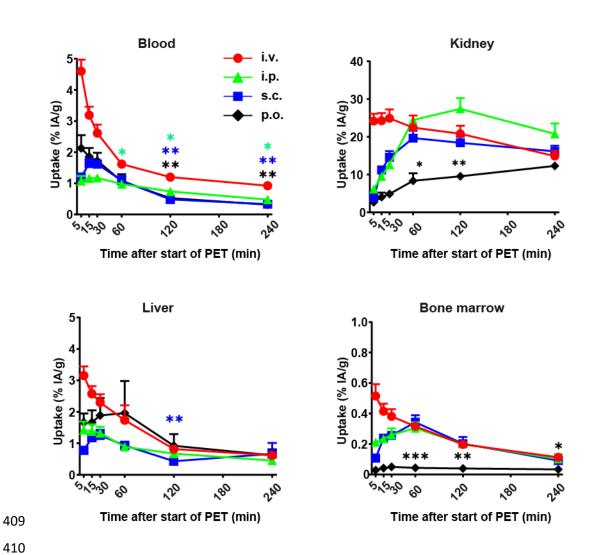


Figure 3. In healthy mice, organ biodistribution at ≥1h p.i following i.p. and s.c. radioligand application is nearly equivalent to i.v. injection. Healthy mice (*n*=6/group) underwent PET scans following i.v., i.p., s.c., and p.o. radioligand application, respectively, at minute 0-30 after start of PET and after 1h, 2h and 4h with subsequent sacrifice of animals. Time-activity curves illustrate *in vivo* PET biodistribution of ⁶⁸Ga-PSMA dynamics in VOIs at indicated times for i.v., i.p., s.c., and p.o. application. Data are shown as mean+SEM. %IA/g: percent injected activity per gram. Asterisks indicate significance compared to i.v. injection.*p<0.05; **p<0.01; ***p<0.001.



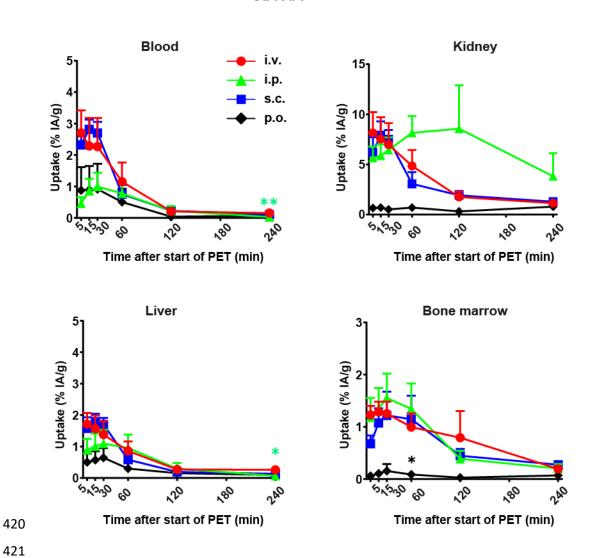


Figure 4. In healthy mice, organ biodistribution at ≥1h p. Healthy mice (*n*=6/group) underwent PET scans following i.v., i.p., s.c., and p.o. radioligand application, respectively, at minute 0-30 after start of PET and after 1h, 2h and 4h with subsequent sacrifice of animals. Time-activity curves illustrate *in vivo* PET biodistribution of ⁶⁸Ga-FAPI dynamics in VOIs at indicated times for i.v., i.p., s.c., and p.o. application. Data are shown as mean+SEM. %IA/g: percent injected activity per gram. Asterisks indicate significance compared to i.v. injection.*p<0.05; **p<0.01.

