A Phase 1 and Biodistribution Study of ABT-806i, An Indium-111 Radiolabeled Conjugate of the Tumor-Specific Anti-EGFR Antibody ABT-806

Hui K. Gan^{1,2,3}, Matthew Burge^{4,5}, Benjamin Solomon^{3,6}, Sze Ting Lee^{1,2,7}, Kyle D. Holen⁸, Yumin Zhang⁹, Marika Ciprotti¹, FT Lee¹, Wijith Munasinghe⁸, JuDee Fischer⁸, Peter Ansell⁸, Gerard Fox⁸, Hao Xiong⁸, Edward B. Reilly⁸, Rod Humerickhouse⁸ and Andrew M. Scott^{1,2,3,7}

¹Olivia Newton-John Cancer Research Institute, Austin Health, Melbourne, Australia; ²School of Cancer Medicine, La Trobe University, Melbourne, Australia; ³Department of Medicine, University of Melbourne, Melbourne, Australia; ⁴Royal Brisbane and Women's Hospital, Brisbane, Australia; ⁵University of Queensland, Brisbane, Australia; ⁶Peter MacCallum Cancer Centre, Melbourne, Australia; ⁷Department of Molecular Imaging and Therapy, Austin Health, Melbourne, Australia; ⁸AbbVie, 1 North Waukegan Road, North Chicago, IL, USA; ⁹Sinotau Pharmaceutical Group, Beijing 102206, China.

Running title: Phase I Study of ABT-806i against EGFR

Corresponding author and First Author:

Hui K. Gan, MBBS, PhD

Director of the Cancer Clinical Trial Center

Austin Hospital, 145 Studley Rd.

Heidelberg VIC 3084, Australia

Phone: +61 3 9496 9925

Fax: +61 3 9457 6698

Email: <u>hui.gan@onjcri.org.au</u>

Word count: 4036/5000

Total figures: 4

Total Tables: 5 and 1 Supplementary (total of 6)

Abstract (283 words)

ABT-806 is a tumor-specific antibody targeting the epidermal growth factor receptor (EGFR). This study assessed safety, biodistribution and pharmacokinetics of indium-111 (¹¹¹In) radiolabeled ABT-806 (ABT-806i) and effects of repeated doses of ABT-806 on receptor occupancy.

Methods: Eligible patients had advanced tumors likely to express EGFR/EGFRvIII; adequate performance status and organ function; and measurable disease by RECIST 1.1. In Cohort 1, 6 patients received a bolus administration of ABT-806i and underwent single-photon emission computed tomography (SPECT) followed by whole body planar scans. In Cohort 2, 12 patients were imaged similarly as in 1 initially; thereafter they received three doses of unlabelled ABT-806, before another dose of ABT-806i with associated SPECT and whole body planar scans. At the end of both cohorts, patients who had stable or responding disease were able to enroll into an extension study (M12-326) where they received unlabelled ABT-806 every 2 weeks until disease progression, withdrawal of consent or intolerable toxicity

Results: No toxicity related to ABT-806i infusion were observed. ABT-806i showed minimal uptake in normal tissues and cleared gradually from blood with t_{42} of 6.0 ± 1.5 days. The mean effective dose of ABT-806i was determined as 0.137 mSv/MBq for males and 0.183 mSv/MBq for females. ABT-806i tumor uptake varied and did not correlate with archived tumor EGFR expression. No change in ABT-806i uptake was observed after interval ABT-806 treatment, indicating stable EGFR expression in tumor. The patient with highest tumor uptake of ABT-806i had advanced head and neck cancer and experienced a partial response.

Conclusion: ABT-806i allows for real-time imaging of EGFR conformational expression in tumors, has an acceptable radiation dosimetry and provides important additional information about

antigen expression compared to standard approaches using archival tissue. Its role to assist in patient selection for EGFR-based therapeutics and investigate treatment resistance should be further investigated.

Keywords: EGFR, EGFRvIII, Solid Tumors, SPECT Imaging, Biodistribution

Introduction

The *Epidermal Growth Factor Receptor* (*EGFR*) gene is a validated target in oncology. Monoclonal antibodies against EGFR are used to treat cancers of the head and neck, colon and lung (1,2). Tyrosine kinase inhibitors of EGFR are used in the treatment of lung cancer with activating kinase mutations (1,2). The most common toxicity of these agents is a well-characterized skin rash (3). Other toxicities include diarrhea, stomatitis, fatigue, and electrolyte disturbances.

ABT-806 is a humanized recombinant IgG1 κ antibody that is specific for a unique, conformationally exposed epitope of EGFR. This epitope is available for binding only under conditions where there is dysregulated EGFR activation due to conditions such as EGFR amplification, presence of specific mutations such as EGFRde2-7 (EGFRvIII) or presence of autocrine loops (1,4-6). The epitope is inaccessible when EGFR is expressed at normal physiological levels, thus ABT-806 has limited binding to normal tissues (1,4-6). ABT-806 reproducibly inhibits the growth of multiple EGFR amplified/EGFRvIII mutated tumors preclinically through down-regulation of EGFR signaling with result anti-tumor changes in proliferation, angiogenesis and apoptosis (1,7-9). When compared to other EGFR-directed therapies, preclinical studies show ABT-806 has a favorable toxicity profile and equivalent/improved efficacy to other anti-EGFR antibodies (1,8,10,11). In a Phase 1 human study of ABT-806 (Study M11-847, NCT01255657), the recommended Phase 2 dose (RPTD) was 24 mg/kg (12). Importantly, ABT-806 was shown to have negligible skin or other organ toxicity (12,13).

