First-in-human evaluation of ¹⁸F-SynVesT-1, a novel

radioligand for PET imaging of synaptic vesicle protein 2A

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Financial support:R01AG052560;Michael J. Fox Foundation;R01AG065474

Running Title:First-in-Human Study of SynVesT-1

ABSTRACT

The use of synaptic vesicle protein 2A (SV2A) radiotracers with positron emission tomography (PET) imaging could provide a way to measure synaptic density quantitatively in living humans. ¹¹C-UCB-J, previously developed and assessed in nonhuman primates and humans, showed excellent kinetic properties as a PET radioligand. However, it is labeled with the short half-life isotope ¹¹C. We developed a new tracer, an ¹⁸F-labeled difluoro-analog of UCB-J (¹⁸F-SynVesT-1, a.k.a. ¹⁸F-SDM-8), which displayed favorable properties in monkeys. The purpose of this first-in-human study was to assess the kinetic and binding properties of ¹⁸F- SynVesT-1 and compare with ¹¹C-UCB-J.

Methods: Eight healthy volunteers participated in a baseline study of ¹⁸F-SynVesT-1. Four of these subjects were also scanned after a blocking dose of the anti-epileptic drug levetiracetam (20 mg/kg). Metabolite-corrected arterial input functions were measured. Regional time-activity curves (TACs) were analyzed using one- and two-tissue compartment (1TC, 2TC) models and multilinear analysis 1 (MA1) to compute distribution volume (V_T) and binding potential (BP_{ND}). The centrum semiovale was used as a reference region. The Lassen plot was applied to compute levetiracetam occupancy and non-displaceable distribution volume (V_{ND}). Standardized uptake value ratio (SUVR) -1 over several time windows was compared with BP_{ND} .

Results: Regional TACs were fitted better with the 2TC model than the 1TC model, but 2TC V_T estimates were unstable. The 1TC V_T values matched well with those from the 2TC model (excluding the unstable values), Thus, 1TC was judged as the most useful model for quantitative analysis of ¹⁸F-SynVesT-1 imaging data. Minimum scan time for stable V_T measurement was 60 min. The rank order of V_T and BP_{ND} values was similar between ¹⁸F-SynVesT-1 and ¹¹C-UCB-J. Regional V_T values were slightly higher for ¹¹C-UCB-J, but BP_{ND} values were higher for ¹⁸F-SynVesT-1, though these differences were not significant. Levetiracetam reduced the uptake of

¹⁸F-SynVesT-1 in all regions and produced occupancy of 85.7%. SUVR-1 of ¹⁸F-SynVesT-1 from 60-90 min matched best with 1TC BP_{ND} .

Conclusion: The novel SV2A tracer, ¹⁸F-SynVesT-1, displays excellent kinetic and in vivo binding properties in humans and holds great potential for the imaging and quantification of synaptic density in neuropsychiatric disorders.

Keywords: Positron emission tomography; SV2A; Brain imaging; Kinetic modeling; Synaptic density

INTRODUCTION

The synaptic vesicle glycoprotein 2A (SV2A) is located in the presynaptic vesicle membrane of virtually all synapses (1), and is the target of the anticonvulsant drug levetiracetam (2). We have previously developed ¹¹C-UCB-J as a positron emission tomography (PET) radiotracer, tested it in nonhuman primates and humans, and found it to have excellent imaging properties (3-5). SV2A PET could provide a way to measure synaptic density quantitatively in living humans and to track changes in synaptic density with disease. For example, SV2A PET imaging with ¹¹C-UCB-J showed lower hippocampal SV2A specific binding in patients with Alzheimer's disease in comparison with cognitively normal subjects (6). In major depressive disorder (7), the severity of depressive symptoms was inversely correlated with SV2A density. Synaptic changes have also been found in Parkinson's disease (8) and schizophrenia (9).

¹¹C-UCB-J has excellent test-retest reproducibility to measure distribution volume (V_T) with a short scan time (~60 min) (5). However, it has a half-life of 20 minutes, which requires the tracer to be produced on-site. A SV2A radiotracer labeled with ¹⁸F for PET imaging is attractive for clinical diagnostic applications. We have therefore developed ¹⁸F-SynVesT-1 (the difluoro-analog of UCB-J) and demonstrated its suitability for imaging SV2A in rhesus monkeys with the same favorable properties as ¹¹C-UCB-J: high brain uptake and good specific binding signals (*10*). Note that the previous identifier for this tracer was ¹⁸F-SDM-8, as it was part of a series of ¹⁸F-labeled UCB-J analogs developed as synaptic density markers (*10*), and has also been studied elsewhere with the identifier ¹⁸F-MNI-1126 (*11*). The new name ¹⁸F-SynVesT-1 was since agreed upon between the groups to eliminate future confusion in the field with two different names for the same radioligand.

In this first-in-human study, we evaluated ¹⁸F-SynVesT-1 in healthy human volunteers. The purpose of this paper is to determine a suitable kinetic model for quantitative analysis of ¹⁸F-

SynVesT-1 imaging data. A subgroup of volunteers also participated in a baseline and blocking study with levetiracetam using both ¹⁸F-SynVesT-1 and ¹¹C-UCB-J for comparison.

MATERIALS AND METHODS

Radiotracer Synthesis

¹⁸F-SynVesT-1 and ¹¹C-UCB-J were synthesized as described previously (3,10).

