

**Repurposing  $^{99m}\text{Tc}$ -mebrofenin as a probe for molecular imaging of hepatocyte transporters**

Solène MARIE<sup>1,2,3</sup>, Irene HERNÁNDEZ-LOZANO<sup>4</sup>, Oliver LANGER<sup>4</sup>, Nicolas TOURNIER<sup>1</sup>

<sup>1</sup> Université Paris-Saclay, CEA, CNRS, Inserm, Laboratoire d'Imagerie Biomédicale Multimodale, BIOMAPS, Service Hospitalier Frédéric Joliot, 4 place du Général Leclerc, 91401, ORSAY France.

<sup>2</sup> Université Paris-Saclay, Faculté de Pharmacie, 92296, Châtenay-Malabry, France

<sup>3</sup> AP-HP, Université Paris-Saclay, Hôpital Bicêtre, Pharmacie Clinique, 94270, Le Kremlin Bicêtre, France

<sup>4</sup> Department of Clinical Pharmacology, Medical University of Vienna, Vienna, Austria

**Short running title:** Molecular imaging of liver transporters

**Category:** Focus on molecular imaging

**Corresponding author:** Solène MARIE

Tel: +33.(0)1.69.86.77.12      Fax: +33.(0)1.69.86.77.86

CEA/SHFJ, 4 place du Général Leclerc 91400 ORSAY, France

solene.marie@aphp.fr

**First author:** Solène MARIE (PhD student), solene.marie@aphp.fr

Word count: 3,996

## **ABSTRACT**

Hepatocyte transporters control the hepatobiliary elimination of many drugs, metabolites and endogenous substances. Hepatocyte transporter function is altered in several pathophysiological situations and can be modulated by certain drugs, with a potential impact for pharmacokinetics and drug-induced liver injury. Development of substrate probes with optimal properties for selective and quantitative imaging of hepatic transporters remains a challenge.  $^{99m}\text{Tc}$ -mebrofenin has been used for decades for hepatobiliary scintigraphy, but the specific transporters controlling its liver kinetics have not been characterized until recently. These include sinusoidal influx transporters (organic anion-transporting polypeptides, OATP) responsible for hepatic uptake of  $^{99m}\text{Tc}$ -mebrofenin, and efflux transporters (multidrug resistance-associated proteins, MRP) mediating its canalicular (liver-to-bile) and sinusoidal (liver-to-blood) excretion. Pharmacokinetic modeling enables molecular interpretation of  $^{99m}\text{Tc}$ -mebrofenin scintigraphy data, thus offering a widely available translational method to investigate transporter-mediated drug-drug interactions *in vivo*.  $^{99m}\text{Tc}$ -mebrofenin allows for phenotyping transporter function at the different poles of hepatocytes as a biomarker of liver function.

## **KEYWORDS**

Solute carrier transporter, ATP-binding cassette transporter, Pharmacokinetics, Mebrofenin, Liver function.

## INTRODUCTION

The quantitative measurement of liver function remains a Holy Grail of modern medicine (1). From a molecular perspective, liver function can be estimated by the activity of membrane transporters expressed in hepatocytes. An array of membrane transporters of the solute carrier (SLC) and adenosine triphosphate-binding cassette (ABC) superfamilies control the hepatobiliary clearance of many endogenous and exogenous compounds. Several SLC influx transporters, including the organic anion-transporting polypeptide (OATP) transporters OATP1B1 and OATP1B3, are expressed in the sinusoidal (blood-facing) membrane of hepatocytes where they mediate the uptake of their substrates from blood into the liver (2). ABC efflux transporters expressed at the canalicular (bile-facing) membrane of hepatocytes, such as the multidrug resistance-associated protein 2 (MRP2) or the bile salt export pump, control the biliary excretion of solutes and bile acids (2). Other ABC transporters are expressed in the sinusoidal hepatocyte membrane where they mediate the backflux of solutes from hepatocytes into blood (Fig. 1). Hepatocyte transporters control the hepatobiliary clearance of many drugs and metabolites. Many marketed drugs have been identified as inhibitors and/or substrates of hepatocyte transporters. There is an associated risk for transporter-mediated drug-drug interaction (DDI) with impact for pharmacokinetics and drug safety (3). Consequently, regulatory authorities request that several hepatocyte transporters are investigated as potential sites for pharmacokinetic DDIs of new drug candidates (2).

Imaging using substrate probes for hepatic transporters bears a great potential to provide molecular insight into liver function (4). However, the development of imaging probes for selective and quantitative imaging of hepatic transporters is a challenge, as described in recent reviews (4–6). Candidate probes are often substrates of multiple hepatic transporters so that true selectivity is difficult to achieve. Moreover, hepatic transporter substrates with predominant liver extraction often undergo extensive metabolism, which complicates the molecular interpretation of the imaging signal measured in the liver

and bile. Metabolically stable probes are therefore preferred for correct estimation of hepatic transporter function (4).

