Small-animal models are crucial to gain insights in the complex recovery mechanisms of liver function during liver regeneration. 99mTc-Mebrofenin hepatobiliary scintigraphy (HBS) has been introduced for noninvasive assessment of liver function in the clinical setting as well as in experimental research. However, HBS is restricted to planar modalities in small animals because hepatic kinetics are generally too fast for SPECT acquisition. 99mTc-DTPA-galactosyl serum albumin (where DTPA is diethylenetriaminepentaacetic acid) (99mTc-GSA) scintigraphy is an alternative, receptor-mediated, noninvasive liver function test. After hepatic uptake, 99mTc-GSA remains trapped in the liver, which readily enables additional SPECT for the assessment of both liver function and liver functional volume within one test. In this study we evaluated the use of 99mTc-GSA scintigraphy combined with SPECT for the assessment of liver function and liver functional volume in normal and regenerating rat livers. **Methods:** The reproducibility of 99mTc-GSA scintigraphy and SPECT was investigated by repeated measurements within the same rat. For the assessment in a regenerating liver, 99mTc-GSA scintigraphy with SPECT was performed on 1, 3, 5, and 7 d (n = 6 rats per time point) after 70% partial hepatectomy (PH). **Results:** The correlation between repeated 99mTc-GSA measurements was strong (r = 0.75, P = 0.019). In normal rat livers, there was a strong, significant correlation between liver functional volume and conventional liver volume (r = 0.93; < 0.0001). The correlation between 99mTc-GSA uptake and liver volume was moderate (r = 0.62, P = 0.043). During the regeneration process, 99mTc-GSA uptake was significantly lower compared with both liver volume (P < 0.001) and liver functional volume (P < 0.001), when expressed as a percentage of baseline levels. There was a strong correlation between liver functional volume and conventional liver volume in the regenerating liver (r = 0.92, P < 0.0001). **Conclusion:** 99mTc-GSA scintigraphy combined with SPECT is a feasible, noninvasive method to assess hepatic functional volume in normal rat liver as well as in the regenerating rat liver. However, the hepatic 99mTc-GSA uptake as a liver function test seems to underestimate hepatic regeneration in comparison to liver volume.

**Key Words:** liver function; liver functional volume; liver regeneration; 99mTc-GSA scintigraphy


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assessment of liver function has been introduced using 99mTc-mebrofenin hepatobiliary scintigraphy (HBS) in both acute and chronic experimental animal models (8–11). However, scintigraphic imaging acquisition is restricted to planar modalities because hepatic kinetics of 99mTc-mebrofenin in small animals are too fast for (pinhole) SPECT acquisition. An alternative radiopharmaceutical for liver function scintigraphy is 99mTc-DTPA-galactosyl serum albumin (where DTPA is diethylenetriaminepentaacetic acid) (99mTc-GSA) (12). Although it is not registered for clinical use outside Japan, 99mTc-GSA is available for preclinical investigation elsewhere. 99mTc-GSA scintigraphy is based on the hepatic pharmacokinetics of asialoglycoproteins (ASGP). ASGP is a serum galactose-terminated glycoprotein, which binds to the ASGP receptor and is subsequently internalized via receptor mediated endocytosis. The ASGP receptor is expressed in large quantities on the hepatocyte sinusoidal surface (13) and its expression reflects the severity of chronic liver disease, such as cirrhosis (14–16). 99mTc-GSA was developed as a synthetic asialoglycoprotein to visualize and quantify its hepatic binding to the ASGP receptor. After intravenous injection and hepatic uptake, 99mTc-GSA remains trapped in the liver for at least 30 min (17). This would potentially enable pinhole SPECT acquisition in small laboratory animals for the assessment of both liver function and liver functional volume.

The aim of this study was to evaluate the use of 99mTc-GSA scintigraphy as a liver function imaging technique in combination with 99mTc-GSA SPECT for additional assessment of liver functional volume in a normal and regenerating rat liver.