Human Subjects

Four healthy volunteers (4 males,44±13y.,body mass index (BMI):30±2) completed a baseline study with ¹⁸F-SynVesT-1. A second group of healthy volunteers (2 males and 2 females,38±15y.,BMI:27±4) were enrolled in a baseline-blocking study of ¹⁸F-SynVesT-1 and ¹¹C-UCB-J using levetiracetam as the blocking drug. All subjects were screened with a physical exam, medical history, routine laboratory studies, pregnancy tests (for females), and electrocardiogram to assess for eligibility. The IRB-approved study was also approved by the Yale-New Haven Hospital Radiation Safety Committee, and performed in accordance with federal guidelines and regulations of the United States for the protection of human research subjects contained in Title 45 Part 46 of the Code of Federal Regulations (45 CFR 46). All subjects signed a written informed consent.

Brain PET Studies

PET imaging. PET images were acquired using the High Resolution Research Tomograph (HRRT, Siemens Medical Systems, Knoxville, TN), which acquires 207 slices (1.2mm slice separation), in list mode for 120 min for ¹⁸F-SynVesT-1 and 90 min for ¹¹C-UCB-J. A 6-min transmission scan was conducted for attenuation correction. Dynamic scan data were reconstructed in 27 frames (6×30sec, 3×1min, 2×2min, and 16×5min) for ¹¹C-UCB-J, with 6 additional 5 min frames for ¹⁸F-SynVesT-1 with corrections for attenuation, normalization, scatter, randoms, and

dead time using the MOLAR algorithm (12). Event-by-event motion correction (13) was included in the reconstruction based on measurements with the Polaris Vicra sensor (NDI Systems, Waterloo, Canada). Subjects were administered ¹⁸F-SynVesT-1 as an intravenous injection over 1 min by an automatic pump (Harvard PHD 22/2000, Harvard Apparatus Holliston, MA, USA). ¹¹C-UCB-J was administered as a bolus for 2 subjects and as a bolus plus infusion for the other 2 subjects. The blocking scan was conducted 3 h after intravenous administration of levetiracetam (¹⁸F-SynVesT-1:20 mg/kg; ¹¹C-UCB-J:10 mg/kg (*n*=3) and 20 mg/kg (*n*=1)).

Magnetic resonance imaging. Each subject underwent an MR scan for PET image registration. MR imaging was performed using a 3D MPRAGE pulse sequence with an echo time of 2.78 ms, repetition time of 2,500 ms, inversion time of 1,100 ms, and flip angle of 7° on a 3T whole-body scanner (Trio; Siemens Medical Systems) with a circularly polarized head coil.

Arterial input function measurement. Discrete blood samples were manually drawn every 10 s from 10 to 90 s, every 15 s from 90 s to 3 min, and then at 3.5, 5, 6.5, 8, 12, 15, 20, 25, 30, 45, 60, 75, and 90 min for both tracers, with 2 additional samples taken at 105 and 120 min for ¹⁸F-SynVesT-1. Samples were centrifuged to obtain plasma and then counted with a calibrated well counter.

Analysis of radiotracer metabolism was performed from plasma samples collected at 3, 8, 15, 30, 60, and 90 min after injection for both tracers, with an additional sample at 120 min for ¹⁸F-SynVesT-1. Metabolite analysis was performed using the column-switching HPLC method (*14*) to determine the parent fraction, as previously described (*10*). An ultrafiltration-based method (Centrifree, Millipore) was used to measure the plasma free fraction (f_P) (*10*).

Image registration and regions of interest. PET images were corrected for motion using a mutual information algorithm (FSL-FLIRT) by frame-by-frame registration to a summed image (0-10 min post-injection) using a 6-parameter mutual information algorithm (FLIRT, FSL). The summed PET image was then co-registered to the subject's T1-weighted MR image (6-parameter affine registration), which was subsequently co-registered to the Automated Anatomical Labeling

(AAL) template (15) in Montreal Neurological Institute (MNI) (16) space using a nonlinear transformation (Bioimage suite) (17). Using the combined transformations from template-to-PET space, regional tissue time-activity curves (TACs) were generated in the following regions: amygdala, anterior cingulate cortex, caudate nucleus, cerebellum, frontal cortex, globus pallidus, hippocampus, insular cortex, occipital cortex, parietal cortex, posterior cingulate cortex, putamen, temporal cortex, and thalamus. The region of interest for the centrum semiovale (CS) was designed to minimize the partial volume effect (18).

Quantitative analysis. For ¹⁸F-SynVesT-1, regional distribution volumes (V_T) were computed from the TACs using one-tissue (1TC) and two-tissue (2TC) compartment models. The relative fit quality of 1TC and 2TC models was compared with the *F* test. Percentage standard error (%SE) was estimated from the theoretical parameter covariance matrix. Multilinear analysis 1 (MA1) (*19*) was also applied to estimate V_T by changing the starting time t^* (from 10 to 60 min with 10 min increments). The best kinetic model for ¹⁸F-SynVesT-1 was chosen based on analysis of data from the eight baseline scans.

For comparison of parametric images to ¹¹C-UCB-J, which used the 1TC model (5), parametric $V_{\rm T}$ images for ¹⁸F-SynVesT-1 were also generated with the 1TC model using a basis function method with k_2 limited to the range of 0.01–1.0 min⁻¹ and without post-smoothing. All modeling was performed with in-house programs using IDL 8.0 (ITT Visual Information Solutions, Boulder, CO, USA).

The minimal scan duration for V_T quantification was evaluated by considering shorter datasets (30 to 120 min in 10 min increments, n=8) using the selected kinetic model. The percent differences in V_T values derived using data from the shorter intervals and 120 min were calculated. The minimum scan time was chosen based on criteria defined previously (20).

The CS region was used as a reference region to compute regional binding potential (BP_{ND}) from V_T values. The Lassen plot was used to determine target occupancy by levetiracetam and the non-displaceable distribution volume (V_{ND}) (21). The V_{ND} from gray matter regions was compared to baseline V_T in CS to test the suitability of CS as a reference region.