ABT-806i, an indium-111 radiolabeled conjugate of ABT-806, is a novel radiopharmaceutical which was developed for real time scintigraphic imaging of EGFR. It utilizes

the tumor-specific binding affinity of ABT-806 for the imaging of EGFR-positive tumors to determine if EGFR amplification, overexpression or mutation is present. We report herein the results of a first-in-human trial of ABT-806i, exploring the ability to image the conformational epitope of EGFR bound by ABT-806, the impact of ABT-806 therapy on ABT-806i uptake, and the relationship of ABT-806i uptake to tumor EGFR by immunohistochemistry (IHC).

Materials and Methods

Trial Objectives

This first-in-human trial (M11-849, NCT01472003) was a two-cohort, open label, multicenter study to determine the biodistribution and dosimetry of ABT-806i. Secondary objectives included characterization of ABT-806i pharmacokinetics (PK); examination of ABT-806i SPECT imaging in tumors of different histologies, EGFR expression and size; and determining the effect of unlabelled ABT-806 on ABT-806i receptor occupancy. The study was approved by the Austin Health Human Research Ethics Committee and all subjects signed an informed consent form.

Patient Eligibility for M11-849 Bioimaging Study

Eligible patients were aged ≥ 18 years; had tumors of a type likely to overexpress wildtype EGFR/EGFRvIII; were Eastern Cooperative Oncology Group (ECOG) Performance Status of 0-2; had measurable disease by RECIST version 1.1 with at least one extrahepatic 2 cm lesion; had adequate organ function, defined as absolute neutrophil count $\geq 1500/\text{mm}^3$; platelets \geq 100,000/mm³; hemoglobin ≥ 9.0 g/dL; creatinine ≤ 1.5 times the upper limit of the institution's normal range (xULN); bilirubin, aspartate aminotransferase (AST), and alanine aminotransferase (ALT) ≤ 1.5 x ULN (those with liver metastases were eligible if their AST and ALT was ≤ 5.0 x ULN). Exclusion criteria included anticancer therapy within 14 days of the first ABT-806i dose; prior use of EGFR-directed monoclonal antibody within 4 weeks of the first ABT-806i dose; presence of unresolved Common Terminology Criteria for Adverse Events (CTCAE) Grade 2 or higher toxicity from prior therapy; major surgery within 21 days prior to the first ABT-806i dose; significant comorbidities which posed an unacceptable risk of toxicities; history of major immunologic reaction to any IgG containing agent; and pregnancy or lactating patients.

Radiolabelling of ABT-806i

ABT-806i was formed by mixing a formulated precursor, DTPA-ABT-806, with commercially available indium-111 trichloride. DTPA-ABT-806 was produced from the conjugation of ABT-806 with the isothiocyanate of the phenylisothiocyanate derivative of dientylene triamine pentacetic acid (CHX-A"-DTPA). An average of 2-3 CHX-A"-DTPA moieties were conjugated per antibody. For radiolabeling, 0.5M sodium acetate (pH 7.2, 1.0 mL) was added to indium-111 trichloride (5-7.5 mCi), and then placed in a sterile reaction vial. DTPA-ABT-806 (amount 5mg/2ml) was added to the reaction vial at room temperature, and after mixing was withdrawn into a 5 ml syringe. The total reaction time was 30 minutes. Preclinical studies confirmed retention of binding affinity after radiolabeling (immunoreactive fraction >65%), radiochemical purity >90%, and serum stability studies showed retention of immunoreactivity (data not shown).

Trial Design of M11-849 study

Patients were treated as in Figure 1. Patients in Cohort 1 received a single IV administration of ABT-806i to determine baseline drug distribution. Pre-dose scintigraphic scanning with cobalt-

57 transmission (including whole body planar and SPECT) were performed on day 1 prior to bolus administration of ABT-806i (a protein tracer dose of 3-5 mg labelled with 4-7 mCi (148-259 MBq) of indium) preceded approximately 30 minutes prior by 500-1000 mg acetaminophen and 25-50 mg IV promethazine. We and others have previously demonstrated that this dose of ¹¹¹In (4-7 mCi) is optimal for diagnostic and dosimetric analysis of intact antibodies (*13-15*). Post-ABT-806i administration, patients underwent whole body planar and SPECT scans on days 1, 2, 3 or 4, 5 or 6, and then on day 7. Thereafter, all patients were eligible to enrol on the extension study (Study M12-326, NCT01406119), which is described below.

In Cohort 2, patients underwent an initial week of imaging for the purposes of determining the biodistribution of ABT-806i, in a manner similar to patients treated on Cohort 1. In this first week, they received ABT-806i (4-7 mCi (148-259 MBq)/3-5 mg) on Day 1 of week 1; this was then followed by whole body planar and SPECT scans on days 1, 3 or 4, 5 or 6, and then on day 7. Beginning on day 1 of week 2, they were treated with either 18 mg/kg or 24 mg/kg of unlabelled ABT-806 every fortnight (weeks 2, 4 and 6); in week 6, they also underwent repeat imaging with another dose of ABT-806i after the unlabelled ABT-806 to determine the effects of unlabelled antibody on receptor occupancy. Imaging after this dose of ABT-806i was on days 2 or 3, 4 or 5, and then day 7 of week 6. At the end of the 6 weeks, all patients with disease response or control were eligible to enrol on the extension study (Study M12-326, NCT01406119).

