Journal of Nuclear Medicine, published on December 4, 2020 as doi:10.2967/jnumed.120.256271

State-of-the-Art: FAPI PET/CT-Will it end the hegemony of FDG in oncology?

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**Short running title:** FAPI Review

### **ABSTRACT**

For over 40 years, FDG has been the dominant PET tracer in neurology, cardiology, inflammatory diseases and, most particularly, oncology. Combined with the ability to perform whole-body scanning, FDG has revolutionized the evaluation of cancer and has stifled the adoption of other tracers, except in situations where low avidity or high background activity limits diagnostic performance. The strength of FDG has generally been its ability to detect disease in the absence of structural abnormality, thereby enhancing diagnostic sensitivity, but its simultaneous weakness has been a lack of specificity due to diverse pathologies with enhanced glycolysis. Radiotracers that leverage other hallmarks of cancer or specific cell-surface targets are gradually finding a niche in the diagnostic armamentarium. However, none have had sufficient sensitivity to realistically compete with FDG for evaluation of the broad spectrum of malignancies. Perhaps, this situation is about to change with development of a class of tracers targeting fibroblast activation protein (FAP) that have low uptake in almost all normal tissues but high uptake in most cancer types. In this review, the development and exciting preliminary clinical data relating to various FAP-specific small-molecule inhibitor (FAPI) tracers in oncology, will be discussed along with potential non-oncological applications.

Key words: PET, theranostics, fibroblast activation protein, oncology, radiochemistry

### INTRODUCTION

Inspired by Sokoloff's autoradiographic assessment of cerebral glucose metabolism, David Kuhl approached the Brookhaven National Laboratory to develop a positron-emitting tracer of deoxyglucose for his new tomographic scanner. As a consequence, <sup>18</sup>F-fluorodeoxyglucose (FDG), which Dr Henry Wagner once described as being "the molecule of the 20<sup>th</sup> century", was conceived. Roles in epilepsy (1), neurodegeneration (2) and ischemic heart disease (3) were initially described, but it was in oncology that FDG has most profoundly impacted the current practice of molecular imaging. This was accelerated by the development of whole-body scanning capability (4) and particularly PET/CT (5).

In the developed world, PET/CT is now widely used to diagnose, stage and monitor treatment response with more accurate diagnosis sparing inappropriate utilization of expensive and toxic therapies (6). FDG is by far the dominant tracer in oncology, despite increasing use of more specific tracers in certain malignancies, such as  $^{68}$ Ga-DOTA-octreotate in neuroendocrine neoplasia (7) and various prostate-specific membrane antigen (PSMA) tracers for prostate cancer evaluation (8,9). These have become particularly important as part of theranostic paradigms including peptide receptor radionuclide therapy (10) and PSMA-based radioligand therapy (11), respectively. Similarly, high uptake of FDG in the brain has opened the way for amino acid analogs in the evaluation of brain tumors (12).

Despite the attraction of such tracers in advancing precision medicine and replacing standard nuclear medicine procedures (13), none have the broadly-based appeal of FDG across the spectrum of malignancies in which it currently holds hegemony. For a tracer to compete, it would need to vie with FDG for sensitivity and surpass it with respect to specificity, or alternatively offer the opportunity for theranostic application. Although data are preliminary, emerging evidence suggests that small-molecule fibroblast activation protein inhibitor (FAPI) tracers may be candidates for widespread oncological application. In this review, the development and performance of FAPI-PET/CT will be discussed along with possible therapeutic

use. As with FDG, FAPI agents may also find an important diagnostic niche in non-oncological conditions.

#### The role of CAFs in cancer

While neoplastic cells have traditionally been thought to be primary determinants of cancer behavior, non-malignant cells within the tumor microenvironment (TME) are increasingly recognized as modulators of both tumor progression and therapeutic response. Accordingly, the stromal compartment, including immune cells, vascular elements and cancerassociated fibroblasts (CAFs), has been a focus of investigation. Key functions of CAFs are deposition and remodeling of extracellular matrix as part of a fibrotic reaction, which, when prominent in tumors such as pancreatic adenocarcinoma (14), is termed desmoplasia. However, CAFs are commonly found in solid tumor types without desmoplasia and therefore represent an attractive diagnostic and therapeutic target (15). Indeed, there is a concerted international effort to develop therapeutic agents targeting CAFs (16). Recently, it has become clear that the complex interplay of CAFs and immune cells can both suppress and accelerate tumor growth. This is likely to be driven by the significant biological heterogeneity that has been identified in CAF subtypes both within and between cancers (17,18). The origins of CAFs are postulated to include activation of quiescent tissue fibroblasts, trans-differentiation of other stromal or epithelial cells, recruitment of circulating mesenchymal stem cells, or by differentiation from tissue-resident stem cells (19). Any or all of these mechanisms may co-exist but how this ontogeny influences their biological behavior remains to be fully elucidated. Epigenetic processes are thought to be involved in the development of pro-tumorogenic CAFs (20).

One of the major impacts of CAFs is the development of an immunosuppressive TME (21) by processes including desmoplastic stroma excluding T cells from tumor deposits and secretion of various chemokines that recruit myeloid-derived suppressor cells and regulatory T cells. Chief among these is transforming growth factor  $\beta$  (TGF $\beta$ ) (22). The desmoplasia initiated by TGF $\beta$  has recently been shown to be mediated through downregulation of signaling through the type I interferon receptor (23). CAFs also interact to alter the behavior or epithelial cancer

cells through cell-cell interactions mediated by cadherins, cytokine signaling and secretion of exosomes (24). Further, by modifying the extra-cellular matrix CAFs are permissive for cancer invasion and tumor cell survival (25,26).

The biology and nomenclature of subsets of CAFs in different cancer types is still evolving (18) but recent advances, including single cell technologies, have helped to define specialized subsets of CAFs (21). In pancreatic cancer, for instance, CAFs can be classified as myofibroblasts (myCAFs), inflammatory (iCAFs) and antigen presenting (apCAFs) (27).