In addition, a simplified outcome measure, the standardized uptake value ratio (SUVR), was evaluated in comparison to BP_{ND} . Static SUVR-1 values, which would equal BP_{ND} at equilibrium, were computed for 9 time windows of 30-min duration: 10-40, 20-50, 30-60, 40-70, 50-80, 60-90, 70-100, 80-110, and 90-120 min, and compared to BP_{ND} , calculated from regional V_T ratio (target/reference–1). All outcome measures were computed from regional TAC analysis of baseline scans.

RESULTS

Human Injection and Scan Parameters

The mean administered activity dose of ¹⁸F-SynVesT-1 was $180\pm7MBq$ (range,167-186MBq) and $180\pm4MBq$ (range,175-185 MBq) for the baseline (*n*=8) and blocking (*n*=4) scans, respectively. Injection and scan parameters are listed in Table 1.

Safety

No significant clinical changes were observed with the administration of 18 F-SynVesT-1 in injected mass dose of up to 0.55 µg.

Plasma Analysis

Data from plasma analysis are displayed in Figure 1. At 60 min after radiotracer injection, the fraction of radioactivity corresponding to the parent compound was $26\pm9\%$ (*n*=8,baseline) and $26\pm9\%$ (*n*=4,blocking) for ¹⁸F-SynVesT-1, and $23\pm8\%$ (*n*=2,baseline) and $23\pm10\%$ (*n*=2, blocking) for ¹¹C-UCB-J. The plasma free fraction was 0.31 ± 0.01 (*n*=8,baseline) and 0.30 ± 0.02 (*n*=4, blocking) for ¹⁸F-SynVesT-1 and 0.27 ± 0.02 (*n*=4,baseline) and 0.28 ± 0.02 (*n*=4,blocking) for ¹¹C-

UCB-J. The parent fraction at 60 min and free fraction in plasma were not significantly different between baseline and blocking conditions with ¹⁸F-SynVesT-1.

Brain Distribution and Kinetics

Typical TACs and their fitting are shown in Figure 2. High uptake was seen in gray matter regions and low uptake in white matter regions. The order of regional uptake levels was similar between ¹⁸F-SynVesT-1 and ¹¹C-UCB-J. Standardized uptake value (SUV) in brain regions peaked at 5-20 min and 10-25 min post-injection, respectively, for ¹⁸F-SynVesT-1 and ¹¹C-UCB-J.

The 2TC model was favored over the 1TC model by the F-test (66% of fits), and the differences were found predominantly in the cerebellum, hippocampus, and neocortical regions. In particular, a moderate lack of fit was seen in the cerebellum with the 1TC model. Although the 2TC fits were excellent, kinetic parameters were not reliably estimated (%SE>100% in 32% of the fits for k_3 and 77% of fits for k_4), which led to poor estimation of 2TC V_T values in 60% of fits (%SE>10%). In stable $V_{\rm T}$ cases, (%SE<10%) the estimated k_4 value was not small (~0.02). But, in unstable $V_{\rm T}$ cases (%SE>100%), the estimated k_4 value was very small (<10⁻⁹). The ratio of k_3/k_4 was also unstable (%SE>1000%). Excluding the unreliable 2TC $V_{\rm T}$ estimates, 1TC $V_{\rm T}$ values matched exceptionally with those from 2TC (1TC V_T =1.01×2TC V_T -0.32, R^2 =1.00). The MA1 method was also tested with a range of t^* settings. If t^* has a very small effect on V_T , then the 1TC model is appropriate. The results showed that MA1 $V_{\rm T}$ values derived with different t* did not differ from those estimated by 1TC: the percent differences ranged from $0.2\%\pm0.3\%$ (t*=10 min) to $2.7\% \pm 0.8\%$ (t*=60 min). The largest difference was seen in the cerebellum and hippocampus with $t^*=60 \min (4-5\%)$. Given the insensitivity of V_T estimates to t^* setting in MA1, the quality of fitting by 1TC, and excellent match with reliable 2TC $V_{\rm T}$ values, the 1TC was judged as the most appropriate model for analysis of ¹⁸F-SynVesT-1 imaging data.

The mean 1TC-estimated $V_{\rm T}$ values (mL/cm³) of ¹⁸F-SynVesT-1 ranged from 3.5±0.4 in the CS to 19.3±2.8 in the putamen (Table 2), while K_1 estimates (mL/min/cm³) ranged from

0.112±0.011 (CS) to 0.377±0.040 (putamen). These K_1 values were similar to those of ¹¹C-UCB-J (range of 0.113±0.010 to 0.367±0.047), with relative difference of 4±8%. The 1TC k_2 estimates (1/min) ranged from 0.014±0.001 (amygdala) to 0.032±0.003 (CS) for ¹⁸F-SynVesT-1.

Voxel-based parametric images were computed with 120 and 90 min of scan data, respectively, for ¹⁸F-SynVesT-1 and ¹¹C-UCB-J (Figure 3). For ¹⁸F-SynVesT-1, effectively identical V_T values were obtained with both parametric imaging and TAC fitting: V_T (voxel)=1.01× V_T (TAC) + 0.02, R^2 =1.00.

A minimum scan duration of 60 min was required to satisfy the stability criteria for V_T estimates in all brain regions in 1TC TAC analysis. For specific regions, 60 min was required for the hippocampus and cerebellum, 40 min for the anterior cingulate cortex, and 30 min for other regions (Table 2).

