Novel PET Probes $^{18}$F-BCPP-EF and $^{18}$F-BCPP-BF for Mitochondrial Complex I: A PET Study in Comparison with $^{18}$F-BMS-747158-02 in Rat Brain

Hideo Tsukada, Shingo Nishiyama, Dai Fukumoto, Masakatsu Kanazawa, and Norihiro Harada

Central Research Laboratory, Hamamatsu Photonics K.K., Hamamatsu, Shizuoka, Japan

We developed novel PET probes, 2-tert-butyl-4-chloro-5-[6-(2-[18F]fluoroethoxy)-ethoxy]-pyridin-3-ylmethoxy]-2H-pyridazin-3-one (18F-BCPP-EF) and 2-tert-butyl-4-chloro-5-[6-(4-[18F]fluorobutoxy)-pyridin-3-ylmethoxy]-2H-pyridazin-3-one (18F-BCPP-BF), for quantitative imaging of mitochondrial complex I (MC-I) activity in the brain and preliminarily evaluated their properties in comparison with 18F-BMS-747158-02 (18F-BMS).

Methods: The affinity of 18F-BCPP-EF, 18F-BCPP-BF, and 18F-BMS to MC-I was analyzed using in vitro binding assays with 3H-dihydrorotenone and bovine myocyte submitochondrial particles. 18F-BCPP-EF, 18F-BCPP-BF, or 18F-BMS was intravenously injected into rats, and the uptake (standardized uptake value) in each organ was determined by dissection method. The effects of rotenone, a specific MC-I inhibitor, on the uptake of each probe were assessed by whole-body PET imaging in rats. Ischemic brain model rats were imaged using 18F-BCPP-EF. Results: The rank order of affinity to MC-I was 18F-BCPP-BF > 18F-BMS > 18F-BCPP-EF. The uptake of 18F-BCPP-EF and 18F-BMS was high in the heart, intermediate in brain, and low in muscle and bone 60 min after the injection. 18F-BCPP-BF provided increasing bone uptake with time after the injection. The uptake of 18F-BCPP-EF and 18F-BMS into the brain and heart was significantly decreased by predadministration of rotenone; however, the reduction degree of 18F-BCPP-EF was more pronounced than that of 18F-BMS. Rotenone did not affect 18F-BCPP-BF uptake in either the brain or the heart. 18F-BCPP-EF imaged the cortical ischemic neuronal damage without any disturbance by microglial activation even on day 7 when 18F-FDG showed high uptake in the damaged area.

Conclusion: The present study demonstrated that 18F-BCPP-EF could be a potential PET probe for quantitative imaging of MC-I activity and its ischemic damage in the living brain with PET.

Key Words: mitochondria complex I; brain; ischemia; 18F-BCPP-EF; PET

DOI: 10.2967/jnumed.113.125328

In normal mammalian cells, glucose is converted to pyruvate, followed by transformation into acetyl-coenzyme A by pyruvate dehydrogenase (Enzyme Commission Code [EC] 1.2.4.1) within the mitochondria, which is subsequently fed into the tricarboxylic acid cycle, ultimately producing energy via the electron transport system and oxidative phosphorylation. Oxidative phosphorylation may account for approximately 85%–90% of the glucose used in adult humans (1–3). Mitochondrial complex I (MC-I; nicotinamide adenine dinucleotide–ubiquinone oxidoreductase, EC 1.6.5.3) is the first and largest macromodule of the respiratory electron transport chain and oxidative phosphorylation. In contrast, activated inflammatory cells (macrophages and microglia) (4–7) and cancer cells (3,8,9) produce lactate from glucose via lactate dehydrogenase subunit A (EC 1.1.1.27), which is known as the Warburg effect or aerobic glycolysis, showing less than 5% glucose utilization in oxidative phosphorylation and a low contribution of the electron transport system for adenosine triphosphate (ATP) production.

18F-FDG PET is a well-established technique for the quantitative measurement of the regional metabolic rate of glucose in the brain (10). However, this technique measures only the first part of glucose metabolism in the phosphorylation of glucose to glucose-6-phosphate by hexokinase (EC 2.7.1.1). Therefore, 18F-FDG is taken up into not only normal tissues but also inflammatory regions observed in the subacute phase after transient focal ischemia in rodent models (11,12). Because recent reports indicated that several neurodegenerative disorders—schizophrenia, major depression, dementia (13,14), and autism (15)—induced neuroinflammation, 18F-FDG may lead to underestimation of the neurodegenerative damage in these diseases.