# Fibroblast activation protein (FAP) and cancer biology

FAP is a type II transmembrane glycoprotein belonging to the dipeptidyl peptidase IV (DPP-IV) like family of post-prolyl cleaving serine proteases that are expressed in dimeric form on the surface of CAFs (28) and, as such, is an attractive diagnostic and therapeutic target. It plays a complex role in modulating the extra-cellular matrix (29). FAP differs from other members of the dipeptidyl peptidase family in acting as a gelatinase on denatured collagen, especially when cleaved by matrix metallopeptidases (30). Although the role of specific subtypes of CAFs in promoting or suppressing tumorigenesis remains controversial, it is becoming increasing clear that CAFs with high FAP expression are associated with an adverse prognosis by promoting invasion, angiogenesis, micro-environmental immune suppression and metastasis (15). At least in part, this may reflect production of an immunosuppressive TME (31). Interestingly, in addition to expression on CAFs, FAP is also expressed on tumor-associated macrophages, which are important mediators of immune suppression (32). FAP has been shown to be important in recruitment of immunosuppressive myeloid-derived suppressor cells in cholangiocarcinoma (33). One of the potential effector molecules secreted by FAP-expressing CAFs is chemokine (C-X-C motif) ligand 12 (CXCL12), which leads to polarization of macrophages to the immunosuppressive M2 phenotype (34) and also promotes angiogenesis in both breast (35) and colorectal carcinoma (36).

There is still much to learn about the biological and clinical significance of FAP expression in different tumor types. For example, in the brain, where CAFs do not exist in the microenvironment, FAP overexpression has been found in both non-malignant mesenchymal cells in the stroma and in transformed cells but had no association with prognosis (37). FAP expression was particularly associated with dysplastic blood vessels and highest in the mesenchymal subtype of glioblastoma. In a subsequent study of 13 patients with glioblastoma (38), no correlation was found the apparent diffusion co-efficient (ADC), which is generally associated with variation in prognosis. There was, however, significant intra-tumoral heterogeneity with a relationship to better perfused regions, possibly reflecting FAP expression in neovasculature (39). Additionally, FAP expression is also seen in certain sarcomas and some benign mesenchymal tumors including desmoid, giant cell tumors and chondroblastoma (40), suggesting that FAP expression is not limited to CAFs.

Besides the opportunity that FAP provides as a diagnostic target, its high expression in several cancers with a poor response to current therapeutic approaches, including pancreatic (41) and ovarian carcinoma (42,43), makes it also an attractive therapeutic target. Even in more common and generally chemo-responsive tumors like colorectal cancer, high FAP expression has also been found to be associated with poor prognosis (36,44). At least in preclinical models, use of CAR T cells directed against FAP-expressing CAFs has been a successful therapeutic strategy (45). FAP is also being investigated as a target for antibody-drug conjugates (46). Accordingly, companion diagnostics would be valuable for selecting patients for such treatments or even for selecting patients for radioligand therapy, as discussed below. Importantly, FAP appears to be overexpressed across a diverse range of cancers (Fig. 1).

### FAP as a target for imaging cancer

Early attempts to develop FAP imaging probes involved radiolabeled antibodies. The F19 probe that had been used for immunohistochemical identification of FAP expression (47) was labeled with <sup>131</sup>I and evaluated in patients with metastatic colorectal cancer (48). However, the long circulation time of intact antibodies poses logistic challenges and stimulated investigation

of smaller molecules that may be suitable for imaging. It had long been known that novel dipeptides containing the boronic acid analogue of proline could form potent, yet non-selective inhibitors of the DPP family of peptidases (49). It was the introduction of the structurally similar 2-cyanopyrollidine that led to the discovery of FAP inhibitors with increased selectivity and nanomolar affinity (50,51). By building on this work, the Heidelberg Group based their first FAP targeted tracers, FAPI-01 and FAPI-02, radiolabeled with <sup>131</sup>I, and <sup>68</sup>Ga through chelation to DOTA, respectively (52).

Initial reports primarily involved preclinical validation (*52,53*). Compared to FAPI-01, FAPI-02 was found to specifically bind to FAP-expressing cells and have greater cellular retention. Internalization was confirmed using a fluorescent FAPI-02 with colocalization with endosomes. Small animal PET demonstrated high and specific tumor uptake. Encouraged by these results, preliminary human evaluation was performed in 3 patients with metastatic breast, lung and pancreatic cancers, respectively. All three patients had visualization of primary and metastatic disease but declining tumor uptake over 3 hours. The primary route of excretion was renal with little parenchymal retention. A series of variants based on FAPI-02 were designed to increase cellular retention. The most promising agent in preclinical models, FAPI-04, was tested in 2 women with metastatic breast cancer, demonstrating encouraging tumor visualization and favorable biodistribution (*54*).

A further report expanded these preliminary clinical results in 50 patients equally split between <sup>68</sup>Ga-FAPI-02 or <sup>68</sup>Ga-FAPI-04 PET/CT (*55*). As suggested by the earlier reports, there was significantly greater washout of <sup>68</sup>Ga-FAPI-02 from tumor deposits between 1 and 3 h compared to <sup>68</sup>Ga-FAPI-04 but both performed comparably at 1 h with clinically acceptable dosimetry. In 6 patients with contemporaneous FDG PET/CT available for direct correlation, comparable tumor SUV results were observed except in one patient with iodine-negative thyroid cancer in whom, FDG visualized several sites of disease not identified on FAPI PET/CT.

The Heidelberg experience with <sup>68</sup>Ga-FAPI-04 was subsequently expanded in 80 patients with a range of cancers (*56*). Of 28 cancer types evaluated, the highest uptake (average

SUVmax >12) was observed in carcinoma of unknown primary, sarcoma, cholangiocarcinoma, esophageal, breast, and lung cancer but most had significantly higher than blood pool uptake (Fig. 2). However, a case report from other group suggested that inflammatory conditions, such as IgG4-related disease, can also be positive (57). Similarly, another group evaluating <sup>68</sup>Ga-FAPI-04 in 68 patients with equivocal FDG PET/CT found a small number of false-positive results related to inflammatory conditions despite clarifying the extent of disease in the majority of patients (58). Another study evaluated 75 patients with FAPI-04, including 54 with FDG PET/CT correlation (59). Across 12 different cancer groupings, FAPI PET/CT identified more primary, nodal and distant metastatic sites than FDG PET/CT with a significantly higher SUV in lung, pancreatic and gastric cancer. The high tumor-to-background ratios provided by <sup>68</sup>Ga-FAPI has been leveraged to guide radiotherapy planning in 14 patients with the head and neck cancer (60), a site of complex anatomy and where muscular and brown fat activity can make FDG PET/CT difficult to interpret. Given that FAP seems to invoke an immune-excluded TME, FAPI imaging may potentially play a role in identifying patients who are less likely to respond to immune checkpoint inhibitor therapy (61).