Figure 4 shows examples of Lassen plots using $V_{\rm T}$ values measured with ¹⁸F-SynVesT-1 and ¹¹C-UCB-J in the baseline-blocking study with levetiracetam (Table 3). The occupancy by 20 mg/kg of levetiracetam was 85.3±4.7% as measured by ¹⁸F-SynVesT-1 and 82.5% as measured by ¹¹C-UCB-J, while occupancy produced by the 10 mg/kg dose was 76.4±5.6% as measured by ¹¹C-UCB-J. The $V_{\rm ND}$ values, determined as the *x*-intercepts from the Lassen plots, were 2.38±0.33 mL/cm³ for ¹⁸F-SynVesT-1 and 3.13±0.40 mL/cm³ for ¹¹C-UCB-J. The gray matter $V_{\rm ND}$ values were lower than the baseline CS $V_{\rm T}$ values by 32±16% for ¹⁸F-SynVesT-1 and 29±13% for ¹¹C-UCB-J; these offsets were not significantly different between the tracers (*P*=0.83). If $V_{\rm ND}$ in CS equals that in gray matter, these data indicate *BP*_{ND,CS} is 0.47 and 0.41 for ¹⁸F-SynVesT-1 and ¹¹C-UCB-J, respectively. However, we previously showed that $V_{\rm ND}$ in CS is > that in gray matter (*18*).

A comparison of regional BP_{ND} values between the two tracers was performed using either baseline CS V_T values or Lassen plot V_{ND} values as reference values (Supplement Table 1). By either reference method, ¹⁸F-SynVesT-1 had higher BP_{ND} values than ¹¹C-UCB-J, but the difference was not statistically significant. Using the CS as a reference region for calculation of BP_{ND} from regional V_T values, the mean 1TC BP_{ND} values ranged from 2.5±0.5 in the globus pallidus to 4.5±0.5 in the putamen for ¹⁸F-SynVesT-1 (Table 2). These values were then compared to SUV ratios using different 30-min time windows. Percent differences between SUVR-1 and BP_{ND} are shown in Figure 5. The smallest percent difference was seen when the time window for SUVR-1 calculation was 60-90 min. The regression line was SUVR-1(60-90 min)=0.95× BP_{ND} +0.15 (R^2 =0.97).

DISCUSSION

This is the first-in-human study of the novel SV2A PET tracer ¹⁸F-SynVesT-1, including baseline PET scans to evaluate methods for kinetic analysis of imaging data, levetiracetam occupancy scans to assess non-specific binding, and comparison with the established tracer ¹¹C-UCB-J.

In choosing the best kinetic analysis method, the 1TC and 2TC models were compared. The 2TC was the preferred model for analysis of ¹⁸F-SynVesT-1 imaging data based on the *F*-test in 66% of fits. However, the values of k_3 and k_4 were poorly estimated by the 2TC model, which resulted in unreliable V_T estimation. Excluding such unreliable estimates, the 1TC and 2TC models provided almost identical V_T values. Judged from visual inspection of the curve fitting, the 2TC model fitted better in the cerebellum than the 1TC model, but the fitting results were similar in other regions. Similar results were reported for ¹¹C-UCB-J (*5*), i.e., the 2TC model was favored in 73% of fits and a small lack of fit was seen in the cerebellum and hippocampus.

We then assessed the MA1 method with different t^* settings. For tracers that require the use of the 2TC model, i.e., 1TC-derived V_T values differ from those by 2TC, MA1-derived V_T values with early and late t^* settings will be similar to those from 1TC and 2TC, respectively. In our study, however, the change in MA1 V_T values over different t^* settings was small in all regions, with the largest change in the cerebellum (4.6%). This supports the result that the 1TC and 2TC models provide similar V_T estimates. Given the quality of curve fitting, small difference in V_T values estimated by the 1TC and 2TC models, and unreliable $V_{\rm T}$ estimation by 2TC, we chose the 1TC as the best model for analysis of ¹⁸F-SynVesT-1 imaging data.

Regional $V_{\rm T}$ values derived from parametric images matched well with those from TAC analysis. The statistical quality of $V_{\rm T}$ images was similarly high for ¹⁸F-SynVesT-1 and ¹¹C-UCB-J, so that no spatial smoothing was required for either tracer. For centers with the ability to produce C-11 tracer, ultimately it will be of interest to assess which tracer provides the best image quality with matched radiation dose.

The CS has been proposed as a suitable reference region because of its negligible SV2A level observed in the baboon brain by Western blot analysis (4). Since CS has been used as a reference region to quantify ¹¹C-UCB-J specific binding (6,22), we also evaluated the use of CS as a reference region to compute BP_{ND} and SUVR of ¹⁸F-SynVesT-1. Rossano et al. (18) investigated the CS reference region for ¹¹C-UCB-J quantification and optimized the location and size of the ROI, which was adopted in the current study. As observed previously with ¹¹C-UCB-J, a decrease in ¹⁸F-SynVesT-1 V_T values was seen in the CS following SV2A blockade with levetiracetam. For both tracers, the V_{ND} determined from the occupancy plot was ~30% lower than the CS V_T . Thus, use of the CS as reference region might lead to underestimation of BP_{ND} and SUVR. Nonetheless, the CS may still serve as a useful reference region if there is a consistent relationship between CS V_T and V_{ND} , as was shown for ¹¹C-UCB-J (18).