For the quantitative neuroimaging of normal tissues without interference by neuroinflammation, we proposed to apply a specific PET probe for MC-I, 18F-BMS-747158-02 (18F-BMS) (16), which was originally developed as a myocardial perfusion imaging agent (17,18). Because whole-body PET imaging indicated high uptake and long retention not only in heart but also in the brain (17), we hypothesized that 18F-BMS might be applicable for detecting neuronal damage in the living brain using PET. In fact, we demonstrated that 18F-BMS might be useful to detect ischemic neuronal damage at the subacute phase 7 d after ischemic insult (16), at which time markedly higher uptake of 18F-FDG, induced by microglial activation, was observed in the ischemic area (12,16). In parallel, we also found that 18F-BMS revealed relatively high nonspecific binding in the brain against the inhibition with rotenone, a specific MC-I inhibitor, in both in vitro and in vivo assessments (16). An ideal PET probe for MC-I should provide the specificity for MC-I with in vitro and in vivo assessments, an appropriate logD7.4 (distribution coefficient at pH 7.4) value, relatively long stability in plasma with rapid clearance, high brain uptake, high metabolic stability in the brain, and long retention with gradual elimination from the brain (19). According
to these criteria, we recently designed novel PET probes, 2-tert-butyl-4-chloro-5-{[6-(2-18F-fluoroethoxy)-ethoxy]-pyridin-3-ylmethoxy}-2H-pyridazin-3-one (18F-BCPP-EF) and 2-tert-butyl-4-chloro-5-{[6-(4-18F-fluorobutoxy)-pyridin-3-ylmethoxy]-2H-pyridazin-3-one (18F-BCPP-BF), for quantitative imaging of MC-I in the living brain (20). A preliminary in vitro binding assay indicated that 18F-BCPP-EF was more sensitive to rotenone than the others, with significantly lower nonspecific binding (16,20). In the present study, the properties of 18F-BCPP-EF and 18F-BCPP-BF were further evaluated regarding the affinity to MC-I using bovine cardiomyocyte submitochondrial particles (SMP), the rat whole-body kinetics by a tissue dissection method, and the effects of rotenone on MC-I binding in the rat brain and heart using small-animal PET. Furthermore, the capability of 18F-BCPP-EF to detect neuronal damage was evaluated in a rat model of ischemic brain damage.

MATERIALS AND METHODS

Animals and Chemicals

The following experiments were approved by the Ethical Committee of the Central Research Laboratory, Hamamatsu Photonics. Male Sprague–Dawley rats (age, 8 wk; weight, 260–280 g) (Japan SLC) were used. 3H-dihydrorotenone was obtained from American Radiolabeled Chemicals. Halothane and chloral hydrate were purchased from Takeda Pharmaceutical and Sigma-Aldrich, respectively. Rotenone was from MP Biochemicals LLC. Rose bengal and rabbit anti-Iba1 polyclonal antibody were from Wako Pure Chemical Industry. Mouse anti-NeuN monoclonal antibody and EnVision were obtained from Millipore and DAKO, respectively. 4,7,13,16,21,24-Hexaoxa-1,10-diazacyclodecane (8,8,8)hexa-oxane (K[2,2,2]) was obtained from Aldrich. Mannose triflate, the precursor of 18F-BMS-747158-02 (18F-BMS), and its standard compounds were obtained from NARD Institute.

In Vitro Binding Assay

For analyses of the affinity of BCPP-EF, BCPP-BF, and BMS to MC-I, an in vitro binding assay was conducted using 3H-dihydropyrone and bovine cardiomyocyte SMP prepared according to a previous report (21). In competition experiments, the radioligand concentration was fixed at 4.5 nM, and variable concentrations of each testing compound were included in dimethyl sulfoxide to add to test tubes containing SMP (45 μg/mL protein) and incubated at 22°C for 30 min. After the incubation, the solutions were filtrated through filter papers (GF/C; Whatman) using a cell harvester (M-30R; Brandel) and washed 3 times with 50 mM Tris/HCl buffer solution (pH 7.4), and then protein-bound radioactivities remaining in the filter papers were measured using a liquid scintillation counter (1300; PerkinElmer). Specific binding was plotted against the concentration of each testing compound to determine the 50% inhibition values, which were converted to the inhibition constant (Ki) using the Cheng and Prusoff equation (22).