MATERIALS AND METHODS

Animals and Reagents

Male Wistar rats (Harlan) were acclimatized for 1 wk under standardized laboratory conditions in a temperature-controlled room with a 12 h-light/12-h dark cycle on standard rat chow and water ad libitum. 99mTc-GSA was kindly provided by Nihon MediPhysics. This study was approved by the Animal Ethics and Welfare Committee of the Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands.

Camera Design

For imaging of 99mTc-GSA uptake in the rat liver, a γ-camera (ARC 3000; Philips) available in a dedicated animal care facility was equipped with a pinhole collimator fitted with a 3-mm tungsten insert. The pinhole collimator was facing up. On the detector, a mechanical support was mounted in which the animal is fixed in an acrylic cylinder positioned exactly above the pinhole collimator. The mechanical support was designed to place the midline of the cylinder exactly in the middle of the pinhole. The position of the animal is adjustable in the axial dimension. Furthermore, the distance from the cylinder to the pinhole aperture is adjustable. Therefore, this gantry permits optimal pinhole scintigraphy of anterior projections of rats, standardizing magnification and orientation (18). The γ-camera is interfaced to a Hermes acquisition station (Nuclear Diagnostics).

Scintigraphy and Interpretation

The animals were sedated with a mixture of O2/N2O (1:1 v/v, 2 L/min) and isoflurane (2%–3% Florene; Abbott Laboratories Ltd.). Once anesthetized, a bolus of 40 MBq 99mTc-GSA in 0.3 mL saline was injected in the tail vein. The animals were scanned on injection of the radiopharmaceutical in anterior position with the liver and the mediastinum in the field of view (FOV). For dynamic 99mTc-GSA scintigraphy, images were obtained for 30 min (10 min at 5 s per frame and 20 min at 60 s per frame) at the 140-keV 99mTc peak with a 20% window in a 128 × 128 matrix. Data were processed on a Hermes workstation (Nuclear Diagnostics). In clinical studies, liver function, measured by 99mTc-GSA scintigraphy, is expressed by many different, sometimes complex, parameters. The most commonly used parameters in planar, dynamic 99mTc-GSA scintigraphy are the hepatic uptake ratio of 99mTc-GSA (LHL15) and the blood clearance ratio (HH15) (12,19). We used the 99mTc-GSA liver uptake based on a technique described by Ekman et al. (20). The algorithm was adapted for rat scintigraphy based on the faster hepatic uptake of 99mTc-GSA in rats (21). Regions of interest (ROIs) were drawn around the liver, heart, and large vessels within the mediastinum (serving as blood pool) and around the total FOV (indicative of total activity). The liver ROI was drawn automatically on a threshold-based algorithm using 20% of the maximum liver value on a summed image of the first 2.5 min of the acquisition as cutoff. Three different time–activity curves were generated based on the liver, blood pool, and total FOV. Liver uptake was calculated in %/min, based on these 3 parameters. Calculations of hepatic 99mTc-GSA uptake were performed using scanned radioactivity values acquired between 60 and 180 s after injection to make sure that calculations were made during a phase of homogeneous distribution of the agent in the blood pool.

SPECT acquisition was performed 30 min after intravenous injection of 99mTc-GSA. Fifty projections (128 × 128 matrix) were made in a 360° orbit. The acquisition time per projection was 25 s. SPECT reconstruction was performed with a Hermes application program adapted to pinhole SPECT, using filtered backprojection (Fig. 1) (18). A Butterworth postreconstruction filter (order 5; 0.8 cycle/cm) was applied. After completion of the reconstruction, the outline extraction method with a threshold of 40% of the maximal voxel count value was applied for determination of the liver functional volume (22).
Reproducibility
Nine rats were anesthetized by inhalation with a mixture of O₂/N₂O (1:1 v/v, 2 L/min) and isoflurane 2%–3% and scanned on 2 separate days with a 2 d time interval, to assess the reproducibility of scintigraphic liver function testing and intrasubject variation. Subsequently, a SPECT acquisition of 25 min was made. After the final scintigraphy, livers were removed and the liver volume and weight were determined. Conventional liver volume was calculated as previously described (23).