Several  $^{99m}\text{Tc}$ -labeled radiopharmaceuticals have been approved for evaluating liver function in patients. For instance, scintigraphy with  $^{99m}\text{Tc}$ -sulfur colloids has been used to visualize phagocytosis by the reticuloendothelial cells of the liver (7). The binding of  $^{99m}\text{Tc}$ -labeled galactosylneoglycoalbumin to the asialoglycoprotein receptors expressed at the surface of hepatocytes is considered a biomarker of liver function (7). In addition, iminodiacetate derivatives are preferentially taken up by hepatocytes and follow intracellular paths similar to that of endogenous substances making them suitable to assess important liver functions.  $^{99m}\text{Tc}$ -HIDA ( $^{99m}\text{Tc}$ -(2,6-dimethylacetanilide)iminodiacetate) was approved in the mid-1970s for hepatobiliary scintigraphy, followed by  $^{99m}\text{Tc}$ -disofenin ( $^{99m}\text{Tc}$ -diisopropyliminodiacetic acid) and  $^{99m}\text{Tc}$ -mebrofenin ( $^{99m}\text{Tc}$ -N-(3-bromo-2,4,6-trimethylphenylcarbonylmethyliminodiacetic acid) (7,8).  $^{99m}\text{Tc}$ -mebrofenin benefits from improved pharmacokinetic properties such as fast blood clearance caused by rapid and predominant uptake by the liver followed by biliary excretion, with minimal urinary excretion (9,10). Radiolabeling of  $^{99m}\text{Tc}$ -mebrofenin from commercial kits is straightforward, yielding high radiochemical purity and stability (11).  $^{99m}\text{Tc}$ -mebrofenin scintigraphy is therefore widely used in Nuclear Medicine to explore the dynamics of biliary excretion or estimate hepatic extraction capacity in order to assess biliary disorders, such as bile duct obstruction, cholecystitis or gallbladder dysfunction (10,12,13).  $^{99m}\text{Tc}$ -mebrofenin is increasingly used to monitor liver function in the setting of liver surgery and transplantation, for preoperative assessment of future remnant liver function (7,14).

The factors governing the liver specificity and hepatobiliary elimination of  $^{99m}\text{Tc}$ -mebrofenin were not known when this imaging agent was initially developed (15–17). The characterization of various carrier-mediated processes in hepatocytes enabled identifying the transporters involved in the liver kinetics of  $^{99m}\text{Tc}$ -mebrofenin (5). An in-depth characterization of the hepatic transport profile of  $^{99m}\text{Tc}$ -

mebrofenin led to use this widely available radiopharmaceutical as a probe for translational molecular imaging of hepatocyte transporters.

#### **NON-CLINICAL STUDIES ON TRANSPORT OF <sup>99m</sup>Tc-MEBROFENIN**

*In vitro* studies in cells overexpressing selected hepatocyte transporters demonstrated that <sup>99m</sup>Tc-mebrofenin is transported by the influx transporters OATP1B1 and OATP1B3 but neither by OATP2B1, nor by Na<sup>+</sup>-taurocholate cotransporting polypeptide and organic cation transporter 1 (15,17,18). The potent OATP inhibitors rifampicin, rifamycin SV and glycyrrhizic acid markedly decreased <sup>99m</sup>Tc-mebrofenin uptake in transporter overexpressing cells, while organic cation and anion transporter inhibitors did not (17–19). Regarding efflux transporters, <sup>99m</sup>Tc-mebrofenin is predominantly transported by the ABC transporters MRP2 and MRP3 (17,19,20). Efflux transport of <sup>99m</sup>Tc-mebrofenin by MRP2 and MRP3 was inhibited by the MRP inhibitors MK571, ritonavir and E<sub>2</sub>17βG (17,19,20). The importance of carrier-mediated systems for the cellular uptake of <sup>99m</sup>Tc-mebrofenin was confirmed by studies in water-injected oocytes, in which low uptake of <sup>99m</sup>Tc-mebrofenin was observed suggesting negligible passive diffusion across cell membranes (15).