The Heidelberg group continued to experiment with variations on FAPI-04 using chemical adaptations including alteration of lipophilicity and modification of the DOTA/linker attachment at the quinoline moiety to improve tumor retention (*53*). Based on cell culture and preclinical imaging, two lead compounds entered clinical evaluation in 8 patients. These were designated FAPI-21 and FAPI-46. While both had high tumor uptake, FAPI-21, inexplicably had higher uptake in the major salivary glands, mouth and thyroid, rendering <sup>68</sup>Ga-FAPI-46 the leading candidate for clinical evaluation. Formal dosimetry studies performed in 6 patients were favorable (*62*). The main off-target organs to receive radiation were, as expected, the urinary bladder and kidneys. This suggests that adequate hydration and frequent voiding would be protective in the therapeutic setting. Using a combination of scans with <sup>68</sup>Ga-FAPI-04 (n =16) and <sup>68</sup>Ga-FAPI-46 (n =6), promising results were reported in lower gastrointestinal tract tumors (*63*). Stage was changed in 50% of patients without prior treatment and identified significantly more metastases in those with already known metastasis and changed management in 17 of 21

evaluable patients. However, relatively few of these patients had been staged with FDG PET/CT. The study included 6 patients with anal cancer, which can be difficult to assess due to physiological FDG uptake in sphincteric muscle. Low bowel activity is a potential advantage of FAPI in delineating peritoneal disease (Fig. 3)

In a further advance, an <sup>18</sup>F-labeled agent, FAPI-74 was recently described that uses an aluminum-fluoride labeling method on a pre-cursor that can also be labeled as a cold-kit using <sup>68</sup>Ga (*64*). Evaluation in 10 patients with lung cancer revealed somewhat higher vascular visualization than FAPI-02/04 but high tumor uptake enabling radiotherapy planning. Radiosynthesis and preclinical evaluation of another fluorinated FAPI tracer, <sup>18</sup>FGlc-FAPI, has also been recently described (*65*). This agent had higher hepatobiliary clearance than <sup>68</sup>Ga-FAPI-04. There is currently limited information on the clinical performance of this tracer.

Low uptake of FAPI agents in the brain is a potential advantage over FDG for evaluation of both primary and secondary malignancies (Fig. 4). Recognition of cerebral metastasis (66) and leptomeningeal disease (67) have been reported in individual patients with adenocarcinoma of the lung. However, this may not necessarily be specific for malignancy with infectious etiologies like tuberculosis also demonstrating increased activity (68). In a series of patients with various primary brain tumors, <sup>68</sup>Ga-FAPI-02 or 04 uptake was observed in all IDH-wildtype glioblastomas and higher-grade IDH-mutant gliomas (38). In a preliminary evaluation, the potential application of FAPI PET/CT or PET/MRI for radiotherapy planning has been described (69).

Beside the diagnostic utility of FAPI agents, the potential ability to select patients for radionuclide therapy is an attractive prospect. Preclinical studies of <sup>177</sup>Lu-labeled internalizing antibodies directed against FAP have been tested in preclinical models and demonstrated to have efficacy (*70*). For agents with very high uptake but a short biological half-life, a radionuclide with a short physical half-life may be more appropriate than a long-lived radionuclide. With these factors in mind, preclinical evaluation of FAPI analogs was performed

using <sup>64</sup>Cu (*71*). The pairing of <sup>64</sup>Cu and <sup>67</sup>Cu represents an exciting theranostic combination. <sup>64</sup>Cu is a positron emitting radioisotope with a 12.7 h half-life, allowing centralized manufacturing and distribution of diagnostic tracers as well as the ability to perform prospective dosimetry (*72*), while <sup>67</sup>Cu is a beta-emitting radiometal with physical characteristics similar to <sup>177</sup>Lu but a shorter half-life. Compared to <sup>68</sup>Ga-FAPI-04, <sup>64</sup>Cu-FAPI-04 showed higher liver and intestinal activity. While it is known that the radiometal component of otherwise identical targeting agents can alter biodistribution (*73*), for copper, there is a possibility that relatively poor chelation by DOTA may increase free-<sup>64</sup>Cu uptake in the liver. For radiopeptides, alternative chelating agents can significantly reduce liver activity (*74*). However, unless related to free copper, altering clearance from rapid renal clearance to slower hepatobiliary excretion may be beneficial for therapeutic applications by increasing bioavailability of the radioligand.

# Therapeutic targeting of FAP in cancer

One of the first attempts to leverage high FAP expression as a therapeutic target was a trial in 26 patients, primarily with metastatic colorectal cancer, using an antibody called sibrotuzumab (75). Unfortunately, no objective responses were observed. Further development of antibodies against FAP identified by phage display, have been labeled with <sup>177</sup>Lu and tested in preclinical models (70). These agents demonstrated encouraging efficacy. Other approaches have included development of FAP-targeting prodrugs, vaccines and nanoparticles (76), although these have largely been limited to preclinical evaluation as yet.

The therapeutic application of FAPI agents has thus far been relatively limited. The original FAPI-04 report included a single patient treated with 2.9 GBq of <sup>90</sup>Y-FAPI-04 for metastatic breast cancer (*54*). Bremsstrahlung imaging post-treatment demonstrated significant tumor retention and the patient experienced a symptomatic improvement in bone pain, thereby demonstrating the theranostic potential of these agents.

## **FAP** imaging in other pathologies

There is a wide range of conditions that involve deposition of collagen. In normal tissue homeostasis, wound healing is the obvious example (77,78). However, excessive fibrotic reaction characterizes a number of pathological states including keloid formation in the skin, pulmonary fibrosis and asbestosis in the lungs, post-myocardial healing and restrictive pericarditis in the heart, interstitial nephritis in the kidney, and sclerosing cholangitis and cirrhosis in the liver, to name but a few. Additionally, increased FAP expression has be reported in synoviocytes in refractory rheumatoid arthritis (79). In pre-clinical models, an antibody targeting FAP, <sup>111</sup>In-28H1, had impressive uptake but the <sup>89</sup>Zn version for PET was less impressive due to higher bone uptake (80).