Overall, the mean $V_{\rm T}$ values of ¹¹C-UCB-J were slightly higher than those of ¹⁸F-SynVesT-1 across all brain regions, including the CS (Table 3). When the CS was used as a reference region, ¹⁸F-SynVesT-1 gave higher $BP_{\rm ND}$ values than ¹¹C-UCB-J, with the ratio of $BP_{\rm ND}$ (¹¹C-UCB-J)/ $BP_{\rm ND}$ (¹⁸F-SynVesT-1) at 0.84±0.16 (Supplement Table 1). When the $V_{\rm ND}$ values derived from the Lassen occupancy plots were used, a similar $BP_{\rm ND}$ ratio of 0.79±0.36 was obtained. That is, the mean $BP_{\rm ND}$ values of ¹⁸F-SynVesT-1 were ~20% higher than those of ¹¹C-UCB-J. Note, however, that with the small size of this study (*n*=4), these differences were not statistically significant. Lower logD of ¹⁸F-SynVesT-1 (2.32 vs. 2.53 for ¹¹C-UCB-J) contributes to higher $f_{\rm ND}$ (0.130 vs. 0.086 for ¹¹C-UCB-J) and lower V_{ND} (2.38 vs. 3.13 for ¹¹C-UCB-J). However, the relative values of $BP_{ND}(=f_{ND}B_{max}/K_D)$ also depend on the in vivo K_D . The average in vitro K_i was 3.3nM for SynVesT-1 and 2.7nM for UCB-J, taken from two sets of measurements published previously (K_i of 2.2 and 4.7nM, respectively, for SynVesT-1; 1.5 and 3.0nM for UCB-J) (23) and one set of our own measurement (K_i of 3.1nM for SynVesT-1 and 3.7nM for UCB-J, unpublished) using ³H-UCB-J, and human cortex homogenates. Using these average in vitro K_i values as surrogates for in vivo K_D , we found $f_{ND}/K_D(^{11}C-UCB-J)/f_{ND}/K_D(^{18}F-SynVesT-1)=0.81$, which matches well with $BP_{ND}(^{11}C-UCB-J)/BP_{ND}(^{18}F-SynVesT-1)=0.79$ found in the present study.

Another way to compare specific binding between tracers is to use the graphical method of Guo et al. (Guo plot) (24), where the sign of the *y*-intercept predicts which tracer has a higher specific binding signal. However, when the Guo plot was applied to compare ¹⁸F-SynVesT-1 and ¹¹C-UCB-J, the *y*-intercept could not be reliably estimated since all the data points are far from the origin.

It should be pointed out that, of the 4 subjects with both ¹¹C-UCB-J and ¹⁸F-SynVesT-1 scans, two had their ¹¹C-UCB-J scans conducted under a bolus plus infusion protocol, and the other two received a bolus injection of the radiotracer. This difference in tracer administration methods should not affect the results, since the outcome measure, V_T , was based on kinetic analysis, which accounts for the difference in input function. Although the number of subjects with ¹¹C-UCB-J scans was limited in this study, their regional V_T values were close to those in the literature (*5*, *18*). Note that for comparison of regional TACs, input functions, and parent fraction curves between tracers, we used the data from the subjects receiving bolus injections of the radiotracers.

In order to simplify the imaging and analysis protocol for ¹⁸F-SynVesT-1, shorter scan times and simplified quantification method are desirable. By comparing V_T values from different scan lengths to those derived from the 120-min scan data, it was found that a scan time of 60 min was sufficient to provide stable V_T estimates for ¹⁸F-SynVesT-1. For quantification without arterial sampling, we assessed the possibility to use SUVR-1 as a surrogate for *BP*_{ND}. The SUVR-1 values underestimated BP_{ND} at early time windows, and the difference between SUVR-1 and BP_{ND} monotonically increased by shifting the time window. The best match was seen when the time window for SUVR-1 calculation was 60-90 min. Thus, SUVR-1 calculated from 60-90 min post injection of ¹⁸F-SynVesT-1 can be used as an appropriate substitute for BP_{ND} as a measure of specific binding signal, thus simplifying the imaging and quantification protocols for this novel SV2A radiotracer. Note, this result was obtained in healthy control baseline scans and could be affected by disease or drug administration. Indeed, SUVR-1 from 60-90 min was ~20% lower than BP_{ND} in the levetiracetam blocking scans.

CONCLUSIONS

This first-in-human study showed that ¹⁸F-SynVesT-1 is an excellent PET tracer for SV2A. ¹⁸F-SynVesT-1 showed similarly good properties to the existing radiotracer ¹¹C-UCB-J: high brain uptake, fast and reversible kinetics and high specific binding. The 1TC model was chosen as the best model for quantitative kinetic analysis of ¹⁸F-SynVesT-1 imaging data. Regional BP_{ND} levels of ¹⁸F-SynVesT-1 were higher than those of ¹¹C-UCB-J. SUVR-1 from 60-90 min post-injection provided an excellent match with the 1TC BP_{ND} of ¹⁸F-SynVesT-1 and thus can serve as a surrogate quantitative measurement of specific binding in a short scan time without invasive arterial sampling. The longer half-life of this tracer will facilitate its broad application in studies of synaptic density in many neurodegenerative and neuropsychiatric populations.

DISCLOSURE/CONFLICT OF INTEREST

The radioligand ¹⁸F-SynVesT-1 (formerly referred to as ¹⁸F-SDM-8) is contained in the international patent application PCT/US2018/018388, Radiolabeled Pharmaceuticals and Methods of Making and Using Same, filed on February 15, 2018 (Inventors:YH, ZC, SL, NN, and REC). The other authors declare that they have no conflict of interest.

ACKNOWLEDGMENTS

The authors appreciate the excellent technical assistance of the staff at the Yale University PET Center. This publication was made possible by CTSA Grant UL1 RR024139 jointly from the National Center for Research Resources (NCRR) and the National Center for Advancing Translational Sciences (NCATS), components of the National Institutes of Health (NIH). Its contents are solely the responsibility of the authors and do not necessarily represent the official view of NIH.

KEY POINTS

Question:Does ¹⁸F-SynVesT-1 show suitable kinetic properties to quantify SV2A density in humans in comparison with the existing SV2A tracer ¹¹C-UCB-J?