The importance of carrier-mediated transport processes was confirmed *in vivo* using transporter-deficient animals. In OATP-deficient mice, more radioactivity was measured in the blood and urinary bladder, with a considerably lower signal in the liver (18). In MRP2-deficient rats and mice, biliary excretion of <sup>99m</sup>Tc-mebrofenin was reduced compared with wild-type animals, while liver exposure was increased despite a similar initial liver uptake (16,18). Rifampicin, which is an inhibitor of both OATP and MRP2, significantly decreased the hepatic uptake of <sup>99m</sup>Tc-mebrofenin and its transfer to the gallbladder and intestine (18). Consequently, radioactivity was higher in the blood and urinary bladder, suggesting a shift from hepatobiliary to urinary excretion. Cyclosporin A (CsA), known to inhibit several efflux transporters including MRP2, did not further decrease the biliary excretion of <sup>99m</sup>Tc-mebrofenin in MRP2-deficient rats,

suggesting a limited role of other ABC transporters at the liver-bile interface (16). We recently investigated the impact of increasing doses of CsA on the liver kinetics of  $^{99m}\text{Tc}$ -mebrofenin, revealing that CsA was more potent in inhibiting MRP2 than OATP (21). Low-dose CsA (0.01 mg/kg administered intravenously) completely inhibited the MRP2-mediated biliary excretion of  $^{99m}\text{Tc}$ -mebrofenin without affecting the OATP-mediated sinusoidal uptake. It is noteworthy that liver exposure to  $^{99m}\text{Tc}$ -mebrofenin was significantly increased under conditions of decreased biliary excretion, while no significant changes in blood radioactivity were observed. This illustrates that dramatic changes in the liver exposure to drugs can be observed, which may potentially lead to hepatotoxicity, with a non-detectable impact on plasma pharmacokinetics. Pharmacokinetic modeling revealed a dose-dependent inhibition of the sinusoidal efflux (liver-to-blood) of  $^{99m}\text{Tc}$ -mebrofenin by CsA, consistent with the expression of MRP3 at this specific interface (17,21). In rats, the extent of MRP3-mediated sinusoidal and MRP2-mediated canalicular efflux transport of  $^{99m}\text{Tc}$ -mebrofenin was similar but remained much lower than the OATP-mediated sinusoidal uptake, thus providing information regarding the relative importance of carrier-mediated processes in hepatocytes (21). Imaging probes for quantitative determination of MRP2 and MRP3 function in hepatocytes are still not available (6). The non-clinical transporter selectivity profile of  $^{99m}\text{Tc}$ -mebrofenin suggests it may be repurposed as a translational molecular imaging probe for simultaneous exploration of OATP, MRP2 and MRP3 function at the different poles of hepatocytes (Fig. 1).

#### **CLINICAL STUDIES WITH $^{99m}\text{Tc}$ -MEBROFENIN TO INVESTIGATE HEPATOCYTE TRANSPORTERS**

Prof. Kim Brouwer and colleagues have pioneered the use of  $^{99m}\text{Tc}$ -mebrofenin as a phenotypic probe to explore hepatocyte transporter function in humans (17,20,22). The kinetics of  $^{99m}\text{Tc}$ -mebrofenin in humans were first interpreted using a two-compartment model, which suggested canalicular efflux (MRP2) rather than sinusoidal efflux (MRP3) as the rate limiting step for controlling the plasma and liver kinetics of  $^{99m}\text{Tc}$ -mebrofenin in humans (17). This modeling approach was also applied to simulated pathophysiological abnormalities, such as hyperbilirubinemia and cholestasis. Mathematical simulations

concluded that hyperbilirubinemia would decrease the hepatic exposure to  $^{99m}\text{Tc}$ -mebrofenin, associated with increased blood concentrations (17). This may be explained by competition of plasma bilirubin with  $^{99m}\text{Tc}$ -mebrofenin for OATP-mediated uptake into the liver (23). Simulations of impaired biliary excretion due to cholestasis resulted in prolonged hepatic exposure to  $^{99m}\text{Tc}$ -mebrofenin. According to the model, cholestasis is expected to increase the blood exposure to  $^{99m}\text{Tc}$ -mebrofenin due to extensive hepatic accumulation and increased sinusoidal efflux, suggesting an involvement of MRP3-mediated sinusoidal transport (17).

This molecular imaging approach was then tested in a cohort of patients suffering from non-alcoholic steatohepatitis (NASH) (22), in whom regulatory changes in hepatocyte transporter expression have been reported (24,25). Plasma and liver exposure to  $^{99m}\text{Tc}$ -mebrofenin were increased in the NASH cohort by 1.4 and 1.6-fold, respectively (22). Both the hepatic uptake and biliary efflux clearances of  $^{99m}\text{Tc}$ -mebrofenin were reduced in NASH patients compared to healthy subjects. From a molecular perspective, this suggested impaired MRP2-mediated canalicular efflux transport of  $^{99m}\text{Tc}$ -mebrofenin in NASH, consistent with previous studies finding a mislocalization of MRP2 in the canalicular hepatocyte membrane of NASH patients (25). No difference was observed in the sinusoidal efflux clearance of  $^{99m}\text{Tc}$ -mebrofenin although an increase in hepatic MRP3 expression and function has been previously found in NASH patients using other methods (25,26). Interestingly, liver uptake of  $^{99m}\text{Tc}$ -mebrofenin was slightly lower in healthy subjects harbouring a low (*SLCO1B1*\*15/\*15) and intermediate (*SLCO1B1*\*1A/\*15) function genotype of the gene encoding for OATP1B1, although liver uptake remained higher than that observed in NASH patients (22). These pioneer clinical data highlight the potential of  $^{99m}\text{Tc}$ -mebrofenin scintigraphy with kinetic modeling to explore the functional consequences of the patho-physiological regulation of certain hepatocyte transporters.