Benign processes characterized by fibrosis can, of course, co-exist with cancer or be caused by cancer treatments, particularly radiotherapy and bleomycin chemotherapy. For example, radiotherapy can induce radiation pneumonitis that can be identified by a geographic increase in uptake of FDG (81), which often precedes CT abnormality (82). In a preclinical model of bleomycin lung injury, FAP levels were increased and fibrosis correspondingly reduced by use of a FAP inhibitor (83). Non-malignant fibrosis of the mesentery and cardiac valves is a characteristic feature of carcinoid syndrome. Serotonin and bradykinins secreted by enterochromaffin-like (ECL) cells have been implicated in this process through stimulating fibroblasts (84) and, in the heart, TGF $\beta$ , which is also a key cytokine in cancer-associated desmoplasia, has been shown to be involved in the development of cardiac valvular thickening that characterizes carcinoid heart disease (85). Under the influence of TGF $\beta$ , FAP is also instrumental in remodeling in the peri-infarct zone following ischemic injury to the myocardium (86,87). The potential for FAPI PET to image this process has been demonstrated clinically (88,89).

Thus far, only clinical limited studies have been performed in benign fibrotic conditions using newer FAPI agents. Diffusely increased uptake in a cirrhotic liver but not in associated nodules, one of which was an hepatic adenoma on biopsy, has been reported (90). High uptake

in hepatic cirrhosis could potentially impair the detection of hepatocellular carcinoma (HCC) arising in this setting. However, in a recent series of 17 patients, which included 11 patients with intrahepatic HCC, all 15 identified lesions were positive on FAPI-04 PET/CT despite the SUVmean in the liver parenchyma in patients with cirrhosis being significantly increased compared with that in patients without cirrhosis (91).

IgG4-related disease seems to have high FAPI-avidity, which may be focal or diffuse in the pancreas and involve other organs, particularly the lungs, salivary glands and biliary tract (57,92,93). In the largest series published as yet, 26 patients were evaluated with both FDG and FAPI-04 with the latter demonstrating active disease in all cases with addition sites of involvement identified in 50% of cases (93). Activation of myofibroblasts by polarized CD4+ T lymphocytes is the likely explanation for this (94). Large vessel vasculitis is another potential application because of the rapid blood clearance and high uptake in vasculitis lesions (Fig. 5). The ability to differentiate vasculitis from active atherosclerotic plaques may, however, be limited with an autoradiographic using an <sup>125</sup>I-labeled FAPI agent demonstrating high binding to plaques in an animal model (95).

## **Future Directions**

There is a large number of ongoing or planned clinical trials using various FAPI tracers suitable for PET imaging, primarily in cancer (*96*). The availability of agents suitable for use in parts of the world where PET is not routinely available is potentially addressed by the development of FAPI agents labeled with <sup>99m</sup>Tc (*97*). <sup>99m</sup>Tc FAPI-34 has been tested in preclinical models and evaluated in one patient with ovarian cancer and another with pancreatic cancer with similar biodistribution in each case with prior <sup>68</sup>Ga-FAPI PET/CT and also post-treatment <sup>90</sup>Y-FAPI bremsstrahlung imaging. A further attraction of this agent is the potential for therapeutic use with <sup>188</sup>Re. This generator-produced beta-emitting radionuclide has a relatively short half-life (16.9 h), which may be advantageous given the clearance kinetics of FAPI-34, and an imageable gamma emission (155keV), allowing post-treatment dosimetry (*98*).

### Conclusions

Although preliminary, there are several reasons to be hopeful that FAPI agents will play an important role in the molecular imaging armamentarium. These include high expression across a wide range of cancer types including several with typically low FDG-avidity, low uptake in almost all normal tissues, including the brain and bowel, where high physiological uptake can obscure primary or metastatic disease, and the ability to provide prognostic information that might guide therapeutic options. Although it remains unclear whether targeting FAP will be an effective therapeutic strategy, the possibility of theranostics remains intriguing. As with FDG, the causes of "false-positive" cancer diagnoses open possibilities for a broader use of these agents, particularly in a range of fibrotic diseases that are difficult to monitor non-invasively.

#### REFERENCES

- 1. Kuhl DE, Engel J, Jr., Phelps ME, Kowell AP. Epileptic patterns of local cerebral metabolism and perfusion in man: investigation by emission computed tomography of 18Ffluorodeoxyglucose and 13N-ammonia. Trans Am Neurol Assoc. 1978;103:52-53.
- 2. Kuhl DE, Metter EJ, Riege WH, Phelps ME. Effects of human aging on patterns of local cerebral glucose utilization determined by the [18F]fluorodeoxyglucose method. J Cereb Blood Flow Metab. 1982;2:163-171.
- 3. Phelps ME, Schelbert HR, Hoffman EJ, Huang SC, Kuhl DE. Positron tomography of the heart. Prog Nucl Med. 1980;6:183-209.
- 4. Hoh CK, Schiepers C, Seltzer MA, et al. PET in oncology: will it replace the other modalities? Semin Nucl Med. 1997;27:94-106.
- 5. Beyer T, Townsend DW, Brun T, et al. A combined PET/CT scanner for clinical oncology. J Nucl Med. 2000;41:1369-1379.
- 6. Sullivan R, Peppercorn J, Sikora K, et al. Delivering affordable cancer care in highincome countries. Lancet Oncol. 2011;12:933-980.
- 7. Kwekkeboom DJ, Kam BL, van Essen M, et al. Somatostatin-receptor-based imaging and therapy of gastroenteropancreatic neuroendocrine tumors. Endocr Relat Cancer. 2010;17:R53-73.
- 8. Hofman MS, Lawrentschuk N, Francis RJ, et al. Prostate-specific membrane antigen PET-CT in patients with high-risk prostate cancer before curative-intent surgery or radiotherapy (proPSMA): a prospective, randomised, multicentre study. Lancet. 2020;395:1208-1216.
- 9. Fendler WP, Eiber M, Beheshti M, et al. Ga-PSMA PET/CT: Joint EANM and SNMMI procedure guideline for prostate cancer imaging: version 1.0. Eur J Nucl Med Mol Imaging. 2017;44:1014-1024.
- 10. Hicks RJ, Kwekkeboom DJ, Krenning E, et al. ENETS consensus guidelines for the standards of care in neuroendocrine neoplasia: Peptide receptor radionuclide therapy with radiolabeled somatostatin analogues. Neuroendocrinology. 2017;105(3):295-309.