PET Ligand Syntheses

18F was produced by 18O(p, n) 18F nuclear reaction using the cyclotron (HM-18; Sumitomo Heavy Industry, Osaka, Japan) at Hamamatsu Photonics PET center. Labeled compounds were synthesized using a modified CUPID system (Sumitomo Heavy Industry). High-performance liquid chromatography (HPLC) analyses of labeled compounds were performed on a GL-7400 low-pressure-gradient HPLC system (GL Sciences, Inc.) with a radioactivity detector (RLC-700; Hitachi Aloka Medical, Inc.). 18F-BCPP-EF and 18F-BCPP-BF were radiolabeled as reported previously (Supplemental Fig. 1; supplemental materials are available at http://jnm.snmjournals.org) (20). Radioactivity yields, radiochemical purities, and specific radioactivities of 18F-BCPP-EF and 18F-BCPP-BF were 5.1 ± 0.9 and 3.7 ± 1.1 GBq (mean ± SD, n = 5), 30.8 ± 4.2 and 16.3% ± 2.7%, 99.1 ± 0.7 and 99.6% ± 0.6%, and 139.6 ± 37.0 and 111.8 ± 40.1 GBq/μmol, respectively.

18F-BMS was labeled as reported previously (17,18). Radioactivity yield, radiochemical yield, radiochemical purity, and specific radioactivity were 4.7 ± 1.0 GBq, 19.1% ± 3.7%, 99.6% ± 0.7%, and 67.2 ± 17.6 GBq/μmol, respectively.

18F-FDG was produced by nucleophilic 18F-fluorination of mannose triflate after basic hydrolysis of 2-18F-fluoro-1,3,4,6-tetra-α-acetamido-1-glucose according to a method described previously (23).

Tissue Dissection Assay

Three rats per each time point were used in this assessment. Five megabecquerels of 18F-BCPP-EF, 18F-BCPP-BF, or 18F-BMS were injected into each rat through the tail vein. The animals were sacrificed by decapitation under chloral hydrate (400 mg/kg, intraperitoneally) anesthesia at 1, 5, 10, and 30 min after the injection; samples of blood, heart, lung, liver, kidney, spleen, muscle, bone, small intestine, large intestine, pancreas, and brain were rapidly removed; the weight and radioactivity were measured using a γ counter (1480 WIZARD, Perkin Elmer); and standardized uptake values (SUVs) were calculated.

For plasma metabolic analyses, the blood samples were obtained at 1, 5, 10, 30, and 60 min after the injection; centrifuged to separate plasma; and weighed. The radioactivity of the samples was measured, and methanol was added to plasma samples (sample/methanol = 1/1), followed by centrifugation. The obtained supernatants were developed with thin-layer chromatography plates (AL SIL GUV; Whatman) with a mobile phase of ethyl acetate. The ratio of unmetabolized fraction was determined using a phosphoimaging plate (FLA-7000, Fuji Film) to calculate unmetabolized ratios of PET probes.

For brain metabolite analyses, the brain samples obtained at 5, 30, and 60 min after the injection were minced, supplemented with ice-cooled EtOH (2.0 mL), and homogenized (Polytron PT300; Kinematica AG) for 1 min. Aliquots of the homogenate (0.4 mL) were transferred to a tube and centrifuged. The resulting supernatants were separated and analyzed in the same manner as plasma samples.

Immunohistochemical assessments were performed with brains sampled at day 7 after ischemic insults using rabbit anti-Iba1 polyclonal antibody and mouse anti-NeuN monoclonal antibody as reported previously (12,16).

PET Imaging

Vehicle alone or rotenone, a specific MC-I inhibitor, at a dose of 0.1 mg/kg in 10 mL of vehicle (N,N-dimethylformamide/polyethylene glycol 400/saline = 1/1/2), was infused to rats through the tail vein cannula for 1 h, then 18F-BCPP-EF, 18F-BCPP-BF, or 18F-BMS was injected as a bolus for PET measurement. For PET imaging of ischemic neuronal damage, 18F-BCPP-EF or 18F-FDG was injected 1 (day 1) and 7 d (day 7) after the ischemic insult with photochemically induced thrombosis (PIT) as reported previously (12,16,24). PET scans were conducted with a high-resolution small-animal PET scanner (ClairviewPET, Shimadzu Corp. (25)) as described previously (12,16). Dynamic images and summation images of 18F-BCPP-EF, 18F-BCPP-BF, and 18F-BMS from 10 to 30 min, and of 18F-FDG from 40 to 60 min after the injection, were reconstructed, and created SUV images. Body temperature in rats was monitored to control using a heat pad during PET measurement. Because of technical limitation, physiologic vital signs could not be monitored.