Pertinent Findings:¹⁸F-SynVesT-1 showed excellent *in vivo* properties with high brain uptake, reversible kinetics, and high specific binding, similar to ¹¹C-UCB-J.

Implications for Patient Care: This longer half-life tracer will be more useful for clinical studies in terms of allowing off-site production and distribution.

References

- 1. Bajjalieh SM, Frantz GD, Weimann JM, McConnell SK, Scheller RH. Differential expression of synaptic vesicle protein 2 (SV2) isoforms. *J Neurosci.* 1994;14:5223-5235.
- 2. Lynch BA, Lambeng N, Nocka K, et al. The synaptic vesicle protein SV2A is the binding site for the antiepileptic drug levetiracetam. *Proc Natl Acad Sci U S A*. 2004;101:9861-9866.
- **3.** Nabulsi NB, Mercier J, Holden D, et al. Synthesis and preclinical evaluation of 11C-UCB-J as a PET tracer for imaging the synaptic vesicle glycoprotein 2A in the brain. *J Nucl Med.* 2016;57:777-784.
- **4.** Finnema SJ, Nabulsi NB, Eid T, et al. Imaging synaptic density in the living human brain. *Sci Transl Med.* 2016;8:348ra396.
- **5.** Finnema SJ, Nabulsi NB, Mercier J, et al. Kinetic evaluation and test-retest reproducibility of [(11)C]UCB-J, a novel radioligand for positron emission tomography imaging of synaptic vesicle glycoprotein 2A in humans. *J Cereb Blood Flow Metab.* 2018;38:2041-2052.
- 6. Chen MK, Mecca AP, Naganawa M, et al. Assessing synaptic density in Alzheimer disease with synaptic vesicle glycoprotein 2A positron emission tomographic imaging. *JAMA Neurol.* 2018;75:1215-1224.
- 7. Holmes SE, Scheinost D, Finnema SJ, et al. Lower synaptic density is associated with depression severity and network alterations. *Nat Commun.* 2019;10:1529.
- **8.** Matuskey D, Tinaz S, Wilcox KC, et al. Synaptic changes in Parkinson disease assessed with in vivo imaging. *Ann Neurol.* 2020;87:329-338.
- **9.** Onwordi EC, Halff EF, Whitehurst T, et al. Synaptic density marker SV2A is reduced in schizophrenia patients and unaffected by antipsychotics in rats. *Nat Commun.* 2020;11:246.
- **10.** Li S, Cai Z, Wu X, et al. Synthesis and in vivo evaluation of a novel PET radiotracer for imaging of synaptic vesicle glycoprotein 2A (SV2A) in nonhuman primates. *ACS Chem Neurosci.* 2019;10:1544-1554.
- **11.** Constantinescu CC, Tresse C, Zheng M, et al. Development and in vivo preclinical imaging of fluorine-18-labeled synaptic vesicle protein 2A (SV2A) PET tracers. *Mol Imaging Biol.* 2019;21:509-518.
- 12. Carson RE, Barker WC, Liow JS, Johnson CA. Design of a motion-compensation OSEM list-mode algorithm for resolution-recovery reconstruction for the HRRT. *IEEE 2003 Nuclear Science Symposium Conference Record.* 2003;5:3281-3285.
- **13.** Jin X, Mulnix T, Gallezot JD, Carson RE. Evaluation of motion correction methods in human brain PET imaging--a simulation study based on human motion data. *Med Phys.* 2013;40:102503.
- 14. Hilton J, Yokoi F, Dannals RF, Ravert HT, Szabo Z, Wong DF. Column-switching HPLC for the analysis of plasma in PET imaging studies. *Nucl Med Biol.* 2000;27:627-630.
- **15.** Tzourio-Mazoyer N, Landeau B, Papathanassiou D, et al. Automated anatomical labeling of activations in SPM using a macroscopic anatomical parcellation of the MNI MRI single-subject brain. *Neuroimage*. 2002;15:273-289.
- **16.** Holmes CJ, Hoge R, Collins L, Woods R, Toga AW, Evans AC. Enhancement of MR images using registration for signal averaging. *J Comput Assist Tomogr*. 1998;22:324-333.
- 17. Papademetris X, Jackowski M, Rajeevan N, Constable RT, Staib LH. Bioimage suite: An integrated medical image analysis suite. *Insight J.* 2006;2006:209.
- **18.** Rossano S, Toyonaga T, Finnema SJ, et al. Assessment of a white matter reference region for (11)C-UCB-J PET quantification. *J Cereb Blood Flow Metab.* 2019:[Epub ahead of print].

- **19.** Ichise M, Toyama H, Innis RB, Carson RE. Strategies to improve neuroreceptor parameter estimation by linear regression analysis. *J Cereb Blood Flow Metab.* 2002;22:1271-1281.
- **20.** Frankle WG, Huang Y, Hwang DR, et al. Comparative evaluation of serotonin transporter radioligands ¹¹C-DASB and ¹¹C-McN 5652 in healthy humans. *J Nucl Med.* 2004;45:682-694.
- **21.** Cunningham VJ, Rabiner EA, Slifstein M, Laruelle M, Gunn RN. Measuring drug occupancy in the absence of a reference region: the Lassen plot re-visited. *J Cereb Blood Flow Metab.* 2010;30:46-50.
- **22.** Koole M, van Aalst J, Devrome M, et al. Quantifying SV2A density and drug occupancy in the human brain using [(11)C]UCB-J PET imaging and subcortical white matter as reference tissue. *Eur J Nucl Med Mol Imaging*. 2019;46:396-406.
- **23.** Patel S, Knight A, Krause S, et al. Preclinical in vitro and in vivo characterization of synaptic vesicle 2A-targeting compounds amenable to F-18 labeling as potential PET radioligands for imaging of synapse integrity. *Mol Imaging Biol.* 2019.
- 24. Guo Q, Owen DR, Rabiner EA, Turkheimer FE, Gunn RN. A graphical method to compare the in vivo binding potential of PET radioligands in the absence of a reference region: application to [(1)(1)C]PBR28 and [(1)(8)F]PBR111 for TSPO imaging. *J Cereb Blood Flow Metab.* 2014;34:1162-1168.