- 11. Kratochwil C, Fendler WP, Eiber M, et al. EANM procedure guidelines for radionuclide therapy with 177Lu-labeled PSMA-ligands (177Lu-PSMA-RLT). Eur J Nucl Med Mol Imaging. 2019;46(12):2536-2544.
- 12. Galldiks N, Lohmann P, Albert NL, Tonn JC, Langen KJ. Current status of PET imaging in neuro-oncology. Neurooncol Adv. 2019;1(1):vdz010.
- 13. Hicks RJ, Hofman MS. Is there still a role for SPECT-CT in oncology in the PET-CT era? Nat Rev Clin Oncol. 2012;9:712-720.
- 14. Ho WJ, Jaffee EM, Zheng L. The tumour microenvironment in pancreatic cancer clinical challenges and opportunities. Nat Rev Clin Oncol. 2020;17:527-540.
- 15. Kalluri R. The biology and function of fibroblasts in cancer. Nat Rev Cancer. 2016;16:582-598.
- 16. Chen X, Song E. Turning foes to friends: targeting cancer-associated fibroblasts. Nat Rev Drug Discov. 2019;18:99-115.
- 17. Helms E, Onate MK, Sherman MH. Fibroblast heterogeneity in the pancreatic tumor microenvironment. Cancer Discov. 2020;10:648-656.
- 18. Sahai E, Astsaturov I, Cukierman E, et al. A framework for advancing our understanding of cancer-associated fibroblasts. Nat Rev Cancer. 2020;20:174-186.
- 19. Kobayashi H, Enomoto A, Woods SL, Burt AD, Takahashi M, Worthley DL. Cancer associated fibroblasts in gastrointestinal cancer. Nat Rev Gastroenterol Hepatol. 2019;16:282-295.
- 20. Fiori ME, Di Franco S, Villanova L, Bianca P, Stassi G, De Maria R. Cancer-associated fibroblasts as abettors of tumor progression at the crossroads of EMT and therapy resistance. Mol Cancer. 2019;18:70.
- 21. Kieffer Y, Hocine HR, Gentric G, et al. Single-cell analysis reveals fibroblast clusters linked to immunotherapy resistance in cancer. Cancer Discov. 2020;10:1330-1351.
- 22. Flavell RA, Sanjabi S, Wrzesinski SH, Licona-Limón P. The polarization of immune cells in the tumour environment by TGFbeta. Nat Rev Immunol. 2010;10:554-567.

- 23. Cho C, Mukherjee R, Peck AR, et al. Cancer-associated fibroblasts downregulate type I interferon receptor to stimulate intratumoral stromagenesis. Oncogene. 2020;39:61296137.
- 24. Ireland AS, Micinski AM, Kastner DW, et al. MYC Drives Temporal Evolution of Small Cell Lung Cancer Subtypes by Reprogramming Neuroendocrine Fate. Cancer Cell. 2020;38(1):60-78.e12.
- 25. Goetz JG, Minguet S, Navarro-Lérida I, et al. Biomechanical remodeling of the microenvironment by stromal caveolin-1 favors tumor invasion and metastasis. Cell. 2011;146:148-163.
- 26. Riching KM, Cox BL, Salick MR, et al. 3D collagen alignment limits protrusions to enhance breast cancer cell persistence. Biophys J. 2014;107:2546-2558.
- 27. Elyada E, Bolisetty M, Laise P, et al. Cross-species single-cell analysis of pancreatic ductal adenocarcinoma reveals antigen-presenting cancer-associated fibroblasts. Cancer Discov. 2019;9:1102-1123.
- 28. Scanlan MJ, Raj BK, Calvo B, et al. Molecular cloning of fibroblast activation protein alpha, a member of the serine protease family selectively expressed in stromal fibroblasts of epithelial cancers. Proc Natl Acad Sci U S A. 1994;91:5657-5661.
- 29. Simková A, Busek P, Sedo A, Konvalinka J. Molecular recognition of fibroblast activation protein for diagnostic and therapeutic applications. Biochim Biophys Acta Proteins Proteom. 2020;1868:140409.
- 30. Park JE, Lenter MC, Zimmermann RN, Garin-Chesa P, Old LJ, Rettig WJ. Fibroblast activation protein, a dual specificity serine protease expressed in reactive human tumor stromal fibroblasts. J Biol Chem. 1999;274:36505-36512.
- 31. Kraman M, Bambrough PJ, Arnold JN, et al. Suppression of antitumor immunity by stromal cells expressing fibroblast activation protein-alpha. Science. 2010;330:827-830.
- 32. Arnold JN, Magiera L, Kraman M, Fearon DT. Tumoral immune suppression by macrophages expressing fibroblast activation protein- and heme oxygenase-1. Cancer Immunol Res. 2014;2:121-126.