Regions of interest (ROIs) for rotenone effect assessment were placed on PET images in the cortex, striatum, and cerebellum with the aid of a rat brain atlas (26). In the ischemic PET studies, ROIs for the
perinfarct and infarct core areas were defined as the ischemic left hemisphere region showing 40%–60% (perinfarcted region) and less than 40% (core region), respectively, of $^{18}$F-BCPP-EF uptake in the corresponding intact side, and they were applied to a normal condition (16). The same ROIs were set on the intact right hemisphere of each condition.

**Statistical Analysis**

Results are expressed as mean ± SD. Comparisons between conditions were performed using paired, 2-tailed Student $t$ tests. A probability level of less than 5% ($P < 0.05$) was considered to indicate statistical significance.

**RESULTS**

We determined 50% inhibition values of $^{18}$F-BCPP-EF, $^{18}$F-BCPP-BF, and $^{18}$F-BMS for MC-I to calculate their $K_i$ values in the in vitro assay using bovine cardiomyocyte SMP and $^3$H-dihydrodrotenone. $^{18}$F-BCPP-BF had the highest affinity ($K_i = 0.70$ nM), $^{18}$F-BMS was intermediate (0.95 nM), and $^{18}$F-BCPP-EF showed the lowest affinity (2.31 nM) for MC-I of bovine cardiomyocyte (Supplemental Fig. 2).

Radiosyntheses of $^{18}$F-BCPP-EF, $^{18}$F-BCPP-BF, and $^{18}$F-BMS were conducted, and the biodistribution of these PET probes was determined by a tissue dissection method in rats (Fig. 1). The kinetics of these 3 probes in whole blood had a U shape, and thus the radioactivity levels decreased up to 10 min after the injection, followed by a gradual increase. Although precise mechanisms remain unclear, the decrease might be attributable to their release from organs with high uptake of PET probes at early times, such as the heart. $^{18}$F-BCPP-EF showed rapid uptake that was retained up to 30 min after injection, followed by a decline thereafter in the heart, kidney, small and large intestines, and pancreas. The liver exhibited rapid uptake and then remained constant throughout the study. In contrast, the brain, lung, spleen, and bone revealed gradual elimination just after the injection (Fig. 1A). At 60 min after the injection, the distribution of $^{18}$F-BCPP-EF was the highest in the heart; high in the kidney and liver; intermediate in the brain, pancreas, and small intestine; and low in the spleen, large intestine, lung, muscle, and bone (Fig. 1A).

The uptake levels of $^{18}$F-BCPP-BF in each organ were slightly lower than those of others. At 60 min after the injection, the distribution of $^{18}$F-BCPP-BF was the highest in the heart; high in the kidney; intermediate in the liver, brain, pancreas, small intestine, and bone; and low in the spleen, large intestine, lung, and muscle (Fig. 1B). The kidney, small and large intestines, pancreas, and liver showed rapid uptake of $^{18}$F-BCPP-BF that was retained up to 10 min after injection, followed by a decline thereafter. The lung and spleen revealed gradual elimination just after the injection. In contrast, the uptake in the heart and bone continuously increased with time after $^{18}$F-BCPP-BF injection (Fig. 1B).

The patterns of time–activity curves of $^{18}$F-BMS uptake were generally the same as those of $^{18}$F-BCPP-EF, except in the liver, in which the uptake of $^{18}$F-BMS increased up to 30 min after injection, followed by a gradual decline (Fig. 1C).

Peak values of brain-to-blood (15.5) and heart-to-blood (63.0) ratios of $^{18}$F-BCPP-EF were observed 10 min after the injection (Table 1), whereas the ratios of $^{18}$F-BCPP-BF continuously increased up to 60 min after the injection, showing 14.9 and 83.7, respectively (Table 1). These peak values of $^{18}$F-BCPP-EF and $^{18}$F-BCPP-BF were significantly higher than those of $^{18}$F-BMS (11.5 and 56.2, respectively, Table 1).