The (M12-326, NCT01406119) was an extension study to allow patients on the bioimaging study above (M11-849) who had completed the required imaging and had not progressed to continue treatment with unlabelled ABT-806 until withdrawal of consent, disease progression, need for other cancer therapy or intolerable toxicity.

Pharmacokinetics during M11-849 study

Pharmacokinetic samples for ABT-806i and ABT-806 were collected prior to dosing and up to 168 hours post dosing on day 1 of week 1 (Cohorts 1 and 2) and up to 336 hours post dosing on day 1 of week 6 (Cohort 2 only). These were measured using the same validated electrochemiluminescence immunoassay and analysed using noncompartmental methods (*16*). Serum samples for antibody drug antibody (ADA) were collected prior to dosing on week 1, day 1 and at the end of study.

Biodistribution and Dosimetry during M11-849 study

From whole-body planar and SPECT images, qualitative analysis of the distribution of ABT-806i in the body and tumor uptake was assessed. Tumor uptake in defined target lesions was evaluated on a four-point scale, with comparison of tumor uptake between initial trace infusion of ABT-806i, and following the second infusion of ABT-806i, to evaluate alteration in EGFR expression following ABT-806 treatment, as well as to define possible receptor occupancy with ABT-806 treatment. The quantitative analysis of ABT-806i in normal tissues was determined in the patients in Cohort 1 after injection of ABT-806i by Region of Interest (ROI) technique. Radioactivity as percentage of injected dose per organ (%ID) was measured in various organs of the patients at 5 time points over 7 days (approximately 168 hours) post-injection. For each patient the %ID values of each organ were plotted against time to generate organ biokinetics to obtain organ radiation exposure (*17*).

Tumor uptake of ABT-806i was measured based on SPECT imaging before and after ABT-806 treatment and plotted against time. Target lesions were selected based on tumor size, with minimum size of approximately 2 cm in the longest transverse diameter on computerized tomography (CT), and sufficient tumor-to-background ratio, ideally greater than approximately 1.5. A major vessel or the heart, selected based on the anatomy in the field of view (FOV), was chosen as the blood pool ROI to estimate radioactivity in the whole blood at the time of imaging (14).

Adverse events during M11-849 study

Analyses of toxicity included only treatment-emergent adverse events (TEAEs) in the first 30 days after first dose of ABT-806i. TEAEs were summarized by system organ class and preferred term according to the Medical Dictionary for Regulatory Activities (MedDRA). The severity of toxicity was assessed using NCI CTCAE Version 4.0 toxicity.

Sample size of M11-849 study

A pragmatic sample size of 26-30 patients was chosen, with 6 patients in Cohort 1 and the rest in Cohort 2. The study was terminated after 18 patients as sufficient data had been gathered with tolerability of highest available dose. Summary statistics for demographics are provided in Table 1.

Results

Patients

Patient demographics are shown in Table 1. The first 6 patients were imaged in Cohort 1 and then proceeded immediately to treatment with the ABT-806 on the M12-326 extension study

(Figure 1 and Table 2). In Cohort 2, 6 patients were treated at the 18 mg/kg dose level, and 6 patients at the 24 mg/kg dose level. At the end of Cohort 2, 5/12 patients had stable disease (SD) with formal MRI restaging at the end of the imaging study and proceeded to the M12-326 extension study (Figure 1 and Table 2). Of these, 1 patient in Cohort 2 with SCCHN experienced partial response (Patient 8, Table 2). In total, 11 out of the 18 patients on the M11-849 biodistribution study had stable or better disease at the end of the study and were screened for enrolment in the M12-326 extension study (Table 2).

Biodistribution of ABT-806i in the M11-849 Biodistribution Study

Evaluation of imaging datasets showed initial pooling of ABT-806i in the cardiac blood pool and within large vessels, liver, spleen and kidneys, followed by gradual blood clearance and washout of ABT-806i from normal tissues over the 8 days of imaging in all patients (Figure 2). In the patients with uptake of ABT-806i in tumor, this was best seen after day 3 with increasing intensity up to day 8 (Figure 2). High, specific uptake of ABT-806i in tumor was visualized in the target lesions for 5/6 patients in Cohort 1 with the other patients having uptake. High specific uptake was seen in all 12 patients in Cohort 2. The uptake of ABT-806i was observed to peak at the 7th or 8th day post-injection. Interestingly, both patients with primary brain tumors showed excellent ABT-806i uptake (Figure 2). Furthermore, the increasing accumulation of ABT-806i over time in tumor (maximal uptake around 4 days), despite decline in serum concentration of ABT-806i during this timeframe, would support specific uptake in the glioblastoma tumors imaged in the study. The peak uptake of ABT-806i in target tumor volumes in Cohort 1 and 2 are shown in Tables 3 and 4 respectively.

Pharmacokinetics during the M11-849 Biodistribution Study

ABT-806i pharmacokinetics were similar between Cohorts 1 and 2 (Table 5 and Figure 3). The apparently shorter observed half-life and higher clearance of ABT-806i compared to ABT-806 (Figure 3) may be due to a shorter pharmacokinetic sampling period for ABT-806i, although nonlinear pharmacokinetics between the low ABT-806i and high ABT-806 doses cannot be completely excluded. The biodistribution images did not indicate any saturable normal tissue compartment, which would be seen if a nonlinear PK at low protein dose did exist. The ABT-806 pharmacokinetics were consistent with results in the previous Phase 1 dose escalation study with ABT-806. No ADA was detected post-ABT-806 administration in this study.