- 33. Lin Y, Li B, Yang X, et al. Fibroblastic FAP promotes intrahepatic cholangiocarcinoma growth via MDSCs recruitment. Neoplasia. 2019;21:1133-1142.
- 34. Feig C, Jones JO, Kraman M, et al. Targeting CXCL12 from FAP-expressing carcinoma associated fibroblasts synergizes with anti-PD-L1 immunotherapy in pancreatic cancer. Proc Natl Acad Sci U S A. 2013;110:20212-20217.
- 35. Orimo A, Gupta PB, Sgroi DC, et al. Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion. Cell. 2005;121:335-348.
- 36. Coto-Llerena M, Ercan C, Kancherla V, et al. High expression of FAP in colorectal cancer is associated with angiogenesis and immunoregulation processes. Front Oncol. 2020;10:979.
- 37. Busek P, Balaziova E, Matrasova I, et al. Fibroblast activation protein alpha is expressed by transformed and stromal cells and is associated with mesenchymal features in glioblastoma. Tumour Biol. 2016;37:13961-13971.
- 38. Röhrich M, Floca R, Loi L, et al. FAP-specific PET signaling shows a moderately positive correlation with relative CBV and no correlation with ADC in 13 IDH wildtype glioblastomas. Eur J Radiol. 2020;127:109021.
- 39. Aimes RT, Zijlstra A, Hooper JD, et al. Endothelial cell serine proteases expressed during vascular morphogenesis and angiogenesis. Thromb Haemost. 2003;89:561-572.
- 40. Dohi O, Ohtani H, Hatori M, et al. Histogenesis-specific expression of fibroblast activation protein and dipeptidylpeptidase-IV in human bone and soft tissue tumours. Histopathology. 2009;55:432-440.
- 41. Cohen SJ, Alpaugh RK, Palazzo I, et al. Fibroblast activation protein and its relationship to clinical outcome in pancreatic adenocarcinoma. Pancreas. 2008;37:154-158.
- 42. Zhang Y, Tang H, Cai J, et al. Ovarian cancer-associated fibroblasts contribute to epithelial ovarian carcinoma metastasis by promoting angiogenesis, lymphangiogenesis and tumor cell invasion. Cancer Lett. 2011;303:47-55.

- 43. Mhawech-Fauceglia P, Yan L, Sharifian M, et al. Stromal expression of fibroblast activation protein alpha (FAP) predicts platinum resistance and shorter recurrence in patients with epithelial ovarian cancer. Cancer Microenviron. 2015;8:23-31.
- 44. Wikberg ML, Edin S, Lundberg IV, et al. High intratumoral expression of fibroblast activation protein (FAP) in colon cancer is associated with poorer patient prognosis. Tumour Biol. 2013;34:1013-1020.
- 45. Lo A, Wang LS, Scholler J, et al. Tumor-promoting desmoplasia is disrupted by depleting FAP-expressing stromal cells. Cancer Res. 2015;75:2800-2810.
- 46. Ostermann E, Garin-Chesa P, Heider KH, et al. Effective immunoconjugate therapy in cancer models targeting a serine protease of tumor fibroblasts. Clin Cancer Res. 2008;14:4584-4592.
- 47. Garin-Chesa P, Old LJ, Rettig WJ. Cell surface glycoprotein of reactive stromal fibroblasts as a potential antibody target in human epithelial cancers. Proc Natl Acad Sci U S A. 1990;87:7235-7239.
- 48. Welt S, Divgi CR, Scott AM, et al. Antibody targeting in metastatic colon cancer: a phase I study of monoclonal antibody F19 against a cell-surface protein of reactive tumor stromal fibroblasts. J Clin Oncol. 1994;12:1193-1203.
- 49. Flentke GR, Munoz E, Huber BT, Plaut AG, Kettner CA, Bachovchin WW. Inhibition of dipeptidyl aminopeptidase IV (DP-IV) by Xaa-boroPro dipeptides and use of these inhibitors to examine the role of DP-IV in T-cell function. Proc Natl Acad Sci U S A. 1991;88:1556-1559.
- 50. Tsai TY, Yeh TK, Chen X, et al. Substituted 4-carboxymethylpyroglutamic acid diamides as potent and selective inhibitors of fibroblast activation protein. J Med Chem. 2010;53:6572-6583.
- 51. Jansen K, Heirbaut L, Cheng JD, et al. Selective inhibitors of fibroblast activation protein (FAP) with a (4-Quinolinoyl)-glycyl-2-cyanopyrrolidine scaffold. ACS Med Chem Lett. 2013;4:491-496.
- 52. Loktev A, Lindner T, Mier W, et al. A tumor-imaging method Ttargeting cancer associated fibroblasts. J Nucl Med. 2018;59:1423-1429.

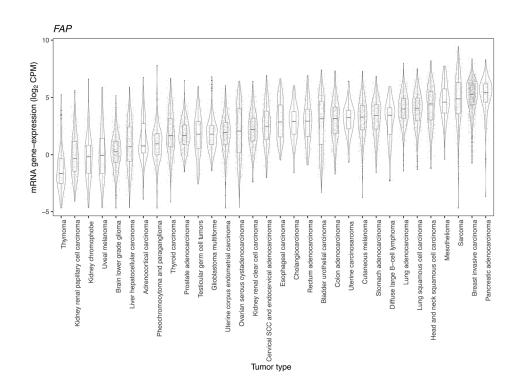
- 53. Loktev A, Lindner T, Burger EM, et al. Development of fibroblast activation protein targeted radiotracers with improved tumor retention. J Nucl Med. 2019;60:1421-1429.
- 54. Lindner T, Loktev A, Altmann A, et al. Development of quinoline-based theranostic ligands for the targeting of fibroblast activation protein. J Nucl Med. 2018;59:1415-1422.
- 55. Giesel FL, Kratochwil C, Lindner T, et al. Ga-FAPI PET/CT: Biodistribution and preliminary dosimetry estimate of 2 DOTA-Containing FAP-targeting agents in patients with various cancers. J Nucl Med. 2019;60:386-392.
- 56. Kratochwil C, Flechsig P, Lindner T, et al. Ga-FAPI PET/CT: Tracer uptake in 28 different kinds of cancer. J Nucl Med. 2019;60:801-805.
- 57. Luo Y, Pan Q, Zhang W. IgG4-related disease revealed by 68Ga-FAPI and 18F-FDG PET/CT. Eur J Nucl Med Mol Imaging. 2019;46:2625-2626.
- 58. Chen H, Zhao L, Ruan D, et al. Usefulness of 68Ga-FAPI and 18F-FDG PET/CT in a Patient With Cholangiocellular Carcinoma. Eur J Nucl Med Mol Imaging. 2020 Jun 25. doi: 10.1007/s00259-020-04940-6.
- 59. Chen H, Pang Y, Wu J, et al. Comparison of [68Ga]Ga-DOTA-FAPI-04 and [18F] FDG PET/CT for the diagnosis of primary and metastatic lesions in patients with various types of cancer. Eur J Nucl Med Mol Imaging. 2020;47:1820-1832.
- 60. Syed M, Flechsig P, Liermann J, et al. Fibroblast activation protein inhibitor (FAPI) PET for diagnostics and advanced targeted radiotherapy in head and neck cancers. Eur J 2020. Nucl Med Mol Imaging.
- 61. Iravani A, Hicks RJ. Imaging the cancer immune environment and its response to pharmacological intervention- Part 2- The Role of Novel PET Agents. J Nucl Med. 2020;61(11):1553-1559.
- 62. Meyer C, Dahlbom M, Lindner T, et al. Radiation dosimetry and biodistribution of 68Ga-FAPI-46 PET imaging in cancer patients. J Nucl Med. 2020;61:1171-1177.