#### **MOLECULAR INSIGHT INTO $^{99m}\text{Tc}$ -MEBROFENIN LIVER KINETICS**

Knowledge of the molecular determinants of the hepatic handling of  $^{99m}\text{Tc}$ -mebrofenin may shed light on results of abnormal  $^{99m}\text{Tc}$ -mebrofenin scintigraphy. A case of a patient with Rotor syndrome was reported 30 years ago, who exhibited decreased and delayed liver uptake of  $^{99m}\text{Tc}$ -mebrofenin with normal biliary excretion and persistent visualization of the cardiac blood pool, kidneys and urinary bladder (27). The etiology of this disease is now identified and consists of mutations in the *SLCO1B1* and *SLCO1B3* genes, encoding non-functional OATP1B1 and OATP1B3 transport proteins (28). This provided mechanistic explanation for the altered liver kinetics of  $^{99m}\text{Tc}$ -mebrofenin observed in this patient. A study in rats with inflammatory induced-liver injury showed a reduced hepatic uptake of  $^{99m}\text{Tc}$ -mebrofenin as compared to control rats (29), which is consistent with changes in hepatic transporter expression, including a decrease in OATP1B1 expression (30).

These data suggest that pathophysiological modulation of hepatocyte transporter function may impact the liver kinetics of  $^{99m}\text{Tc}$ -mebrofenin. Moreover, concomitant drug intake may also alter the hepatic disposition of  $^{99m}\text{Tc}$ -mebrofenin. Many marketed drugs are known to inhibit hepatic transporters handling  $^{99m}\text{Tc}$ -mebrofenin (Table 1).  $^{99m}\text{Tc}$ -mebrofenin may therefore become an unintended victim of transporter-mediated DDIs, which may confound the interpretation of liver scintigraphy images. For instance, low hepatic clearance of  $^{99m}\text{Tc}$ -mebrofenin was recently reported in a patient treated for hepatitis C with grazoprevir, an OATP1B1/3 substrate, suggesting competitive inhibition of hepatic uptake transport (Fig. 2) (31). CsA is a potent inhibitor of the OATP- and MRP2-mediated transport of  $^{99m}\text{Tc}$ -mebrofenin (21). A study performed in patients undergoing liver transplantation showed that hepatic extraction of  $^{99m}\text{Tc}$ -mebrofenin was consistently lower in CsA-treated patients compared with tacrolimus-treated patients, despite similar liver function assessed using biochemistry assays (32). These reports provide evidence that concomitant drug intake may precipitate transporter-mediated DDIs with  $^{99m}\text{Tc}$ -mebrofenin (21).



On the other hand, several pharmacokinetic studies have illustrated the potential of  $^{99m}\text{Tc}$ -mebrofenin as a probe to specifically investigate DDIs involving OATP, MRP2, and probably MRP3, at both sinusoidal and canalicular membranes of hepatocytes in drug development. The great advantage of  $^{99m}\text{Tc}$ -mebrofenin is that it is a metabolically stable probe which allows to selectively assess hepatocyte transporter function and which is not affected by changes in the activity of metabolizing enzymes. For instance, ritonavir was shown to inhibit several hepatocyte transporters *in vitro* and is also a powerful inhibitor of cytochrome P450 enzymes (20). A semiphysiologically-based pharmacokinetic model showed that ritonavir significantly increased systemic exposure to  $^{99m}\text{Tc}$ -mebrofenin, without affecting overall hepatic exposure or biliary recovery. Simulations suggested that clinical doses of ritonavir may inhibit OATP-mediated hepatic uptake but not the MRP2-mediated biliary excretion, thus pinpointing the corresponding risk of transporter-mediated DDIs and its consequences at the organ and hepatocyte level (20).