Toxicity within M11-849 study

No patient in the M11-849 study experienced an adverse event related to ABT-806i. Two patients (33%) in Cohort 1 of M11-849 study each experienced an unrelated adverse event of Grade 1 upper respiratory tract infection and Grade 2 dyspnoea. Ten patients (83%) in Cohort 2 experienced toxicity. One patient in Cohort 2 (8%) experienced Grade 1 rash that the investigator considered possibly related to ABT-806 but no toxicities related to ABT-806i were reported. Toxicity occurring in \geq 2 patients in Cohort 2 included decreased appetite, fatigue (4 patients each, 33.3%), constipation, nausea, upper respiratory tract infection (3 patients each, 25%), arthralgia, dizziness, and productive cough (2 patients each, 17%) that were unrelated. Two of 10 patients each experienced 2 Grade 3 events; the remaining 8 patients experienced Grade 1 or 2 events.

Tumor uptake and Dosimetry of ABT-806i

ABT-806i tumor uptake was quantitatively evaluated for patients in Cohorts 1 and 2 (n=18). Tumor uptake varied greatly between patients, and ranged from 0.009-5.837% injected dose for a reference lesion. The only non-tumor region that showed specific ABT-806i uptake was

in patient 1, who had uptake in an area of previous lung radiotherapy, most likely related to enhanced permeability into the area of inflammation. No clear pattern between ABT-806i uptake and EGFR staining in archival tissue was seen (Table 2), although the median interval between tissue acquisition and enrolment on study was 1.5 years (range, 0.3-6.5 years), and archived samples tested were typically from previously removed primary tumors. Repeated treatment with ABT-806 had little effect on ABT-806i tumor accumulation irrespective of ABT-806 dose.

On a per target lesion basis, there were 12 lesions in Cohort 1 of which 7 lesions demonstrated qualitative uptake of ABT-806i, , 2 lesions with mild uptake score of 2; 3 lesions with moderate uptake score of 3; and 2 lesions with marked uptake score of 4. There was 1 target lesion with no uptake (score 0), 1 lesion with equivocal uptake, and 1 lesion with non-evaluable uptake. In Cohort 2, there were 34 lesions, of which 21 lesions had uptake of ABT-806i, including 14 lesions with moderate uptake score of 3, and 1 lesion with marked uptake score of 4. Importantly, of the 13 patients who had more than one evaluable lesion, 4 patients (31%) showed marked inter-tumoral variation of uptake of ABT-806i in target lesions. The patient who had a partial response had high (score 3) uptake of ABT-806i (Figure 4).

The normal tissue dosimetry of ABT-806i is shown in Supplementary Table 1. The mean effective dose of ABT-806i was calculated to be 0.137 mSv/MBq for males and 0.183 mSv/MBq for females.

Discussion

The advent of next-generation antibodies like ABT-806 represents the next wave of EGFRtargeting antibodies with enhanced tumor specificity and minimal cutaneous toxicities (1,7,12,13). ABT-806 has been shown to be well-tolerated and to have little of the cutaneous and other toxicities seen with other anti-EGFR antibodies (12). Our current study confirmed this profile with only 1 patient (6%) experiencing a Grade 1 rash that was possibly related to ABT-806. Like other anti-EGFR antibodies that have shown some activity in head and neck cancer (18), there was one patient with head and neck cancer who had a prolonged objective response.

Administration of ABT-806i was safe and well-tolerated, with no drug-related toxicities. The lung is the organ that had the highest dosimetry for ABT-806i, although the dose received from a tracer infusion of ABT-806i is well within acceptable levels, and consistent with other ¹¹¹Inlabeled antibodies (15,19). In comparison to imaging of wtEGFR, using ¹¹¹In-225 (20) and ⁸⁹Zrcetuximab (21), ABT-806i showed less liver uptake on imaging, with dosimetry and linear pharmacokinetics at low protein doses consistent with lack of normal tissue "sink" as well as tumor-specific EGFR imaging. This imaging illustrated several key observations. Firstly, marked inter-tumoral heterogeneity exists in nearly a third of patients between different metastatic sites. Secondly, it is not possible to predict this inter-tumoral heterogeneity with EGFR testing using archival tissue (or likely even with prospective tissue collection from single tumor sites). Lastly, the observation that the one responder in this study also had the higher ABT-806i uptake suggests a possible use to enrich for patients who will respond to anti-EGFR treatment. Clearly, our results require further validation given the relatively small study size and the heterogeneous patient population. However, the possibility that prospective and contemporaneous assessment of antigen target expression using radiolabeled tracers may impact on patient selection for antibody-based therapeutics has been shown by other trials, including T-DM1 (22) and may have an important role in drug development (14,23). Arguably, ABT-806i imaging could present way to enrich for responders for therapy with ABT-414, which comprised of ABT-806 conjugated to monomethyl auristatin F (MMAF)(24). Using selection on EGFR amplification in archival tissue, adding ABT-

414 to standard post-operative chemo-radiation did not improve survival in newly diagnosed patients (25) but there was a trend, though not statistically significant, toward survival benefit when combining ABT-414 with temozolomide in patients with recurrent GBM (26). In both studies, the need for better patient selection for highly targeted drugs like ABT-414 was emphasized (25,26) and imaging with ABT-806i could represent one way to do so.

Conclusion

In conclusion, this study has established the safety and feasibility of molecular imaging of intra-tumoural EGFR expression using ABT-806i scanning and illustrates possible roles in the development of antibody-based therapeutics.