- 63. Koerber SA, Staudinger F, Kratochwil C, et al. The role of FAPI-PET/CT for patients with malignancies of the lower gastrointestinal tract first clinical experience. J Nucl Med. 2020;61(9):1331-1336.
- 64. Giesel F, Adeberg S, Syed M, et al. FAPI-74 PET/CT using either 18F-AlF or cold-kit 68Ga-labeling: Biodistribution, radiation dosimetry and tumor delineation in lung cancer. J Nucl Med. 2020 Jun 26:jnumed.120.245084. doi: 10.2967/jnumed.120.245084
- 65. Toms J, Kogler J, Maschauer S, et al. Targeting fibroblast activation protein: Radiosynthesis and preclinical evaluation of 18F-labeled FAP inhibitor. J Nucl Med. 2020 Apr 24:jnumed.120.242958. doi: 10.2967/jnumed.120.242958.
- 66. Giesel FL, Heussel CP, Lindner T, et al. FAPI-PET/CT improves staging in a lung cancer patient with cerebral metastasis. Eur J Nucl Med Mol Imaging. 2019;46:1754-1755.
- 67. Hao B, Wu J, Pang Y, Sun L, Chen H. 68Ga-FAPI PET/CT in assessment of leptomeningeal metastases in a patient with lung adenocarcinoma. Clin Nucl Med. 2020;45:784-786.
- 68. Hao B, Wu X, Pang Y, et al. [18F]FDG and [68Ga]Ga-DOTA-FAPI-04 PET/CT in the evaluation of tuberculous lesions. Eur J Nucl Med Mol Imaging. 2020 Jul 8. doi: 10.1007/s00259-020-04941-5.
- 69. Windisch P, Röhrich M, Regnery S, et al. Fibroblast activation protein (FAP) specific PET for advanced target volume delineation in glioblastoma. Radiother Oncol. 2020;150:159-163.
- 70. Fischer E, Chaitanya K, Wüest T, et al. Radioimmunotherapy of fibroblast activation protein positive tumors by rapidly internalizing antibodies. Clin Cancer Res. 2012;18:62086218.
- 71. Watabe T, Liu Y, Kaneda-Nakashima K, et al. Theranostics targeting fibroblast activation protein in the tumor stroma: 64Cu- and 225Ac-Labeled FAPI-04 in pancreatic cancer xenograft mouse models. J Nucl Med. 2020;61:563-569.
- 72. Hicks RJ, Jackson P, Kong G, et al. Cu-SARTATE PET imaging of patients with neuroendocrine tumors demonstrates high tumor uptake and retention, potentially allowing prospective dosimetry for peptide receptor radionuclide therapy. J Nucl Med. 2019;60:777-785.

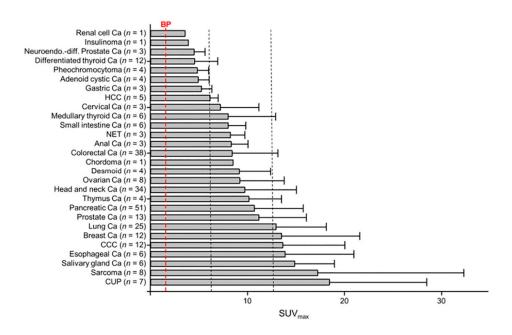
- 73. Hicks RJ. Citius, Altius, Fortius: An Olympian Dream for Theranostics. J Nucl Med. 2017;58:194-195.
- 74. Paterson BM, Roselt P, Denoyer D, et al. PET imaging of tumours with a 64Cu labeled macrobicyclic cage amine ligand tethered to Tyr3-octreotate. Dalton Trans. 2014;43:13861396.
- 75. Scott AM, Wiseman G, Welt S, et al. A Phase I dose-escalation study of sibrotuzumab in patients with advanced or metastatic fibroblast activation protein-positive cancer. Clin Cancer Res. 2003;9:1639-1647.
- 76. Puré E, Blomberg R. Pro-tumorigenic roles of fibroblast activation protein in cancer: back to the basics. Oncogene. 2018;37:4343-4357.
- 77. Rettig WJ, Garin-Chesa P, Beresford HR, Oettgen HF, Melamed MR, Old LJ. Cell-surface glycoproteins of human sarcomas: differential expression in normal and malignant tissues and cultured cells. Proc Natl Acad Sci U S A. 1988;85:3110-3114.
- 78. Gabbiani G, Ryan GB, Majne G. Presence of modified fibroblasts in granulation tissue and their possible role in wound contraction. Experientia. 1971;27:549-550.
- 79. Bauer S, Jendro MC, Wadle A, et al. Fibroblast activation protein is expressed by rheumatoid myofibroblast-like synoviocytes. Arthritis Res Ther. 2006;8:R171.
- 80. Laverman P, van der Geest T, Terry SY, et al. Immuno-PET and immuno-SPECT of rheumatoid arthritis with radiolabeled anti-fibroblast activation protein antibody correlates with severity of arthritis. J Nucl Med. 2015;56:778-783.
- 81. Iravani A, Turgeon GA, Akhurst T, et al. PET-detected pneumonitis following curative-intent chemoradiation in non-small cell lung cancer (NSCLC): recognizing patterns and assessing the impact on the predictive ability of FDG-PET/CT response assessment. Eur J Nucl Med Mol Imaging. 2019;46:1869-1877.
- 82. Mac Manus MP, Ding Z, Hogg A, et al. Association between pulmonary uptake of fluorodeoxyglucose detected by positron emission tomography scanning after radiation therapy for non-small-cell lung cancer and Radiation Pneumonitis. Int J Radiat Oncol Biol Phys. 2011;80(5):1365-71.