Ga-DOTATOC

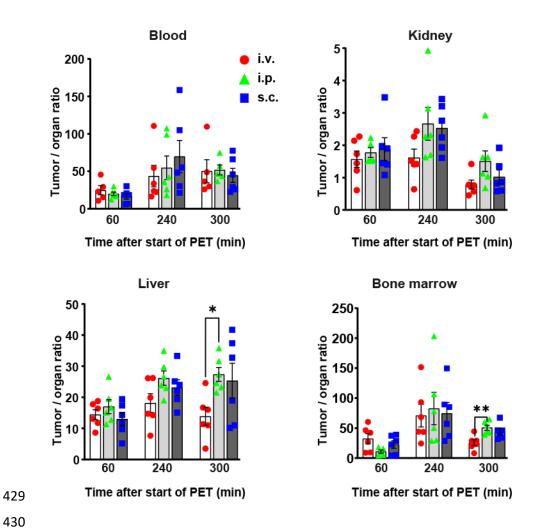


Figure 5. I.p. and s.c. radioligand application increase tumor-to-liver uptake compared to i.v. injection. Mice with subcutaneous RM1-SSTR tumors (*n*=6/group) with i.v., i.p. and s.c. application of ⁶⁸Ga-DOTATOC and PET scans after 1h and 4h, followed by sacrifice (5h) and subsequent assessment of radioactivity in organs and tumors by gamma counter. Plots show tumor-to-organ ratios after i.v., i.p. and s.c. of ⁶⁸Ga-DOTATOC. Each dot represents a mouse. Data are shown as mean±SEM. Asterisks indicate significance compared to i.v. injection. *p<0.05; **p<0.01.

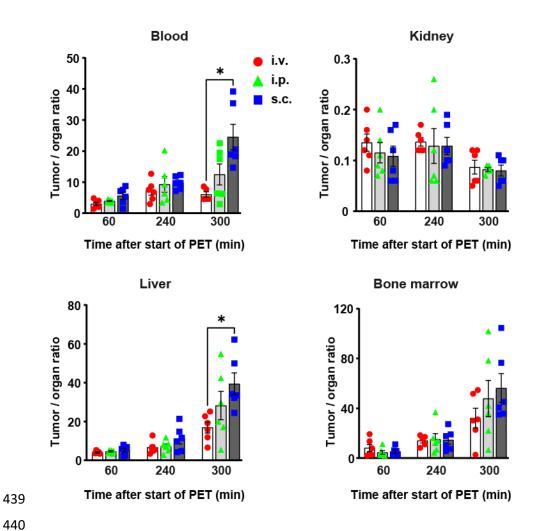


Figure 6. I.p. and s.c. radioligand application increase tumor-to-liver uptake compared to i.v. injection. Mice with subcutaneous RM1-PSMA tumors (*n*=6/group) with i.v., i.p. and s.c. application of ⁶⁸Ga-PSMA and PET scans after 1h and 4h, followed by sacrifice (5h) and subsequent assessment of radioactivity in organs and tumors by gamma counter. Plots show tumor-to-organ ratios after i.v., i.p. and s.c. of ⁶⁸Ga-DOTATOC. Each dot represents a mouse. Data are shown as mean±SEM. Asterisks indicate significance compared to i.v. injection. *p<0.05.

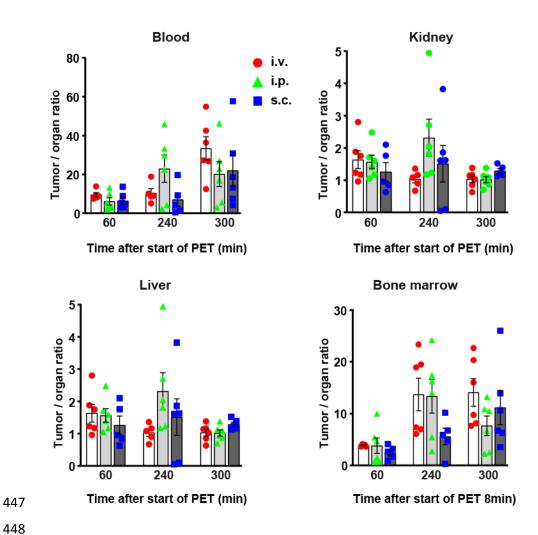
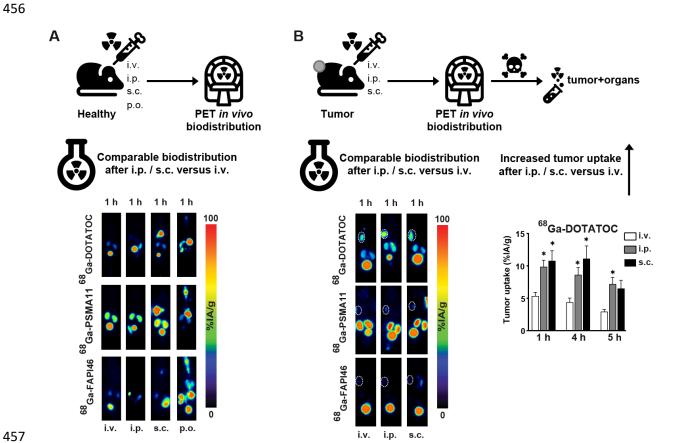