Disclosures

Conflict of Interest Disclosures

HKG: received honoraria, travel support and/or research funding from AbbVie, BMS, Eisai, EMD Serono and MSD.

MB: served on advisory boards and received honoraria from Roche, Amgen and Sirtex

BS: served on advisory boards and received honoraria from Pfizer, Merck, Roche-Genetech, AstraZeneca, Bristol-Myers Squibb and Novartis

STL: Nil

KDH, WM, JF, PA, GF, HX, EBR and RH: Employees of AbbVie and may own stock

YZ: Previous AbbVie employee and may own stock

MC: Employee of Bristol-Myers Squibb and may own stock

AMS: Received research funding and travel support from AbbVie; received research funding from Daiichi-Sankyo, Merck, Telix, Avid, Celgene, ITM; is a consultant and has stock in Life Science Pharmaceuticals

Financial support: Abbvie provided financial support for this study (M11-849, NCT01472003) and participated in the study design, conduct, analysis and interpretation of data, as well as the writing, review and approval of the manuscript. All authors were involved in the data gathering, analysis, review, and interpretation.

Acknowledgements

We thank Mrinal Shah, PhD, for medical writing support, and Thomas Merdan, PhD, for technical support and assistance (both of Abbvie).

Key Points

- 1) We confirmed the safety of ABT-806i and acceptable radiation dosimetry
- We demonstrate the ability to successfully obtain real-time imaging of EGFR conformational expression in tumors
- Our data provides support for the use of theranostics to guide patient selection for EGFRtargeting therapies

References

1. Gan HK, Burgess AW, Clayton AHA, Scott AM. Targeting of a conformationally exposed, tumor-specific epitope of EGFR as a strategy for cancer therapy. *Cancer Res.* 2012;72:2924-2930.

2. Dhomen NS, Mariadason J, Tebbutt N, Scott AM. Therapeutic targeting of the epidermal growth factor receptor in human cancer. *Crit RevOncog.* 2012;17:31-50.

3. Robert C, Soria J-C, Spatz A, et al. Cutaneous side-effects of kinase inhibitors and blocking antibodies. *Lancet Oncol.* 2005;6:491-500.

4. Garrett TPJ, Burgess AW, Gan HK, et al. Antibodies specifically targeting a locally misfolded region of tumor associated EGFR. *Proc Nat Acad Sci (USA).* 2009;106:5082-5087.

5. Johns TG, Adams TE, Cochran JR, et al. Identification of the Epitope for the Epidermal Growth Factor Receptor-specific Monoclonal Antibody 806 Reveals That It Preferentially Recognizes an Untethered Form of the Receptor. *J Biol Chem.* 2004;279:30375-30384.

6. Jungbluth AA, Stockert E, Huang HJ, et al. A monoclonal antibody recognizing human cancers with amplification/overexpression of the human epidermal growth factor receptor. *Proc Nat Acad Sci (USA).* 2003;100:639-644.

7. Reilly EB, Phillips AC, Buchanan FG, et al. Characterization of ABT-806, a Humanized Tumor-Specific Anti-EGFR Monoclonal Antibody. *Mol Cancer Ther.* 2015;14:1141-1151.

8. Perera RM, Narita Y, Furnari FB, et al. Treatment of human tumor xenografts with monoclonal antibody 806 in combination with a prototypical epidermal growth factor receptor-specific antibody generates enhanced antitumor activity. *Clin Cancer Res.* 2005;11:6390-6399.

9. Gan HK, Lappas M, Cao DX, Cvrljevdic A, Scott AM, Johns TG. Targeting a unique EGFR epitope with monoclonal antibody 806 activates NF-kappaB and initiates tumour vascular normalization. *J Cell Molec Med.* 2009;13:3993-4001.

10. Luwor RB, Johns TG, Murone C, et al. Monoclonal antibody 806 inhibits the growth of tumor xenografts expressing either the de2-7 or amplified epidermal growth factor receptor (EGFR) but not wild-type EGFR. *Cancer Res.* 2001;61:5355-5361.

11. Cokgor I, Akabani G, Kuan CT, et al. Phase I trial results of iodine-131-labeled antitenascin monoclonal antibody 81C6 treatment of patients with newly diagnosed malignant gliomas. *J Clinl Oncol.* 2000;18:3862-3872.

12. Cleary JM, Reardon DA, Azad N, et al. A phase 1 study of ABT-806 in subjects with advanced solid tumors. *Invest New Drugs*. 2015;33:671-678.

13. Scott AM, Lee F-T, Tebbutt N, et al. A phase I clinical trial with monoclonal antibody ch806 targeting transitional state and mutant epidermal growth factor receptors. *Proc Nat Acad Sci (USA).* 2007;104:4071-4076.

14. Ciprotti M, Tebbutt NC, Lee FT, et al. Phase I Imaging and Pharmacodynamic Trial of CS-1008 in Patients With Metastatic Colorectal Cancer. *J Clin Oncol.* 2015;33:2609-2616.

15. Vallabhajosula S, Kuji I, Hamacher KA, et al. Pharmacokinetics and biodistribution of 111In- and 177Lu-labeled J591 antibody specific for prostate-specific membrane antigen: prediction of 90Y-J591 radiation dosimetry based on ¹¹¹In or ¹⁷⁷Lu? *J Nucl Med.* 2005;46:634-641.