- 83. Egger C, Cannet C, Gérard C, et al. Effects of the fibroblast activation protein inhibitor, PT100, in a murine model of pulmonary fibrosis. Eur J Pharmacol. 2017;809:64-72.
- 84. Seuwen K, Magnaldo I, Pouysségur J. Serotonin stimulates DNA synthesis in fibroblasts acting through 5-HT1B receptors coupled to a Gi-protein. Nature. 1988;335:254256.
- 85. Waltenberger J, Lundin L, Oberg K, et al. Involvement of transforming growth factorbeta in the formation of fibrotic lesions in carcinoid heart disease. Am J Pathol. 1993;142:71-78.
- 86. Nagaraju CK, Dries E, Popovic N, et al. Global fibroblast activation throughout the left ventricle but localized fibrosis after myocardial infarction. Sci Rep. 2017;7:10801.
- 87. Tillmanns J, Hoffmann D, Habbaba Y, et al. Fibroblast activation protein alpha expression identifies activated fibroblasts after myocardial infarction. J Mol Cell Cardiol. 2015;87:194-203.
- 88. Heckmann MB, Reinhardt F, Finke D, et al. Relationship between cardiac fibroblast activation protein activity by positron emission tomography and cardiovascular disease. Circ Cardiovasc Imaging. 2020;13:e010628.
- 89. Siebermair J, Köhler MI, Kupusovic J, et al. Cardiac fibroblast activation detected by Ga-68 FAPI PET imaging as a potential novel biomarker of cardiac injury/remodeling. J Nucl Cardiol. 2020.
- 90. Zhao L, Gu J, Fu K, Lin Q, Chen H. 68Ga-FAPI PET/CT in assessment of liver nodules in a cirrhotic patient. Clin Nucl Med. 2020;45:e430-e432.
- 91. Shi X, Xing H, Yang X, et al. Fibroblast imaging of hepatic carcinoma with 68Ga-FAPI04 PET/CT: a pilot study in patients with suspected hepatic nodules. Eur J Nucl Med Mol Imaging. 2020 Sep 25. doi: 10.1007/s12350-020-02307-w.
- 92. Pan Q, Luo Y, Zhang W. Recurrent immunoglobulin G4-related disease shown on 18F-FDG and 68Ga-FAPI PET/CT. Clin Nucl Med. 2020;45:312-313.
- 93. Luo Y, Pan Q, Yang H, Peng L, Zhang W, Li F. Fibroblast activation protein targeted PET/CT with 68Ga-FAPI for imaging IgG4-related disease: comparison to 18F-FDG PET/CT. J Nucl Med. 2020 Jun 8:jnumed.120.244723. doi: 10.2967/jnumed.120.244723.

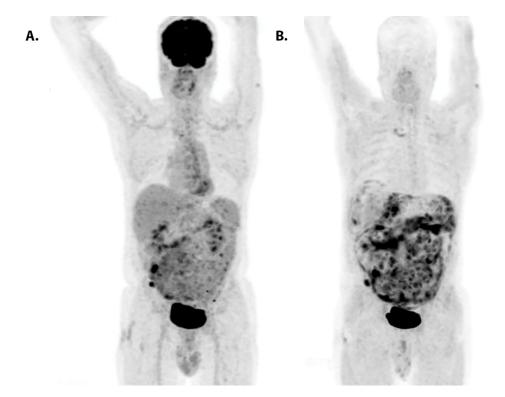
- 94. Kamisawa T, Zen Y, Pillai S, Stone JH. IgG4-related disease. Lancet. 2015;385:14601471.
- 95. Meletta R, Müller Herde A, Chiotellis A, et al. Evaluation of the radiolabeled boronic acid-based FAP inhibitor MIP-1232 for atherosclerotic plaque imaging. Molecules. 2015;20:2081-2099.
- 96. Windisch P, Zwahlen DR, Koerber SA, et al. Clinical results of fibroblast activation protein (FAP) specific PET and implications for radiotherapy planning: Systematic review. Cancers (Basel). 2020;12.
- 97. Lindner T, Altmann A, Kraemer S, et al. Design and development of 99mTc-labeled FAPI-tracers for SPECT-imaging and 188Re therapy. J Nucl Med. 2020;61(10):1507-1513.
- 98. Lepareur N, Lacoeuille F, Bouvry C, et al. Rhenium-188 labeled radiopharmaceuticals: Current clinical applications in oncology and promising perspectives. Front Med (Lausanne). 2019;6:132.
- 99. Cerami E, Gao J, Dogrusoz U, et al. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discov. 2012;2:401404.



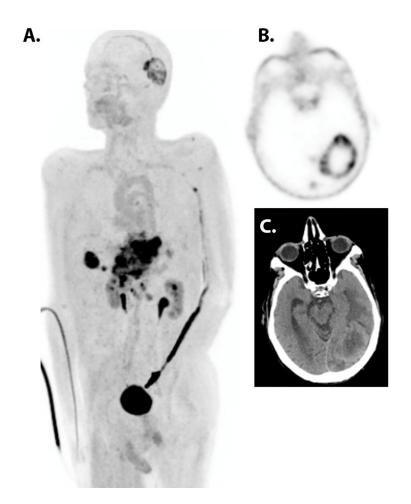
**Figure 1**. Cancer FAP expression results based upon data generated by the TCGA Research Network (99).



**Figure 2.** Average SUVmax of  $^{68}$ Ga-FAPI PET/CT in various tumor entities in comparison to blood pool. Ca = cancer; CCC = cholangiocellular carcinoma; CUP = carcinoma of unknown primary; HCC = hepatocellular carcinoma; NET = neuroendocrine tumor (Reproduced with permission from (56))



**Figure 3.** Peritoneal carcinomatosis. A. FDG PET maximum intensity projection (MIP) image demonstrates focal uptake in the region of the cecum and transverse colon but impression of diffuse peritoneal disease. B. The corresponding FAPI-04 PET MIP clearly demonstrates diffuse peritoneal disease including involvement of the subphrenic spaces.



**Figure 4.** Gastric cancer with nodal, liver and brain metastases. A. FAPI PET MIP, B. Necrotic left occipital metastasis on transaxial FAPI PET, C. Correlative CT.



**Figure 5.** Giant cell arteritis. A. FAPI PET MIP, B. Coronal slice through aortic arch and subclavian arteries, C. Sagittal slice through thoracic aorta.