Surgical Procedure
Two groups of rats in different weight categories, group A; 250–300 g (n = 6) and group B; 300–350 g (n = 6), were used to compare liver volume and liver functional volume measured by 99mTc-GSA SPECT. For the assessment of liver regeneration, 30 rats were randomized into 5 experimental groups. In a control group (n = 6), 99mTc-GSA scintigraphy with SPECT was performed to calculate normal (baseline) 99mTc-GSA uptake and liver functional volume. Subsequently, rats were sacrificed to determine baseline liver weight and liver volume. For the assessment of liver regeneration, rats were anesthetized 1 (n = 6), 3 (n = 6), 5 (n = 6) and 7 (n = 6) days after 70% PH and 99mTc-GSA scintigraphy with SPECT was performed to calculate 99mTc-GSA uptake and liver functional volume. After sacrifice, liver weight and conventional liver volume were determined.

Experimental Groups
Two groups of rats in different weight categories, group A; 250–300 g (n = 6) and group B; 300–350 g (n = 6), were used to compare liver volume and liver functional volume measured by 99mTc-GSA SPECT. For the assessment of liver regeneration, 30 rats were randomized into 5 experimental groups. In a control group (n = 6), 99mTc-GSA scintigraphy with SPECT was performed to calculate normal (baseline) 99mTc-GSA uptake and liver functional volume. Subsequently, rats were sacrificed to determine baseline liver weight and liver volume. For the assessment of liver regeneration, rats were anesthetized 1 (n = 6), 3 (n = 6), 5 (n = 6) and 7 (n = 6) days after 70% PH and 99mTc-GSA scintigraphy with SPECT was performed to calculate 99mTc-GSA uptake and liver functional volume. After sacrifice, liver weight and conventional liver volume were determined.

Statistical Analysis
Commercial computer packages were used for the analysis of the data (GraphPad Prism 4; SPSS 12.01 for Windows). Continuous data was compared by independent sample t test and expressed as mean ± SE of the mean (SEM). A correlation between variables was tested using the Pearson correlation coefficient r. The scintigraphy calculations were further analyzed to the method of Bland–Altman, which is a supplementary method to compare 2 different methods when the true value is unknown. The data was plotted as scatter plot of the mean values versus the difference of both calculations. Differences between postoperative 99mTc-GSA uptake, liver functional volume and conventional volume measurements were evaluated with the Wilcoxon signed rank test and a nonparametric repeated measurement ANOVA test. All statistical tests were 2-tailed and differences were considered significant at a P value < 0.05.

RESULTS
Reproducibility of 99mTc-GSA Calculations
The hepatic uptake of 99mTc-GSA was homogenous in all liver lobes (Fig. 2). The uptake rate remained constant after 10 min of imaging. The 99mTc-GSA uptake was measured twice within the same animal, with a time interval of 2 d. No remaining radioactivity was detected in the second measurement before reinjection. There was a strong corre-

Liver Volume During Liver Regeneration
Mean conventional liver volume was decreased 1 d after 70% PH (6.77 ± 0.12 mL) compared with baseline (12.38 ± 0.32 mL), after which it increased from 9.08 ± 0.77 mL at day 3 to normal liver volume at day 5 (12.15 ± 0.45 mL) and day 7 (11.76 ± 0.50 mL, Fig. 5A).

99mTc-GSA SPECT was used to calculate functional liver volume. The mean baseline functional volume was 12.14 ± 0.20 mL. One day after PH, functional volume was reduced to 8.07 ± 0.27 mL. Similar to conventional liver volume, the functional volume increased by day 3 (9.68 ± 0.92 mL) to normal liver functional volume on day 5 (11.82 ± 0.39 mL) and day 7 (11.80 ± 0.37 mL) (Fig. 5A).