The possibility for DDIs selectively occurring at the level of canalicular hepatocyte transporters, with no impact on plasma pharmacokinetics but dramatic increases in liver exposure has been confirmed with low-dose CsA using  $^{99m}\text{Tc}$ -mebrofenin in rats (21). Such “silent” DDIs, which cannot be detected from conventional plasma pharmacokinetics, may result in increased hepatic accumulation of drugs. This situation is increasingly considered as a mechanistic explanation for drug-induced liver injury (DILI), which is a major safety concern in drug development. Many drugs have an inhibitory effect on MRP2 *in vitro* (Table 1) and  $^{99m}\text{Tc}$ -mebrofenin imaging may be used in drug development to predict the corresponding risk of pharmacokinetic DDIs in humans (33). These new applications extend the scope of this widely available molecular imaging probe beyond “classical” hepatobiliary scintigraphy to explore hepatocyte transporter function in health and disease.

Compared with positron emission tomography (PET), gamma scintigraphy is traditionally considered a poorly quantitative imaging modality. Liver scintigraphy typically involves dynamic

acquisitions with single-head 2D gamma-cameras which enables capturing the rapid uptake and biliary excretion of  $^{99m}\text{Tc}$ -mebrofenin (4). However, such dynamic planar images lack the ability to assess liver function at the segmental level, therefore requiring the assumption of homogenous hepatic function (14). With the introduction of hybrid SPECT-CT systems, significant advances in image reconstruction with sophisticated compensation techniques have been made which now enable correction for photon attenuation and scattering. Quantitative SPECT-CT data can now be obtained in accurately delineated liver volumes and segments, although suffering from low temporal resolution as compared with PET or gamma scintigraphy (34). Detailed knowledge regarding the carrier-mediated systems that control the liver kinetics of [ $^{99m}\text{Tc}$ ]mebrofenin may be used to improve the pharmacokinetic properties of this radiopharmaceutical with respect to tomographic acquisition. In rats, blocking of the biliary excretion of  $^{99m}\text{Tc}$ -mebrofenin was achieved by targeted inhibition of MPR2 using low-dose CsA, which prolonged the liver uptake phase and enhanced the liver exposure to  $^{99m}\text{Tc}$ -mebrofenin (21). Since CsA is a clinically approved drug, these optimized conditions may be potentially applied in a clinical setting for an improved assessment of functional liver volume with  $^{99m}\text{Tc}$ -mebrofenin and SPECT.

## **CONCLUSION**

The discovery and characterization of different hepatocyte transporters has shed new light on the molecular determinants of the liver, bile and plasma kinetics of  $^{99m}\text{Tc}$ -mebrofenin. These properties enlarged the perspectives of this radiopharmaceutical agent to be repurposed for translational molecular imaging of liver transporter function. Enhanced knowledge of the liver transport of  $^{99m}\text{Tc}$ -mebrofenin offers a molecular reading of scintigraphy data in various clinical situations with impaired liver function (Table 2).

## **ACKNOWLEDGEMENTS**

The authors would like to thank Thomas Beyer (Medical University of Vienna) for critically assessing their article.

## **DISCLOSURE STATEMENT**

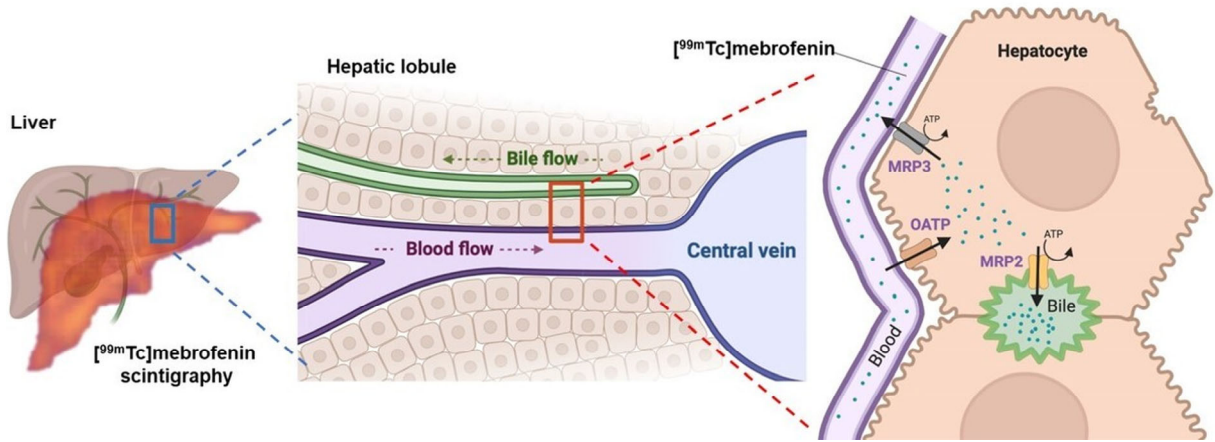
No potential conflicts of interest relevant to this article exist.