The metabolic analyses of $^{18}$F-BCPP-EF and $^{18}$F-BCPP-BF revealed the gradual degradation to polar metabolites at a relatively slow rate, with 17.3% and 18.4%, respectively, remaining as each parent compound 30 min after the injection (Table 2). $^{18}$F-BMS was metabolized more rapidly than others, showing only 7.5% at 30 min after the injection (Table 2). In contrast, the metabolic profiles of $^{18}$F-BCPP-EF, $^{18}$F-BCPP-BF, and $^{18}$F-BMS indicated greater stability in the brain than those in the plasma, showing 90.5%, 97.1%, and 87.2%, respectively, 30 min after the injection (Table 2).

Small-animal PET imaging of rats with $^{18}$F-BCPP-EF, $^{18}$F-BCPP-BF, and $^{18}$F-BMS revealed high uptake into the brain (SUV = 2.9, 2.0, and 3.0, respectively) (Figs. 2 and 3) and heart (SUV = 10.2, 8.6, and 10.6, respectively, at 30 min after injection).
Rats were intravenously injected with 5 MBq of $^{18}$F-BCPP-EF, $^{18}$F-BCPP-BF, or $^{18}$F-BMS via tail vein and sacrificed at 1, 5, 10, 30, and 60 min after injection to obtain tissue samples for uptake ratio analyses. All values are expressed as mean ± SD for 3 animals.

![Fig 4](image)

Under the vehicle condition, the radioactivity level of $^{18}$F-BCPP-EF in the brain showed rapid uptake and gradual decrease with time just after the injection (Fig. 3A), whereas $^{18}$F-BCPP-BF remained constant throughout the PET measurements up to 60 min after the injection (Fig. 3B). $^{18}$F-BMS remained constant for 30 min after the injection, followed by gradual reduction (Fig. 3C). In the heart, $^{18}$F-BCPP-EF exhibited slow accumulation up to 30 min after the injection, followed by slight washout (Fig. 4A), whereas $^{18}$F-BCPP-BF constantly increased up to 60 min after the injection (Fig. 4B). $^{18}$F-BMS exhibited slow accumulation, followed by its maintenance at a constant level at 20 min and later (Fig. 4C). With preadministration of rotenone (0.1 mg/kg/h), the uptake levels of $^{18}$F-BCPP-EF and $^{18}$F-BMS into the brain showed a significant reduction, and the degree of reduction of $^{18}$F-BCPP-EF (Figs. 2A and 3A) was much more pronounced than that of $^{18}$F-BMS (Figs. 2C and 3C, lower). In the heart, rotenone administration induced reduction of $^{18}$F-BCPP-EF (Figs. 2A and 4A) and $^{18}$F-BMS uptake (Figs. 2C and 3C), with faster elimination rates than in the vehicle condition. No significant reduction of $^{18}$F-BCPP-BF uptake was observed in either brain or heart (Figs. 2B, 3B, and 4B). The SUVs of $^{18}$F-BCPP-EF and $^{18}$F-BMS, assessed 30 min after the injection, were suppressed by the preadministration of rotenone (0.1 mg/kg) to 65.4% and 85.6% of the vehicle condition in the brain (Figs. 3A and 3C) and 53.7% and 81.7% of the vehicle condition in the heart, respectively (Figs. 3A and 3C).

At day 1 after ischemic insult, PIT induced a decrease in the uptake of $^{18}$F-FDG (Fig. 5, left, and Fig. 6, left) in the infarct region of the left ischemic hemisphere; however, there was higher uptake in the ischemic region than in the corresponding region in the intact side at day 7 (Fig. 5, left, and Fig. 6, left), which overlapped with high immunoreactivity to anti-Iba 1 antibody (Fig. 7, upper). In contrast, PET imaging with $^{18}$F-BCPP-EF demonstrated a significant reduction in the infarct region of the left ischemic hemisphere.