Figure 7. I.p. and s.c. radioligand application increase tumor-to-liver uptake compared to i.v. injection. Mice with subcutaneous HT-1080 tumors (*n*=6/group) with i.v., i.p. and s.c. application of ⁶⁸Ga-DOTATOC and PET scans after 1h and 4h, followed by sacrifice (5h) and subsequent assessment of radioactivity in organs and tumors by gamma counter. Plots show tumor-to-organ ratios after i.v., i.p. and s.c. of ⁶⁸Ga-DOTATOC. Each dot represents a mouse. Data are shown as mean±SEM.

GRAPHICAL ABSTRACT



1 Supplement

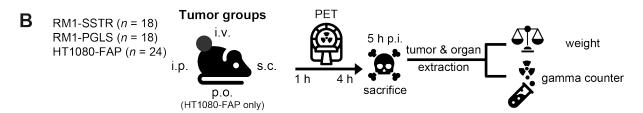
2 Supplemental Tables

Supplemental Table 1. I.v., i.p., and s.c. injection led to near equivalent organ biodistribution of radioligands in healthy mice. Healthy mice were injected with 68Ga-DOTATOC, 68Ga-PSMA11, or 68Ga-FAPI46, respectively. Absolute organ uptake is given as %IA/g at 1h and 4h p.i. (*in vivo* PET), and 5h p.i. (*ex vivo* gamma counter). Data represent mean %IA/g±SEM of *n*=6 mice/group. *p<0.05; **p<0.01.

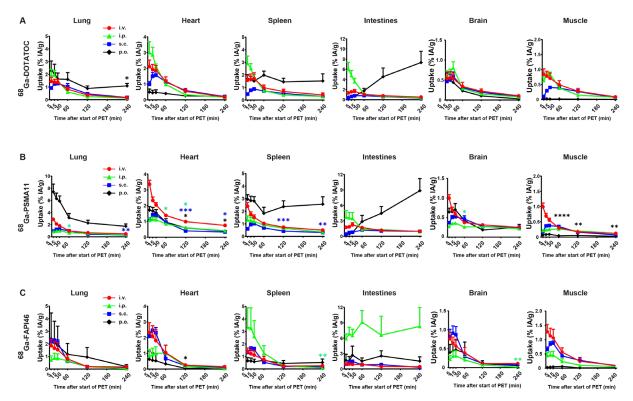
	⁶⁸ Ga-DOTATOC							
	i.v.	i.p.	s.c.	p-value i.v. vs. i.p.	p-value i.v. vs. s.c.			
blood 1h	1.3 ± 0.3	1.0 ± 0.1	1.1 ± 0.1	0.7589	0.8431			
blood 4h	0.3 ± 0.1	0.2 ± 0.02	0.2 ± 0.04	0.5639	0.8229			
kidneys 1h	6.0 ± 1.0	6.2 ± 0.4	5.6 ± 0.5	0.9965	0.9837			
kidneys 4h	3.9 ± 0.7	4.3 ± 0.2	5.9 ± 0.5	0.9393	0.1658			
	⁶⁸ Ga-PSMA11							
	i.v.	i.p.	s.c.	p-value i.v. vs. i.p.	p-value i.v. vs. s.c.			
blood 1h	1.6 ± 0.1	1.0 ± 0.1	1.1 ± 0.1	p=0.0226*	p=0.0880*			
blood 4h	0.9 ± 0.1	0.5 ± 0.1	0.3 ± 0.04	p=0.0394*	p=0.0065**			
kidneys 1h	22.4 ± 3.2	24.4 ± 0.9	19.7 ± 2.0	p=0.9279	p=0.8773			
kidneys 4h	14.9 ± 1.4	20.8 ± 2.8	16.1 ± 1.5	p=0.3040	p=0.9251			
	⁶⁸ Ga-FAPI46							
	i.v.	i.p.	s.c.	p-value i.v. vs. i.p.	p-value i.v. vs. s.c.			
blood 1h	1.2 ± 0.6	0.8 ± 0.4	0.8 ± 0.3	p=0.9505	p=0.9345			
blood 4h	0.2 ± 0.02	0.02 ± 0.01	0.1 ± 0.03	p=0.0036**	p=0.5326			
kidneys 1h	4.9 ± 1.6	8.7 ± 1.7	3.1 ± 1.2	p=0.5065	p=0.8014			
kidneys 4h	1.1 ± 0.2	3.8 ± 2.3	1.3 ± 0.2	p=0.6694	p=0.9513			