## REFERENCES

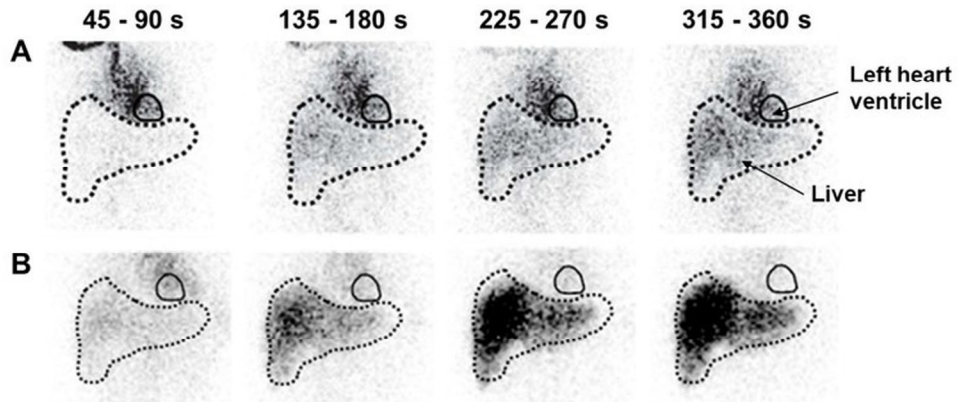
1. Gholam P, Lee Z. Quantitative measurement of liver function: The quest for the holy grail? *J Nucl Med.* 2011;52:169-170.
2. Patel M, Taskar K, Zamek-Gliszczyński M. Importance of hepatic transporters in clinical disposition of drugs and their metabolites. *J Clin Pharmacol.* 2016;56:S23-S39.
3. Giacomini KM, Huang S-M, Tweedie DJ, et al. Membrane transporters in drug development. *Nat Rev Drug Discov.* 2010;9:215-236.
4. Tournier N, Stieger B, Langer O. Imaging techniques to study drug transporter function in vivo. *Pharmacol Ther.* 2018;189:104-122.
5. Marie S, Cisternino S, Buvat I, Declèves X, Tournier N. Imaging probes and modalities for the study of Solute Carrier O (SLCO)-transport function in vivo. *J Pharm Sci.* 2017;106:2335-2344.
6. Hernández Lozano I, Langer O. Use of imaging to assess the activity of hepatic transporters. *Expert Opin Drug Metab Toxicol.* 2020;16:149-164.
7. de Graaf W, Bennink R, Veteläinen R, van Gulik T. Nuclear imaging techniques for the assessment of hepatic function in liver surgery and transplantation. *J Nucl Med.* 2010;51:742-752.
8. Nunn A, Loberg MD, Conley RA. A structure-distribution-relationship approach leading to the development of Tc-99m mebrofenin: an improved cholescintigraphic agent. *J Nucl Med.* 1983;24:423-430.
9. Fritzberg A, Klingensmith W. Quest for the perfect hepatobiliary radiopharmaceutical. *J Nucl Med.* 1982;23:543-546.
10. Krishnamurthy S, Krishnamurthy G. Technetium-99m-iminodiacetic acid organic anions: Review of biokinetics and clinical application in hepatology. *Hepatology.* 1989;9:139-153.
11. Lan J, Chervu L, Johansen K, Wolkoff A. Uptake of technetium 99m hepatobiliary imaging agents by cultured rat hepatocytes. *Gastroenterology.* 1988;95:1625-1631.
12. Gad MA, Krishnamurthy GT, Glowniak JV. Identification and differentiation of congenital gallbladder abnormality by quantitative technetium-99m IDA cholescintigraphy. *J Nucl Med Off Publ Soc Nucl Med.* 1992;33:431-434.
13. Tulchinsky M. The SNM practice guideline on hepatobiliary scintigraphy. *J Nucl Med.* 2010;51:1825-1825.
14. de Graaf W, van Lienden K, van Gulik T, Bennink R. 99mTc-mebrofenin hepatobiliary scintigraphy with SPECT for the assessment of hepatic function and liver functional volume before partial hepatectomy. *J Nucl Med.* 2010;51:229-236.
15. de Graaf W, Häusler S, Heger M, et al. Transporters involved in the hepatic uptake of 99mTc-mebrofenin and indocyanine green. *J Hepatol.* 2011;54:738-745.