16. Gan HK, Reardon DA, Lassman AB, et al. Safety, Pharmacokinetics and Antitumor Response of ABT-414 as Monotherapy or as Combination Therapy with Temozolomide (TMZ) in Patients with Glioblastoma. *Neuro-Oncol.* 2016; 2018;20:838-847.

17. Herbertson RA, Tebbutt NC, Lee FT, et al. Targeted chemoradiation in metastatic colorectal cancer: a phase I trial of ¹³¹I-huA33 with concurrent capecitabine. *J Nucl Med.* 2014;55:534-539.

18. Vermorken JB, Trigo J, Hitt R, et al. Open-label, uncontrolled, multicenter phase II study to evaluate the efficacy and toxicity of cetuximab as a single agent in patients with recurrent and/or metastatic squamous cell carcinoma of the head and neck who failed to respond to platinum-based therapy. *J Clin Oncol.* 2007;25:2171-2177.

19. Fisher DR, Shen S, Meredith RF. MIRD dose estimate report No. 20: radiation absorbed-dose estimates for ¹¹¹In- and ⁹⁰Y-ibritumomab tiuxetan. *J Nucl Med.* 2009;50:644-652.

20. Divgi CR, Welt S, Kris M, et al. Phase I and imaging trial of indium 111-labeled anti-epidermal growth factor receptor monoclonal antibody 225 in patients with squamous cell lung carcinoma. *J Natl Cancer Inst.* 1991;83:97-104.

21. Menke-van der Houven van Oordt CW, Gootjes EC, Huisman MC, et al. 89Zr-cetuximab PET imaging in patients with advanced colorectal cancer. *Oncotarget.* 2015;6:30384-30393.

22. Gebhart G, Lamberts LE, Wimana Z, et al. Molecular imaging as a tool to investigate heterogeneity of advanced HER2-positive breast cancer and to predict patient outcome under trastuzumab emtansine (T-DM1): the ZEPHIR trial. *Ann Oncol.* 2016;27:619-624.

23. de Vries EG, de Jong S, Gietema JA. Molecular Imaging As a Tool for Drug Development and Trial Design. *J Clin Oncol.* 2015;33:2585-2587.

24. Phillips AC, Boghaert ER, Vaidya KS, et al. ABT-414, an antibody–drug conjugate targeting a tumor-selective EGFR epitope. *Mol Cancer Ther.* 2016;15:661-669.

25. Lassman AB, Pugh SL, Wang TJC, et al. Depatuxizumab Mafodotin (ABT-414) in EGFR-amplified Newly Diagnosed GBM: a Randomized, Double-blind, Phase III, International Clinical Trial. *Annual Meeting of the Society for Neuro-Oncology*. Phoenix, AZ, USA; 2019.

26. van den Bent M, Eoli M, Sepulveda JM, et al. INTELLANCE 2/EORTC 1410 randomized phase II study of Depatux-M alone and with temozolomide vs temozolomide or lomustine in recurrent EGFRamplified glioblastoma. *Neuro Oncol.* 2020 doi: 10.1093/neuonc/noaa115. .

Figures

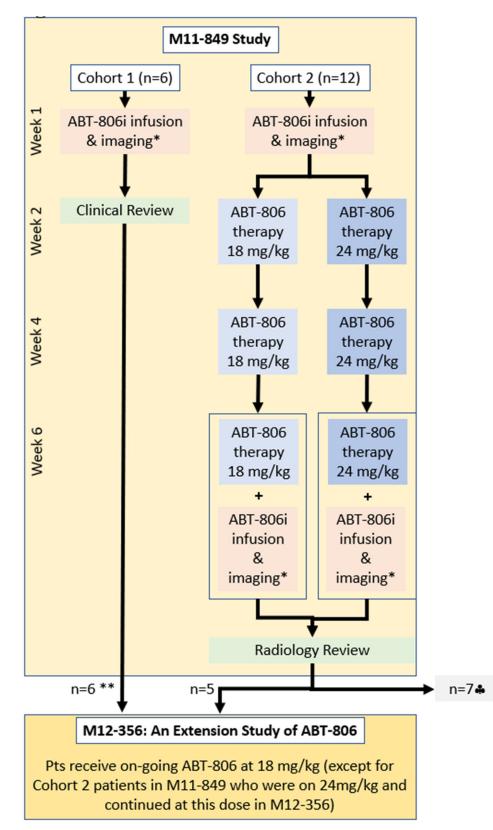


Figure 1. Patient Flow within M11-849 study and subsequently for eligible patient, from the M11-849 study into the M12-356 extension study

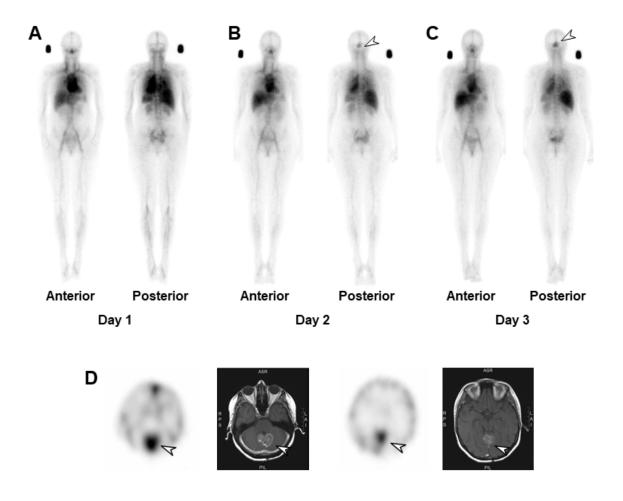


Figure 2. ABT-806i biodistribution and SPECT/CT images of a patient with high grade glioma. (**A**) Anterior and posterior whole body planar images at day 1 (standard located above right shoulder for dosimetry); (**B**) whole body planar images at day 2; (**C**) whole body planar images at day 3 in patient 3. Rapid uptake of ¹¹¹In-ABT-806i in known GBM (arrow) is identified by day 2 and increases by day 3. (**D**) SPECT/MRI images at two planes showing the high uptake of ¹¹¹In-ABT-806i in the GBM (arrow) in the posterior fossa. A focus of ¹¹¹In-ABT-806i uptake in the anterior venous sinus is due to blood pool activity.