**TABLE 1**

<table>
<thead>
<tr>
<th>Probe</th>
<th>1 min</th>
<th>5 min</th>
<th>10 min</th>
<th>30 min</th>
<th>60 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>$^{18}$F-BCPP-EF</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Brain to blood</td>
<td>7.643 ± 0.495</td>
<td>15.058 ± 1.358</td>
<td>15.525 ± 2.178</td>
<td>7.866 ± 0.453</td>
<td>4.411 ± 0.435</td>
</tr>
<tr>
<td>Brain to plasma</td>
<td>5.280 ± 0.217</td>
<td>11.703 ± 1.361</td>
<td>11.879 ± 1.559</td>
<td>5.941 ± 0.502</td>
<td>3.328 ± 0.642</td>
</tr>
<tr>
<td>$^{18}$F-BCPP-BF</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Brain to plasma</td>
<td>3.926 ± 0.319</td>
<td>10.986 ± 1.042</td>
<td>10.039 ± 0.786</td>
<td>11.658 ± 2.918</td>
<td>11.112 ± 2.304</td>
</tr>
<tr>
<td>Heart to blood</td>
<td>19.617 ± 0.680</td>
<td>65.826 ± 5.418</td>
<td>72.429 ± 1.703</td>
<td>76.204 ± 3.909</td>
<td>83.791 ± 18.636</td>
</tr>
<tr>
<td>$^{18}$F-BMS</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Brain to blood</td>
<td>5.483 ± 1.289</td>
<td>11.318 ± 1.328</td>
<td>11.568 ± 2.916</td>
<td>6.141 ± 0.399</td>
<td>3.169 ± 0.124</td>
</tr>
<tr>
<td>Brain to plasma</td>
<td>4.291 ± 1.030</td>
<td>8.804 ± 1.116</td>
<td>9.039 ± 2.329</td>
<td>4.657 ± 0.128</td>
<td>2.511 ± 0.119</td>
</tr>
<tr>
<td>Heart to blood</td>
<td>25.195 ± 4.438</td>
<td>56.286 ± 5.319</td>
<td>53.204 ± 8.714</td>
<td>29.393 ± 2.996</td>
<td>14.379 ± 0.442</td>
</tr>
<tr>
<td>Heart to plasma</td>
<td>19.720 ± 3.658</td>
<td>43.831 ± 5.400</td>
<td>41.555 ± 6.982</td>
<td>22.268 ± 1.422</td>
<td>11.385 ± 0.172</td>
</tr>
</tbody>
</table>

**TABLE 2**

<table>
<thead>
<tr>
<th>Probe</th>
<th>1 min</th>
<th>5 min</th>
<th>10 min</th>
<th>30 min</th>
<th>60 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>$^{18}$F-BCPP-EF</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Brain</td>
<td>ND</td>
<td>98.5 ± 0.1</td>
<td>ND</td>
<td>90.5 ± 0.2</td>
<td>84.4 ± 0.1</td>
</tr>
<tr>
<td>Plasma</td>
<td>84.5 ± 3.2</td>
<td>46.1 ± 8.0</td>
<td>40.4 ± 4.8</td>
<td>17.3 ± 2.6</td>
<td>8.3 ± 0.9</td>
</tr>
<tr>
<td>$^{18}$F-BCPP-BF</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Brain</td>
<td>ND</td>
<td>99.3 ± 0.1</td>
<td>ND</td>
<td>97.1 ± 0.1</td>
<td>95.3 ± 0.1</td>
</tr>
<tr>
<td>Plasma</td>
<td>92.5 ± 0.3</td>
<td>69.5 ± 1.6</td>
<td>39.8 ± 2.0</td>
<td>18.4 ± 3.5</td>
<td>12.1 ± 3.6</td>
</tr>
<tr>
<td>$^{18}$F-BMS</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Brain</td>
<td>ND</td>
<td>97.9 ± 0.3</td>
<td>ND</td>
<td>87.2 ± 0.5</td>
<td>76.3 ± 1.2</td>
</tr>
<tr>
<td>Plasma</td>
<td>89.9 ± 1.5</td>
<td>38.9 ± 3.5</td>
<td>24.3 ± 9.3</td>
<td>7.5 ± 1.8</td>
<td>2.9 ± 0.7</td>
</tr>
</tbody>
</table>

Rats were intravenously injected with 5 MBq of $^{18}$F-BCPP-EF, $^{18}$F-BCPP-BF, or $^{18}$F-BMS via tail vein and sacrificed at 1, 5, 10, 30, and 60 min after injection to obtain brain and plasma for metabolic analyses. All values are expressed as mean ± SD for 3 animals. ND = not determined.
hemisphere from day 1 and later after the ischemic insult induced by PIT (Fig. 5, right, and Fig. 6, right). At day 1, the damaged areas determined by 18F-BCPP-EF as core regions were well correlated with those measured by 2,3,5-triphenyltetrazolium chloride staining (Supplemental Fig. 3). At day 7, the low uptake area determined with 18F-BCPP-EF (Fig. 5, right) corresponded to the area with low immunoreactivity to anti-NeuN antibody (Fig. 7, lower).