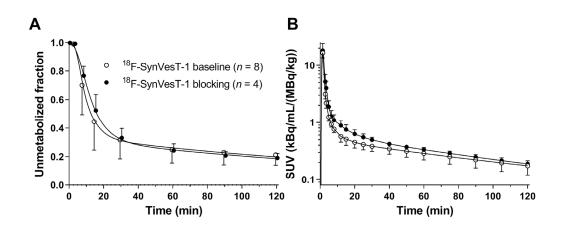


Figure 1:Mean±SD of (A) plasma parent fraction and (B) metabolite-corrected plasma activity in the baseline (closed circles, n=8) and levetiracetam blocking (open circles, n=4) studies with ¹⁸F-SynVesT-1. Panel B is displayed in SUV units [concentration/(injected dose/body weight)].

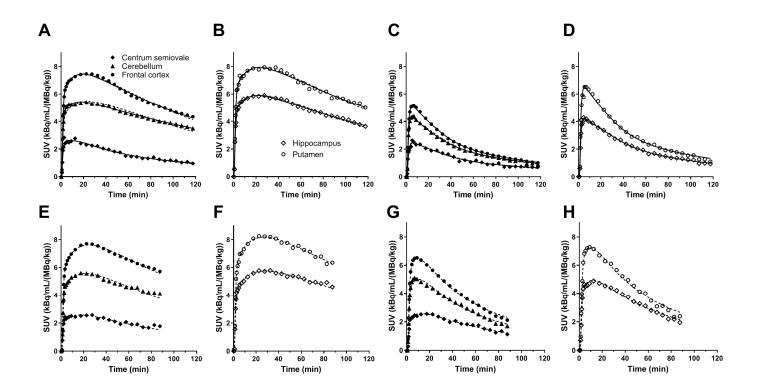


Figure 2:Time-activity curves and fittings with the 1TC (dotted curve) and 2TC (solid curve) models in the same subject under baseline (**A** and **B**) and levetiracetam blocking (**C** and **D**) conditions with ¹⁸F-SynVesT-1, and under the same baseline (**E** and **F**) and blocking (**G** and **H**) conditions with ¹¹C-UCB-J. The 2TC model was not applied to ¹¹C-UCB-J.

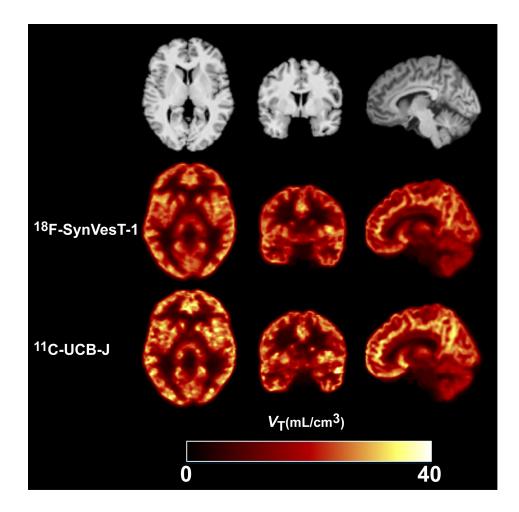


Figure 3:MR and co-registered parametric V_T images of ¹⁸F-SynVesT-1 (120 min PET data) and ¹¹C-UCB-J (90 min PET data) in the same subject, calculated with the 1TC model and the basis function method.

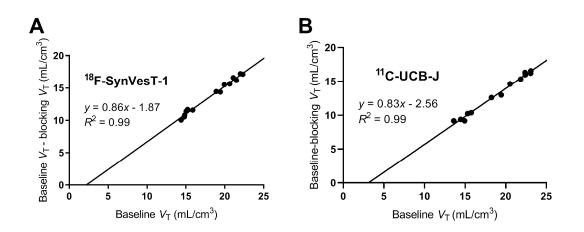


Figure 4:Occupancy plots for blocking scans with 20 mg/kg levetiracetam measured with ¹⁸F-SynVesT-1 (**A**) and ¹¹C-UCB-J (**B**) in the same subject. The estimated SV2A occupancy values were 86% in **A** and 83% in **B**.

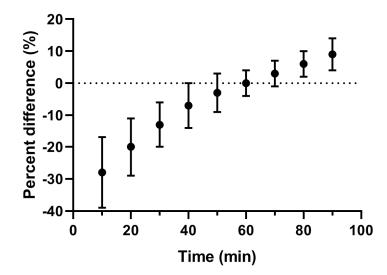


Figure 5:Mean and SD of percent differences between SUVR-1 from different time windows (30-min duration) and 1TC BP_{ND} values. Each data point was plotted at the beginning time of each window for SUVR-1 computation.

Tables

Parameter	¹⁸ F-SynVesT-1		¹¹ C-UCB-J	
	Baseline(n=8)	Blocking(n=4)	Baseline(n=4)	Blocking(<i>n</i> =4)
Age(y)	41±13	39±15	38±15	
Body weight(kg)	87.6±13.9	79.1±13.4	80.0±13.1	
Injected dose(MBq)	180±7	180±4	548±142	637±99
Molar activity at time of injection(MBq/nmol)	218±89	170±49	136±44	127±53
Injected mass(µg)	0.28±0.10	0.35±0.14	1.67±0.58	2.10±0.54

Table 1. Subject information and PET scan parameters

Data are mean±SD.