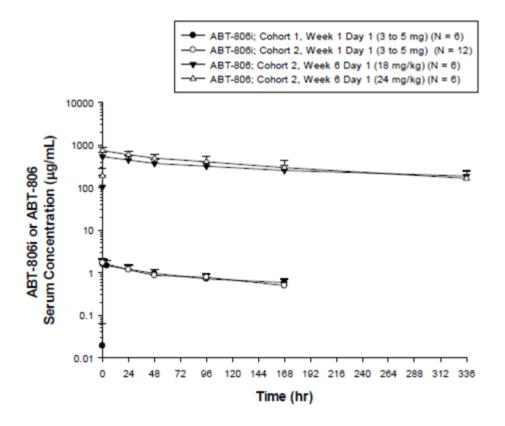


Figure 3. Serum concentration-time profiles of ABT-806i (Week 1, Day 1) or ABT-806 (Week 6, Day 1) following IV administration. Mean + standard deviation (SD) is shown on a log-linear scale.

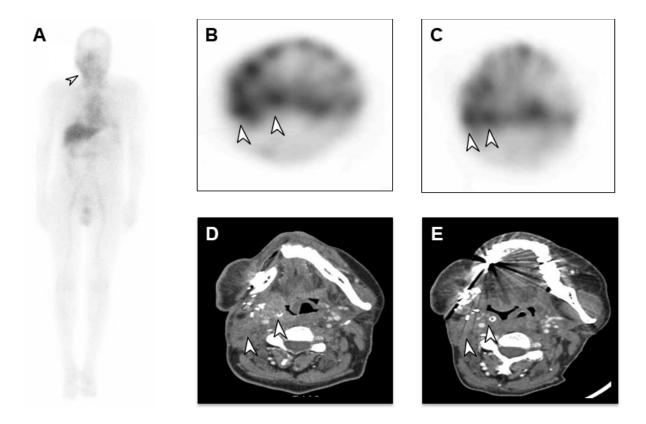


Figure 4. ABT-806i biodistribution and SPECT/CT images of a patient with squamous cell carcinoma of the head and neck. (A) Whole body planar image of ¹¹¹In-ABT-806i biodistribution at day 8 in patient 8. Arrow shows localization in tumor area in the right neck. (B) Week 1 SPECT image of ¹¹¹In-ABT-806i uptake in right parapharyngeal lesion and right cervical node (arrows), with uptake increased compared to normal tissue. (C) Week 16 SPECT image of ¹¹¹In-ABT-806i uptake in right parapharyngeal lesion and right cervical node (arrows), which appear smaller than week 1 images. (D) CT at baseline showing tumor in the right parapharyngeal region and right cervical node (arrows), which also showed ¹¹¹In-ABT-806i uptake. (E) CT at week 16 restaging, showing reduction in size of right parapharyngeal lesion and right cervical node (arrows), assessed as RECIST partial response.

Characteristics, N (%)		All patients N=18
Gender	Male:Female	11:7
Age (years)	Median	57
	Range	37-75
Ethnicity	Caucasian	17 (92)
-	Asian	1 (8)
Tumor type	Head and neck	6 (33)
	Colorectal	3 (17)
	Non-small cell lung cancer	2 (11)
	Glioma	2 (11)
	Other*	5 (28)
ECOG performance	0	5 (28)
status	1	13 (72)

Table 1. Patient demographics.

*Includes adrenal, cervical, cholangiocarcinoma, transitional cell carcinoma of bladder, thymoma

ECOG, Eastern Cooperative Oncology Group.

Patient	Cohort	Cancer type	EGFR H- scoreBest response on M11-849 study		Best response on M12-326 study	
1	1	Colon	0	Clinically stable	PD	
2	1	NSCLC	3	Clinically Stable	PD	
3	1	HGG	50	Clinically Stable	PD	
4	1	H&N	12	Clinically Stable	PD	
5	1	HGG	295	Clinically Stable	PD	
6	1	Adrenal	0	Clinically Stable	SD	
7	2	H&N	80	PD	N/A*	
8	2	H&N	235	SD	PR	
9	2	Thymoma	115	PD	N/A*	
10	2	Cholangio- carcinoma	55	PD	N/A*	
11	2	NSCLC	278	SD	PD	
12	2	Colon	0	SD	PD	
13	2	H&N	170	SD	PD	
14	2	Cervical	13	PD	N/A*	
15	2	Bladder	228	PD	N/A*	
16	2	H&N	67	PD	N/A*	
17	2	Colon	0	SD	PD	
18	2	H&N	32	PD	N/A*	

Table 2. Individual patient data.

NSCLC, non-small cell lung cancer; HGG, high grade glioma; H&N, head and neck; N/A Not applicable; PD, progressive disease; SD, stable disease; PR, partial response.