Supplemental Figures

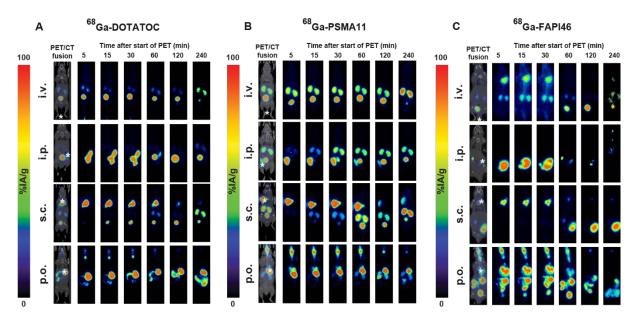
Healthy groups $\begin{array}{c}
68 \text{Ga-DOTATOC } (n = 24) \\
68 \text{Ga-PSMA } (n = 24) \\
68 \text{Ga-FAPI } (n = 24)
\end{array}$ i.v. $\begin{array}{c}
0-30 \\
\text{min}
\end{array}$ sacrifice



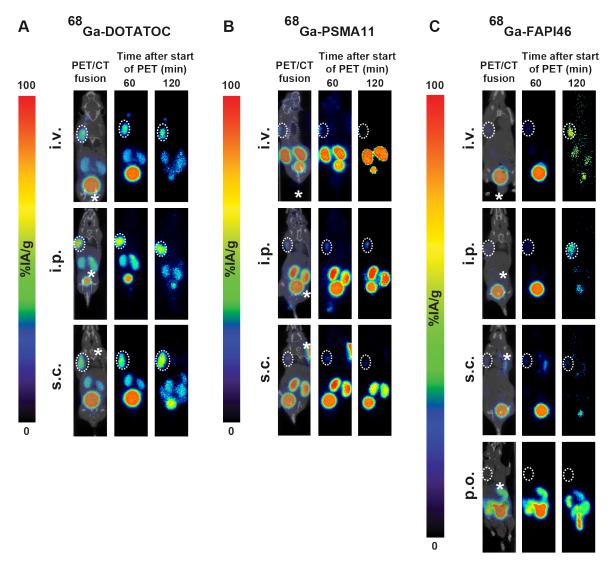
Supplemental Figure 1. Experimental design. (A) Healthy mice (*n*=6/group) underwent PET scans following i.v., i.p., s.c., and p.o. radioligand application, respectively, at minute 0-30 after start of PET and after 1h, 2h and 4h with subsequent sacrifice of animals. (B) Mice with subcutaneous RM1-SSTR, RM1-PSMA, or HT1080-FAP tumors (*n*=6/group) with i.v., i.p. and s.c. application of ⁶⁸Ga-DOTATOC, ⁶⁸Ga-PSMA11, or (C) ⁶⁸Ga-FAPI46, respectively underwent PET scans after 1h and 4h, followed by sacrifice (5h) and subsequent assessment of radioactivity in organs and tumors by gamma counter.