16. Bhargava K, Joseph B, Ananthanarayanan M, et al. Adenosine triphosphate-binding cassette subfamily C member 2 Is the major transporter of the hepatobiliary imaging agent 99mTc-mebrofenin. *J Nucl Med.* 2009;50:1140-1146.
17. Ghibellini G, Leslie E, Pollack G, Brouwer K. Use of Tc-99m mebrofenin as a clinical probe to assess altered hepatobiliary transport: integration of in vitro, pharmacokinetic modeling, and simulation studies. *Pharm Res.* 2008;25:1851.
18. Neyt S, Huisman M, Vanhove C, et al. In vivo visualization and quantification of (disturbed) Oatp-mediated hepatic uptake and Mrp2-mediated biliary excretion of 99mTc-mebrofenin in mice. *J Nucl Med.* 2013;54:624-630.
19. Swift B, Yue W, Brouwer K. Evaluation of 99mtechnetium-mebrofenin and 99mtechnetium-sestamibi as specific probes for hepatic transport protein function in rat and human hepatocytes. *Pharm Res.* 2010;27:1987-1998.
20. Pfeifer N, Goss S, Swift B, et al. Effect of ritonavir on 99mtechnetium-mebrofenin disposition in humans: A semi-PBPK modeling and in vitro approach to predict transporter-mediated DDIs. *CPT Pharmacomet Syst Pharmacol.* 2013;2:e20.
21. Marie S, Hernández-Lozano U, Breuil L, et al. Validation of pharmacological protocols for targeted inhibition of canalicular MRP2 activity in hepatocytes using [99mTc]mebrofenin imaging in rats. *Pharmaceutics.* 2020;12:486.
22. Ali I, Slizgi J, Kaullen J, et al. Transporter-mediated alterations in patients with NASH increase systemic and hepatic exposure to an OATP and MRP2 substrate. *Clin Pharmacol Ther.* 2017;104:749-756.
23. Cui Y, König J, Leier I, Buchholz U, Keppler D. Hepatic uptake of bilirubin and its conjugates by the human organic anion transporter SLC21A6. *J Biol Chem.* 2001;276:9626-9630.
24. Clarke J, Hardwick R, Lake A, et al. Synergistic interaction between genetics and disease on pravastatin disposition. *J Hepatol.* 2014;61:139-147.
25. Hardwick RN, Fisher CD, Canet MJ, Scheffer GL, Cherrington NJ. Variations in ATP-binding cassette transporter regulation during the progression of human nonalcoholic fatty liver disease. *Drug Metab Dispos.* 2011;39:2395-2402.
26. Ferslew B, Johnston C, Tsakalozou E, et al. Altered morphine glucuronide and bile acid disposition in patients with non-alcoholic steatohepatitis. *Clin Pharmacol Ther.* 2015;97:419-427.
27. LeBouthillier G, Morais J, Picard M, Picard D, Chartrand R, Pomier G. Scintigraphic Aspect of Rotor's disease with technetium-99m-mebrofenin. *J Nucl Med.* 1992;33:1550-1551.
28. van de Steeg E, Stránecký V, Hartmannová H, et al. Complete OATP1B1 and OATP1B3 deficiency causes human Rotor syndrome by interrupting conjugated bilirubin reuptake into the liver. *J Clin Invest.* 2012;122:519-528.

29. Joseph B, Bhargava K, Tronco G, Kumaran V, Palestro C, Gupta S. Regulation of hepatobiliary transport activity and noninvasive identification of cytokine-dependent liver inflammation. *J Nucl Med*. 2005;46:146-152.
30. Evers R, Piquette-Miller M, Polli J, et al. Disease-associated changes in drug transporters may impact the pharmacokinetics and/or toxicity of drugs: a white paper from the International Transporter Consortium. *Clin Pharmacol Ther*. 2018;104:900-915.
31. Deshayes E, Fersing C, Meunier L, Quenet F, Guiu B. Antiviral (hepatitis C virus) drug-drug interaction leading to dramatic underestimation of liver function with <sup>99m</sup>Tc-mebrofenin hepatobiliary scintigraphy. *Clin Nucl Med*. 2020;45:133-135.
32. Kaxiras A, Yamamoto S, Söderdahl G, Wernerson A, Axelsson R, Ericzon B. Cyclosporin A, but not tacrolimus, negatively affects the hepatic extraction fraction of hepatobiliary scintigraphy in liver transplant recipients. *EJNMMI Res*. 2014;4:73.
33. Liu H, Sahi J. Role of hepatic drug transporters in drug development. *J Clin Pharmacol*. 2016;56:S11-S22.
34. Bailey D, Willowson K. Quantitative SPECT/CT: SPECT joins PET as a quantitative imaging modality. *Eur J Nucl Med Mol Imaging*. 2014;41:17-25.
35. Matsson P, Pedersen J, Norinder U, Bergström C, Artursson P. Identification of novel specific and general inhibitors of the three major human ATP-binding cassette transporters P-gp, BCRP and MRP2 among registered drugs. *Pharm Res*. 2009;26:1816-1831.
36. Estudante M, Soveral G, Morais J, Benet L. Insights into solute carriers: physiological functions and implications in disease and pharmacokinetics. *MedChemComm*. 2016;7:1462-1478.
37. Karlgren M, Vildhede A, Norinder U, et al. Classification of inhibitors of hepatic organic anion transporting polypeptides (OATPs): influence of protein expression on drug–drug interactions. *J Med Chem*. 2012;55:4740-4763.
38. Köck K, Ferslew B, Netterberg I, et al. Risk factors for development of cholestatic drug-induced liver injury: inhibition of hepatic basolateral bile acid transporters multidrug resistance-associated proteins 3 and 4. *Drug Metab Dispos*. 2014;42:665-674.