There was a significant difference at day 1 between conventional liver volume and liver functional volume assessed by 99mTc-GSA SPECT (6.77 ± 0.12 mL and 8.07 ± 0.27 mL, respectively; P = 0.048, Wilcoxon signed rank test). There were no significant differences between the volumes assessed by both methods at baseline, 3, 5, and 7 d after 70% PH, (P = 0.14, P = 0.71, P = 0.69 and P = 0.92, respectively). The correlation between the 2 volumes in the regenerating liver was strong and significant (Pearson r = 0.924, P < 0.0001; Fig. 5B).
The mean baseline level of $^{99m}$Tc-GSA uptake in a normal liver was $27.08\% \pm 1.82\%/\text{min}$. One day after 70% PH, the $^{99m}$Tc-GSA uptake ($8.01\% \pm 0.62\%/\text{min}$) significantly decreased compared with baseline. At 3, 5, and 7 d after resection, $^{99m}$Tc-GSA uptake increased again to $10.34\% \pm 0.46\%/\text{min}$, $13.53\% \pm 1.20\%/\text{min}$, and $21.71\% \pm 1.94\%/\text{min}$, respectively (Fig. 6).

To investigate if the decrease in uptake of $^{99m}$Tc-GSA was mainly due to the decrease in liver weight after 70% PH, the $^{99m}$Tc-GSA uptake was divided by total liver (wet) weight. The mean baseline level of $^{99m}$Tc-GSA uptake in a normal liver was $2.44\% \pm 0.07\%/\text{min/g}$. One day after 70% PH, the $^{99m}$Tc-GSA uptake per gram liver decreased significantly to $1.51\% \pm 0.08\%/\text{min/g}$ (vs. baseline, $P < 0.008$; Mann–Whitney test) (Table 1). At 3 d after resection, $^{99m}$Tc-GSA uptake per gram liver increased again to $10.34\% \pm 0.46\%/\text{min}$, $13.53\% \pm 1.20\%/\text{min}$, and $21.71\% \pm 1.94\%/\text{min}$, respectively (Fig. 6).

**FIGURE 2.** Dynamic scintigraphy in control rat after intravenous administration of 40 MBq $^{99m}$Tc-GSA in tail vein. (Right panel) Reframed images of dynamic acquisition. There is homogeneous liver uptake from blood-pool activity within 5 min. There is no excretion of tracer within bile to the intestinal tract. (Upper left panel) Summed image with liver in the ROI (white line). (Lower left panel) Time–activity curve of liver ROI, with accumulation to a plateau phase at 10 min after injection.

**FIGURE 3.** (A) Correlation between 2 measurements of $^{99m}$Tc-GSA uptake was significant and strong ($r = 0.754$, $P = 0.019$). (B) The Bland–Altman analysis shows a small bias between the 2 measurements (mean bias, $-1.49\%/\text{min}$), with the 95% limit of agreement between $-10.22\%/\text{min}$ and $7.24\%/\text{min}$. 

**TABLE 1.** Summary of $^{99m}$Tc-GSA uptake in normal and resected liver (mean ± standard error).

<table>
<thead>
<tr>
<th>Time (d)</th>
<th>Baseline</th>
<th>1</th>
<th>3</th>
<th>5</th>
<th>7</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>$27.08% \pm 1.82%/\text{min}$</td>
<td>$2.44% \pm 0.07%/\text{min}$</td>
<td>$10.34% \pm 0.46%/\text{min}$</td>
<td>$13.53% \pm 1.20%/\text{min}$</td>
<td>$21.71% \pm 1.94%/\text{min}$</td>
</tr>
<tr>
<td>Resected</td>
<td>$8.01% \pm 0.62%/\text{min}$</td>
<td>$1.51% \pm 0.08%/\text{min/g}$</td>
<td>$10.34% \pm 0.46%/\text{min}$</td>
<td>$13.53% \pm 1.20%/\text{min}$</td>
<td>$21.71% \pm 1.94%/\text{min}$</td>
</tr>
</tbody>
</table>
and 5 d after 70% PH, it decreased to 1.46% ± 0.13%/min/g and 1.32% ± 0.10%/min/g, respectively (vs. baseline, \( P = 0.008 \) and \( P = 0.004 \), respectively; Mann–Whitney test). At day 7, the \(^{99m}\)Tc-GSA uptake per liver weight had returned to baseline level—that is, 2.18% ± 0.13%/min/g (vs. baseline, \( P = 0.08 \); Mann–Whitney test).