DISCUSSION

The present study evaluated the capability of 18F-BCPP-EF and 18F-BCPP-BF as PET probes for imaging the MC-I activity in the living rat brain in comparison with a previously reported probe 18F-BMS (17,18).

The accumulation of 18F-BMS was reported to depend on cellular MC-I activity, which is the first component of 4 electron transport complexes in the inner mitochondrial membrane (17,18). By an in vitro assay in a monolayer of neonatal rat cardiomyocytes, it was confirmed that 18F-BMS uptake was inhibited by rotenone, a specific MC-I inhibitor (17). 18F-BMS was also expected to show uptake in the brain because of the high density of mitochondria, not only in the heart but also in the brain. Our previous results demonstrated that 18F-BMS might be a useful PET probe to detect the ischemic damages of MC-I not only at the acute but also at the subacute phase, at which time the remarkable microglial activation in the ischemic damaged area was determined by PET with 11C-(R)-PK11195 and 18F-FDG as well as immunohistochemical analysis with anti-Iba1 antibody (12,16). Because activated inflammatory cells (macrophages and microglia) exclusively produce ATP through the enhanced glycolysis with a low contribution of the electron transport system including MC-I for ATP production, these cells need more glucose to survive and also uptake more 18F-FDG than normal neuronal and glial tissues. These metabolic properties resulted in the apparently high uptake of 18F-FDG in damaged areas in which activated inflammatory cells accumulated (12,16). To measure the viability of neuronal and glial tissues by PET and to avoid underestimation of damaged areas as observed by 18F-FDG, PET targeting mitochondrial function was expected to be useful (8,16).

In vitro assessment with living brain slices and in vivo assessment using small-animal PET suggested that 18F-BMS has relatively high nonspecific binding in brain tissues against inhibition with rotenone (16). Therefore, we tried to modify the chemical structure of 18F-BMS to induce lower lipophilicity (20). As reported previously, the lipophilicity indices of logD7.4 were 3.03, 4.27, and 3.69 for 18F-BCPP-EF, 18F-BCPP-BF, and 18F-BMS, respectively (20), which are within the appropriate range of logD7.4 for blood–brain barrier penetration (19). It was expected that, if the brain uptake was exclusively lipophilicity-driven, the rank order of brain uptake should have been high in 18F-BCPP-BF, intermediate in 18F-BMS, and low in 18F-BCPP-EF. However, the assessed results indicated that 18F-BCPP-BF with the highest lipophilicity showed the lowest brain uptake. The prospected transport mechanisms of these compounds need to take into account more complex systems.

The specificity of 18F-BCPP-EF binding to MC-I was confirmed by preadministration of rotenone. Although a dose-escalation

FIGURE 2. Effects of preadministration of rotenone on uptake of 18F-BCPP-EF (A), 18F-BCPP-BF (B), and 18F-BMS (C) in rats imaged by small-animal PET and X-CT. After continuous infusion of vehicle or rotenone at dose of 0.1 mg/kg/h for 1 h, PET scanning was conducted for 60 min after injection of each PET probe. Summation PET data from 10 to 30 min for A, B, and C were reconstructed for SUV images, and then PET images were superimposed on individual X-CT images.

FIGURE 3. Effects of preadministration of rotenone on time–activity curves of 18F-BCPP-EF (A), 18F-BCPP-BF (B), and 18F-BMS (C) in rat brain. PET scans were conducted as described in Figure 2, and then summation data from 10 to 30 min were reconstructed for SUV images. ROIs for brain were set on reconstructed PET images to obtain time–activity curves of each PET probe in these regions. ○ = cortex-vehicle; ● = cortex-rotenone; △ = striatum-vehicle; ▲ = striatum-rotenone; ■ = cerebellum-vehicle; ▧ = cerebellum-rotenone.
study was impossible because of its lethal effects on the cardiac function, a significant reduction of uptake of $^{18}$F-BCPP-EF and $^{18}$F-BMS was observed in the brain and heart, even at a relatively low dose of 0.1 mg/kg/h. In contrast, preadministration of rotenone induced no significant reduction of $^{18}$F-BCPP-BF in both brain and heart, because of its high Ki value to MC-I determined using SMP and $^3$H-dihydrotetenone. The high affinity of $^{18}$F-BCPP-BF to MC-I was also suggested by its nonreversible (brain) and accumulated-type kinetics (heart) in rat using animal PET imaging, the kinetics of which are not always suitable for quantitative imaging analysis of brain function using PET (27,28).