**FIGURE 1.** Multiscale physiological and molecular interpretation of the liver kinetics of  $^{99m}\text{Tc}$ -mebrofenin.



**FIGURE 2.** Impact of concomitant drug intake on clinical <sup>99m</sup>Tc-mebrofenin scintigraphy data. Dynamic acquisition (anterior view, 45-second summed planar images over 6 minutes) was acquired after injection of <sup>99m</sup>Tc-mebrofenin in a 42-year-old woman treated with grazoprevir (A) and at 3 weeks after the end of the treatment (B). Liver uptake of <sup>99m</sup>Tc-mebrofenin was markedly reduced under grazoprevir treatment (A), presumably due to competitive inhibition of OATP1B1/3-mediated liver uptake of <sup>99m</sup>Tc-mebrofenin. Adapted from Clin Nucl Med. Deshayes et al. Clin Nucl Med 2020;45(2)133-135. (31)



**TABLE 1**

Drugs Inhibiting *in vitro* the Transporters Involved in the Hepatic Disposition of <sup>99m</sup>Tc-mebrofenin (35–38)

Transporters	Inhibitors
<b>OATP1B1</b>	atazanavir, atorvastatin, budesonide, carfilzomib, cyclosporin, clarithromycin, diclofenac, dipyridamole, dronedarone, elacridar, erythromycine, flavonoids, fluvastatin, fosinopril, gemfibrozil, glibenclamide, gliquidone, lapatinib, lopinavir, pitavastatin, rapamycin, repaglinide, rifampicin, ritonavir, rosuvastatin, roxithromycin, sirolimus, sulfasalazin, telithromycin, telmisartan, trametinib, valspodar.
<b>OATP1B3</b>	atazanavir, atorvastatin, budesonide, carfilzomib, clarithromycin, cyclosporin, clarithromycin, dipyridamole, dronedarone, flavonoids, fluvastatin, fosinopril, gemfibrozil, glibenclamide, gliquidone, lapatinib, lopinavir, pitavastatin, rapamycin, repaglinide, rifampicin, ritonavir, rosuvastatin, sildenafil, sirolimus, sulfasalazin, telmisartan, trametinib, valspodar.
<b>MRP2</b>	benzbromarone, cyclosporin, diltiazem, dipyridamole, ivermectin, lansoprazole, loperamide, lopinavir, reserpine, rifampicin, ritonavir, saquinavir, silymarin, tamoxifen, terfenadine.
<b>MRP3</b>	alpidem, benzbromarone, chloramphenicol, chlorpromazine, clozapine, cyclosporin, desipramine, dicloxacillin, etoposide, finasteride, fluoxetine, fluvastatin, furosemide, glibenclamide, haloperidol, indinavir, indomethacin, lopinavir, maprotiline, mibefradil, nadolol, nifedipine, nortryptiline, oxybutynin, primaquine, probenecid, promethazine, quinidine, rifamycin SV, ritonavir, simvastatin, sorafenib, sulfasalazine, tamoxifen, taxol, tolbutamide, tolcapone, troglitazone, valinomycin, verapamil, vinblastine.

## TABLE 2

### Future Perspectives of $^{99m}\text{Tc}$ -mebrofenin for Clinical Applications

- $^{99m}\text{Tc}$ -mebrofenin represents a translational molecular imaging probe to explore canalicular MRP2 as well as sinusoidal OATP and MRP3 function in hepatocytes.
- In patients, kinetic modeling may enable interpreting dynamic  $^{99m}\text{Tc}$ -mebrofenin scintigraphy data from a molecular perspective.
- Owing to its widespread availability,  $^{99m}\text{Tc}$ -mebrofenin can be potentially used as a phenotyping probe to safely and selectively investigate the risk for transporter-mediated drug-drug interactions in drug development and to explore pathophysiological regulations of hepatocyte transporter function.
- Preclinical data suggest that the liver uptake phase of  $^{99m}\text{Tc}$ -mebrofenin can be prolonged by targeted inhibition of MRP2, thereby potentially providing an improved assessment of functional liver volume with SPECT.