* Patients who had progressive disease at the end of M11-849 study were not eligible for enrolment on the M12-326 extension study

Table 3. Peak uptake of ¹¹¹In-ABT-806i in target tumor lesion in Cohort 1 patients, calculated from SPECT Images

Subject	Site of reference lesion	Tumor maximal uptake (%ID)
1	left lung	0.023%
2	right lung	0.035%
3	cerebellum	N/A
4	left neck	0.029%
5	left frontal lobe	N/A
6	left adrenal	0.016%

Ist Infusion ¹¹¹In-ABT-806i

N/A - no CT data available

Table 4. Peak uptake of ¹¹¹In-ABT-806i in target tumor lesion in Cohort 2 patients,calculated from SPECT/CT Images

		Ist Infusion ¹¹¹ In-ABT-806i	2nd Infusion ¹¹¹ In-ABT-806i Tumor maximal uptake (%ID)		
Subject	Site of reference lesion	Tumor maximal uptake (%ID)			
7	right hilum	1.290%	1.321%		
8	right pharyngeal	0.078%	0.092%		
9	left lung	0.043%	0.041%		
10	right peritoneum	0.265%	0.264%		
11	subcarinal node	N/A	N/A		
12	right lung	0.038%	0.037%		
13	hypopharynx	0.030%	0.024%		
14	right hilum	0.067%	0.057%		
15	right lung	0.064%	0.087%		
16	left lung	0.050%	0.051%		
17	right lung	5.837%	6.679%		
18	left lung	0.009%	0.009%		

N/A - target lesion not assessable for quantitative dosimetry

Cohort (ABT-806 Dose Group)	ABT- 806i (mg)	ABT- 806 (mg/kg)	N	T _{max} (hr)	C _{max} (µg/ml)	C _{max} /Dose (μg/ml)/(mg/kg)	T _{1/2} (days)*	AUC (mg*h/ml)†	AUC/dose (mg*h/ml)/(mg/kg) †	CL (ml/h)
					ABT-	806i (Week 1, Day 1	1)			
1	3-5	N/A	6	0.41±0.46	1.61±0.50	23.4±5.2	6.0±1.5	0.274±0.080	4.07±1.10	19.4±6.6
2 (18mg/kg)	3-5	N/A	6	0.11±0.04	1.62±0.32	22.2±4.7	5.6±1.5	0.243±0.062	3.36±1.03	21.3±6.1
2 (24 mg/kg)	3-5	N/A	6	0.15±0.10	1.75±0.39	22.7±5.4	5.3±2.4	0.261±0.113	3.49±1.90	24.5±15.9
2 (18 and 24 mg/kg)	3-5	N/A	12	0.13±0.08	1.69±0.35	22.4±4.8	5.4±1.9	0.252±0.088	3.42±1.46	22.9±11.6
	ABT-806 + ABT-806i (Week 6, Day 1)									
2 (18 mg/kg)	3-5	N/A	6	2.25±0.49	531±136	29.5±7.6	14.5±4.3	99.8±27.9	5.54+1.55	13.3±4.6
2 (24 mg/kg)	3-5	N/A	6	2.16±0.61	742±128	30.9±5.3	8.6±2.3	116.2±38.6	4.84±1.61	14.8±5.1

Table 5. Pharmacokinetic parameters of ABT-806i and ABT-806

 T_{max} time to maximum concentration in serum; C_{max} maximum observed plasma concentration; $t_{1/2}$ terminal elimination half-life; AUC area under the curve; CL clearance; N/A not applicable. * Harmonic mean and pseudo-standard deviation

 \dagger AUC_{\scriptscriptstyle \infty} for Week 1, Day 1 and AUC_{\scriptscriptstyle TAU} for Week 6, Day 1

Female (n=4) Male (n=2) Organ Mean ED Mean ED SD SD (mSv/MBq) (mSv/MBq) Adrenals 0.0008 0.0005 0.0005 0.0002 Bladder 0.0103 0.0064 0.0145 0.0048 Brain 0.0002 0.0001 0.0002 0.0001 Breast 0.0032 0.0018 0.0026 0.0001 Gall bladder 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 0.0000 Heart 0.0077 Kidney 0.0018 0.0010 0.0092 Large bowel (upper) 0.0007 0.0004 0.0004 0.0002 Large bowel (lower) 0.0102 0.0059 0.0105 0.0018 0.0261 0.0152 0.0167 0.0052 Liver Lungs 0.0264 0.0165 0.0201 0.0010 Muscle 0.0003 0.0002 0.0002 0.0001 Osteogenic cells 0.0015 0.0009 0.0014 0.0002 Ovaries 0.0195 0.0111 _ _ 0.0008 0.0005 0.0002 Pancreas 0.0005 Red marrow 0.0121 0.0068 0.0134 0.0012 Skin 0.0004 0.0002 0.0003 0.0001 Small bowel 0.0010 0.0005 0.0006 0.0002 Spleen 0.0014 0.0009 0.0011 0.0002 0.0233 Stomach 0.0134 0.0184 0.0028 Testes 0.0000 0.0000 --0.0003 Thymus 0.0005 0.0003 0.0001 Thyroid 0.0052 0.0030 0.0073 0.0004 0.0005 Uterus 0.0003 - $MEAN \pm SD$ 0.183 0.021 0.137 0.008

Supplementary Table 1. Mean organ effective dose (ED) in mSv/MBq for all patients in Cohort 1.

SD, standard deviation