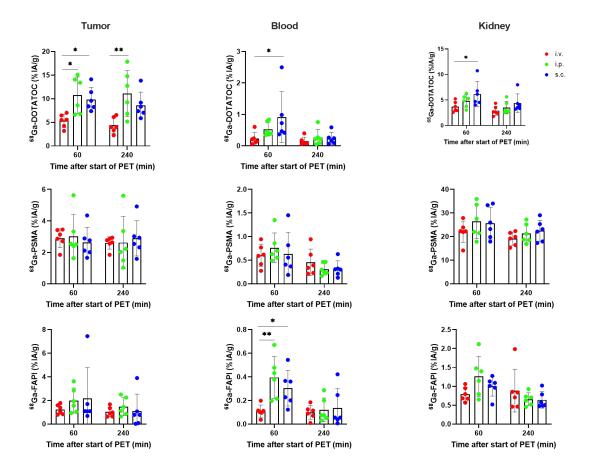
Supplemental Figure 2. Near equivalent radioligand organ biodistribution for i.p. and s.c. compared to i.v. injection in healthy mice. *In vivo* PET biodistribution of ⁶⁸Ga-ligands in healthy mice (*n*=6/group). PET scans with (A) ⁶⁸Ga-DOTATOC, (B) ⁶⁸Ga-PSMA11, and (C) ⁶⁸Ga-FAPI46. Time-activity curves illustrate radioligand dynamics in selected organ VOIs at indicated time points for i.v., i.p., s.c., and p.o. application. Data are shown as mean+SEM. %IA/g: percent of the injected activity per gram. Asterisks indicate significance compared to i.v. application route. *p<0.05; **p<0.01; ***p<0.001; ****p<0.0001.



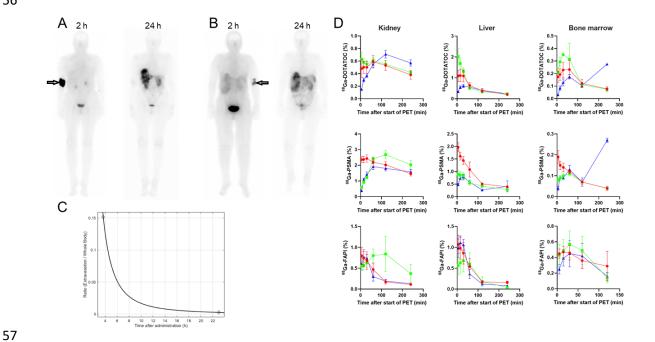
Supplemental Figure 3. PET biodistribution of 68 Ga-ligands in healthy mice. Whole body maximum intensity projections of one representative mouse out of n=6/group for each application route after injection of 68 Ga-labelled ligands. (A) 68 Ga-DOTATOC, (B) 68 Ga-PSMA11, and (C) 68 Ga-FAPI46 after i.v., i.p., s.c., and p.o. application in healthy mice. Asterisks indicate the injection site.



Supplemental Figure 4. PET biodistribution of 68 Ga-ligands in tumor-bearing mice. Whole body maximum intensity projections of one representative mouse out of n=6/group for each application route 1h and 4h after injection of 68 Ga-labelled ligands. (A) 68 Ga-DOTATOC, (B) 68 Ga-PSMA11, and (C) 68 Ga-FAPI46 via i.v., i.p., and s.c. application in RM1-SSTR-, RM1-PSMA-, or HT1080-FAP-tumor-bearing mice with additional p.o. application, respectively. Asterisks indicate the injection site; dashed circles indicate subcutaneous tumor in the right shoulder region.



Supplemental Figure 5. PET biodistribution of ⁶⁸Ga-ligands in tumor-bearing mice. *In vivo* PET uptake of ⁶⁸Ga-ligands in tumor-bearing mice (*n*=6/group). after ⁶⁸Ga-DOTATOC, ⁶⁸Ga-PSMA11, and ⁶⁸Ga-FAPI46. Bars illustrate radioligand uptake in selected organ VOIs at indicated time points for i.v., i.p., and s.c. application. Data are shown as mean±SD. %IA/g: percent of the injected activity per gram. Asterisks indicate significance compared to i.v. application route. *p<0.05; **p<0.01.



Supplemental Figure 6. Anterior whole-body planar images of ¹⁷⁷Lu-DOTATOC distribution 2h and 24h after paravenous infusion (arrow) of the radioligand in two patients (A and B). Radioligand absorption in patient A occurs with a half-life of 3.3 hours (C). Extrapolation from mice to humans suggests comparable biodistribution of ⁶⁸Ga-radioligands in healthy organs (D). Data are shown as mean±SEM.