Regions	$V_{\rm T}$ (mL/cm ³)	$BP_{\rm ND}$	Minimum scan
Regions	(Baseline, <i>n</i> =8)	(Baseline,n=8)	time(min)
Putamen	19.3(14%)	4.5(11%)	30
Insula cortex	18.8(16%)	4.3(13%)	30
Temporal cortex	18.6(15%)	4.3(13%)	30
Parietal cortex	17.7(16%)	4.0(15%)	30
Amygdala	17.6(11%)	4.0(12%)	30
Occipital cortex	17.4(18%)	3.9(17%)	30
Ant. cingulate cortex	17.2(15%)	3.9(13%)	40
Frontal cortex	16.9(15%)	3.8(12%)	30
Caudate nucleus	15.3(15%)	3.3(12%)	30
Post. cingulate cortex	14.2(27%)	3.0(29%)	30
Thalamus	13.5(12%)	2.8(12%)	30
Cerebellum	13.0(12%)	2.7(15%)	60
Hippocampus	13.0(11%)	2.7(14%)	60
Globus pallidus	12.6(20%)	2.5(19%)	30
Centrum semiovale	3.5(11%)		30

Table 2. In vivo binding parameters of ¹⁸F-SynVesT-1 and minimum scan time for stable measurement, derived with the one-tissue compartment model

Regions	¹⁸ F-SynVesT-1(<i>n</i> =4)		¹¹ C-UCB-J(<i>n</i> =4)	
	Baseline	Blocking	Baseline	Blocking
Putamen	18.7(18%)	5.1(14%)	20.5(14%)	7.2(10%)
Insula cortex	18.4(21%)	4.9(13%)	19.6(15%)	6.9(11%)
Temporal cortex	18.8(20%)	4.6(8%)	20.4(15%)	6.8(7%)
Parietal cortex	18.1(21%)	4.3(6%)	20.1(15%)	6.6(7%)
Amygdala	17.4(13%)	4.7(12%)	19.1(13%)	7.0(13%)
Occipital cortex	17.7(24%)	4.2(6%)	19.5(20%)	6.3(4%)
Ant. cingulate cortex	17.6(21%)	4.8(16%)	19.0(20%)	7.0(15%)
Frontal cortex	17.1(18%)	4.3(8%)	18.8(12%)	6.4(8%)
Caudate nucleus	15.9(15%)	4.3(12%)	17.6(14%)	6.3(12%)
Post. cingulate cortex	15.9(32%)	4.0(9%)	16.9(28%)	5.9(19%)
Thalamus	13.3(14%)	4.0(13%)	14.5(12%)	5.7(6%)
Cerebellum	13.0(17%)	3.6(12%)	13.7(13%)	5.2(5%)
Hippocampus	13.2(13%)	3.8(11%)	14.1(10%)	5.5(10%)
Globus pallidus	11.9(27%)	4.2(14%)	12.5(18%)	5.6(6%)
Centrum semiovale	3.6(13%)	2.4(6%)	4.5(10%)	3.4(4%)

Table 3. Distribution volumes derived with the one-tissue compartment model for ¹⁸F-SynVesT-1 and ¹¹C-UCB-J under baseline and levetiracetam blocking conditions

%COV: coefficient of variation (inter-subject variability).

	Reference: 0	Reference: Centrum		Reference: V _{ND} from Lassen	
Regions	semiovale V _T		occupancy plot		
	¹⁸ F-SynVesT-1	¹¹ C-UCB-J	¹⁸ F-SynVesT-1	¹¹ C-UCB-J	
	(n = 4)	(<i>n</i> = 4)	(n = 4)	(<i>n</i> = 4)	
Putamen	4.26 (14%)	3.65 (24%)	7.53 (30%)	5.68 (26%)	
Insula cortex	4.18 (19%)	3.45 (27%)	7.40 (32%)	5.38 (26%)	
Temporal cortex	4.29 (19%)	3.64 (27%)	7.58 (31%)	5.61 (23%)	
Parietal cortex	4.10 (22%)	3.56 (28%)	7.29 (33%)	5.47 (21%)	
Amygdala	3.91 (14%)	3.31 (18%)	6.89 (24%)	5.22 (26%)	
Occipital cortex	3.99 (25%)	3.45 (33%)	7.11 (36%)	5.29 (25%)	
Ant. cingulate cortex	3.92 (16%)	3.29 (30%)	7.01 (33%)	5.20 (33%)	
Frontal cortex	3.83 (17%)	3.25 (23%)	6.82 (30%)	5.09 (23%)	
Caudate nucleus	3.48 (11%)	2.98 (22%)	6.24 (28%)	4.73 (26%)	
Post. cingulate cortex	3.43 (29%)	2.80 (37%)	6.3 (46%)	4.44 (35%)	
Thalamus	2.76 (14%)	2.29 (24%)	5.06 (28%)	3.72 (24%)	
Cerebellum	2.66 (20%)	2.10 (29%)	4.91 (30%)	3.41 (21%)	
Hippocampus	2.74 (16%)	2.17 (17%)	5.01 (25%)	3.58 (26%)	
Globus pallidus	2.34 (28%)	1.85 (38%)	4.47 (42%)	3.07 (30%	

 Table 1: Binding potentials of ¹⁸F-SynVesT-1 and ¹¹C-UCB-J with the one-tissue compartment model under baseline condition

 $\frac{18}{11} \text{F-SynVesT-1: } BP_{\text{ND}} \text{ using } V_{\text{T,CS}} = 0.64 \times (BP_{\text{ND}} \text{ using the Lassen } V_{\text{ND}}) - 0.33, R^2 = 1.00$