**Figure 4.** (A) Correlation between functional volume assessed by \(^{99m}\)Tc-GSA SPECT and conventional liver volume was strong and significant \(( r = 0.93, P < 0.0001 \)). (B) Correlation between \(^{99m}\)Tc-GSA uptake and conventional liver volume was moderate \(( r = 0.62, P = 0.046 \)).

**Comparison Between \(^{99m}\)Tc-GSA Uptake, Liver Functional Volume, and Liver Volume During Liver Regeneration**

To compare \(^{99m}\)Tc-GSA uptake, liver functional volume, and conventional liver volume during liver regeneration, data were expressed as a percentage of baseline measurements (Fig. 7). During the regeneration process, there were no significant differences between conventional liver volume and liver functional volume assessed by \(^{99m}\)Tc-GSA SPECT \(( P = 0.144 \), nonparametric repeated measurement ANOVA). There were, however, significant differences between \(^{99m}\)Tc-GSA uptake and liver volume \(( P < 0.001 \)) and liver functional volume \(( P < 0.001 \)). When the separate time points were analyzed, significant differences between \(^{99m}\)Tc-GSA uptake and liver volume were found at day 1 \(( P < 0.001 \)), day 3 \(( P = 0.09 \)), and day 5 \(( P < 0.001 \)) after 70% PH. At day 7, there were no significant differences \(( P = 0.14 \)).

**DISCUSSION**

The liver has the unique ability to regenerate after a partial liver resection. In the clinical setting, liver regeneration is traditionally evaluated using CT volumetry. However, during liver regeneration, hepatic volume does not always represent hepatocellular function \((5–7)\). Consequently, additional functional assessment of the regenerating liver should be considered.

As liver regeneration is difficult to study in humans, a standardized rat model of 70% PH is commonly used. Similar to the clinical situation, liver regeneration is generally assessed by increase of liver volume. Only a few studies have investigated the recovery of liver function after PH. In this study we evaluated the utility of \(^{99m}\)Tc-GSA scintigraphy combined with SPECT for the assessment of liver function and liver functional volume in normal and regenerating rat liver.

\(^{99m}\)Tc-GSA liver scintigraphy was developed in Japan as a noninvasive liver function test. The liver is the only uptake site for \(^{99m}\)Tc-GSA and, therefore, it is an ideal agent for functional liver scintigraphy. It is assumed that the ligand–receptor binding is a second-order process, in which a significant fraction of receptor is occupied during the course of the imaging study \((17)\). This enables SPECT acquisitions in small laboratory animals for the assessment of both liver function and liver functional volume.

Clinical studies report a correlation with conventional liver function tests, including antithrombin III, total and direct bilirubin, prothrombin time, indocyanine green clearance, Child–Pugh classification, and histology \((hepatic activity index score)\) \((19,24)\). However, the broader use of \(^{99m}\)Tc-GSA liver scintigraphy has been hampered by the fact that it has not been approved for clinical use in Europe.
and the United States. $^{99m}$Tc-GSA is available for preclinical investigation outside of Japan. In the clinical situation, liver function measured by $^{99m}$Tc-GSA scintigraphy, is expressed by many different, sometimes complex, parameters (12,19,25–28). In our study, we applied a method that calculates the hepatic binding rate on the basis of liver $^{99m}$Tc-GSA uptake and the count ratio of liver to blood pool. $^{99m}$Tc-GSA SPECT was used for the calculation of liver functional volume, using the outline extraction method (29).

In the normal rat, as well as during the regeneration process of the liver, we demonstrated a strong correlation between conventional liver volume and liver functional volume. Therefore, $^{99m}$Tc-GSA SPECT is an accurate, non-invasive method to assess liver functional volume in normal rat liver.