For $^{18}$F-labeled PET probes, $^{18}$F-BCPP-EF indicated minimal defluorination in plasma. In contrast, $^{18}$F-BCPP-BF exhibited the increasing accumulation of radioactivity in bone with time after the injection, suggesting the instability with defluorination. It was reported to be a common mechanism of $^{18}$F-fluoride formation that P-450–mediated hydroxylation occurred at $^{18}$F-fluorine–substituted carbon, followed by spontaneous elimination (29,30). In contrast, the substitution of oxygen at β carbon decreased the reactivity toward hydroxylation, leading to slower $^{18}$F-fluoride release (31). Thus, the differences of metabolic stability among these PET probes against P-450–mediated hydroxylation could be considered attributable to the chemical structural differences.

$^{18}$F-BCPP-EF could detect impaired MC-I activity in the living brains of the ischemic rat model, revealing the lower uptake of $^{18}$F-BCPP-EF in the ischemia-damaged area than intact area. As observed at day 7, $^{18}$F-FDG showed high uptake in the ischemia-damaged region, in which $^{18}$F-BCPP-EF revealed low uptake. These results strongly suggested that this reduction was related to MC-I specific binding, not to a simply reduced delivery by ischemic insult. The reactive oxygen species (ROS) play a role in cell injury associated with cortical ischemia and reperfusion. The PIT model has been considered as the most relevant to the pathologic cascade occurring after clinical infarction among the experimental ischemic rodent models (24). Thus, the reperfusion–injurylike phenomenon, occurring after clinical infarction, could be involved in the progress of more severe brain damage than a permanent occlusion model (32,33), suggesting that reperfusion-related ROS production may in part contribute to neuronal damage of reperfusion injury. Mitochondria are considered the main intracellular source of ROS and also the main target of

FIGURE 4. Effects of preadministration of rotenone on time–activity curves of $^{18}$F-BCPP-EF (A), $^{18}$F-BCPP-BF (B), and $^{18}$F-BMS (C) in rat heart. PET scans were conducted as described in Figure 2, and then summation data from 10 to 30 min were reconstructed for SUV images. ROIs for heart (myocardium) were set on reconstructed PET images to obtain time–activity curves of each PET probe in these regions. ○ = vehicle; ● = rotenone.

FIGURE 5. Temporal changes in PET images of $^{18}$F-FDG and $^{18}$F-BCPP-EF in rat brains of PIT model. PET scans were conducted for 60 min with each PET probe before (normal), 1 (day 1), and 7 d (day 7) after PIT operation. #P < 0.01 vs. corresponding normal condition. $^\#P < 0.01$ vs. corresponding day 1 condition.

FIGURE 6. Temporal changes in uptake (SUV ratio) of $^{18}$F-FDG and $^{18}$F-BCPP-EF in rat brains of PIT model. PET scans were conducted for 60 min with each PET probe before (normal), 1 (day 1), and 7 d (day 7) after PIT operation. #P < 0.01 vs. corresponding normal condition. $^\#P < 0.01$ vs. corresponding day 1 condition.
The present preliminary results demonstrated that $^{18}$F-BCPP-EF has the capability to detect neuronal degeneration as impaired MC-I activity in the living rat brains of an ischemic brain model.

DISCLOSURE

The costs of publication of this article were defrayed in part by the payment of page charges. Therefore, and solely to indicate fact, this article is hereby marked “advertisement” in accordance with 18 USC section 1734. No potential conflict of interest relevant to this article was reported.

REFERENCES


22. Cheng Y, Prusoff WH. Relationship between the inhibition constant (K) and the concentration of inhibitor which causes 50 percent inhibition (IC$_{50}$) of an enzymatic reaction. *Biochem Pharmacol.* 1973;22:3099–3108.


ACKNOWLEDGMENTS

We gratefully acknowledge the technical assistance provided by Takeharu Kakiuchi, Shigeyuki Yamamoto, Yuko Kato, and Teruyo Hosoya.