After 70% PH, a discrepancy between volumetric ($^{99m}$Tc-GSA SPECT, conventional analysis) and functional ($^{99m}$Tc-GSA uptake) recovery was observed during the regeneration process. $^{99m}$Tc-GSA uptake underestimated hepatic regeneration in comparison with liver volume. In both clinical and experimental studies, a discrepancy has been described between volumetric regeneration and functional regeneration after PH. Tanaka et al. reported that functional regeneration was impaired in large resections in comparison with volumetric regeneration (28). This can be explained by the theory that, during liver regeneration, hepatocyte proliferation is promoted at the expense of liver function (30). Others, however, concluded that the functional regeneration was more rapid than the morphologic regeneration (6,7). These results support the theory that hepatocyte function is upregulated after major hepatic resection to compensate for the reduced liver mass (22,31,32). Therefore, it remains difficult to draw conclusions with regard to the difference between functional and volumetric regeneration.

In our study, $^{99m}$Tc-GSA uptake was decreased in the first days after partial liver resection, which was not only due to the decrease in liver weight after resection. The $^{99m}$Tc-GSA uptake per gram liver was significantly decreased during the first days after resection, which returned to baseline at day 7. Although the uptake of $^{99m}$Tc-GSA is influenced by blood flow, liver perfusion is increased after partial liver resection due to the fact that the complete portal flow is directed through the small remnant liver (33). When the remnant liver increases, portal perfusion returns to baseline levels. It has been suggested that increased liver perfusion after resection is one of the main triggers for regeneration (33). Therefore, it is unlikely that the decrease in $^{99m}$Tc-GSA uptake, as found in this study, results from a change in liver perfusion because this is increased during the first days after liver resection.

The decrease in uptake per gram liver can be explained by a decrease in ASGP receptors per hepatocyte, a decrease in affinity of the ASGP receptor, or a decrease in the amount of hepatocytes per gram liver. In vitro studies have described a decrease in surface ASGP receptors during liver regeneration.

![Figure 6](image1.png) **FIGURE 6.** $^{99m}$Tc-GSA uptake decreased the first day after 70% PH, after which it slowly recovered to baseline uptake at day 7 after resection.

![Figure 7](image2.png) **FIGURE 7.** During the regeneration process, there were significant differences between $^{99m}$Tc-GSA uptake and both conventional liver volume ($P < 0.001$) and liver functional volume ($P < 0.001$), when data were expressed as a percentage of baseline levels.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Baseline</th>
<th>Day 1</th>
<th>Day 3</th>
<th>Day 5</th>
<th>Day 7</th>
</tr>
</thead>
<tbody>
<tr>
<td>$^{99m}$Tc-GSA (%/min/g)</td>
<td>2.44 ± 0.17</td>
<td>1.51 ± 0.08*</td>
<td>1.46 ± 0.13*</td>
<td>1.32 ± 0.10*</td>
<td>2.18 ± 0.13</td>
</tr>
</tbody>
</table>

*Versus baseline, $P < 0.05$; Mann–Whitney test.

**TABLE 1**

$^{99m}$Tc-GSA Uptake per Liver Weight in Regenerating Liver After 70% PH
regeneration (34,35). Mechanisms of decreased cell-surface ASGP receptors include the secretion of the ASGP receptor into the extracellular space and reduced recycling of the receptor after endocytosis (34). Our results may support the theory that the amount of ASGP receptors is decreased, and it is uncertain whether dynamic 99mTc-GSA scintigraphy still accurately represents liver function during the early phase of regeneration. Therefore, further validation and investigation of possible mechanisms of the decrease of ASGP receptor is advised before it can be used for the assessment of liver function in a regenerating rat liver.

CONCLUSION

99mTc-GSA scintigraphy combined with SPECT is a feasible, noninvasive method to assess hepatic volume in normal rat liver as well as in the regenerating rat liver. However, the hepatic 99mTc-GSA uptake seems to underestimate hepatic regeneration, possibly due to a decrease of ASGP receptors in the early phase after PH.

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