

Distribution Volume of ^{18}F -BMS-986192 in NSCLC Patients

TO THE EDITOR: In an interesting article, Dr. Huisman et al. recently investigated the optimal kinetic model for an ^{18}F -labeled anti-programmed cell death ligand 1 (anti-PD-L1) adnectin, namely, ^{18}F -BMS-986192, to quantify PD-L1 expression in non-small-cell lung cancer (NSCLC) patients (1). A single-tissue-reversible (STR) compartment model, additionally including blood-volume fraction, was found to be the most preferred model for fitting the tumor time-activity curves. Its specific outcome measure is the distribution volume (V_T ; $\text{mL}\cdot\text{cm}^{-3}$) that is the equilibrium ratio of forward/reverse transport-rate constants, that is, K_i/k_b , between blood and reversible-trapping compartment (2). V_T was then used to validate simplified methods, the best correlation being obtained with body weight-normalized SUV (SUV_{BW}) at 50–80 min after injection ($R^2 = 0.92\text{--}0.91$), whereas a lower correlation was obtained with SUV normalized to plasma concentration ($\text{SUV}/C_{\text{plasma}}$, presumably at 50 min after injection; $R^2 = 0.84$). The authors conclude that SUV_{BW} at 60 min after injection is an accurate simplified parameter for uptake assessment of ^{18}F -BMS-986192 baseline studies.

We would like to further analyze the latter lower correlation since, under postinjection time conditions we address in this letter: (i) V_T may be assessed by the ratio of tissue/plasma tracer concentration, i.e., $C_{\text{tissue}}/C_{\text{plasma}}$; and (ii) the $\text{SUV}_{\text{BW}}/C_{\text{plasma}}$ ratio may be also proportional to V_T since SUV_{BW} is proportional to C_{tissue} . To clarify this issue, we have fitted the ^{18}F -BMS-986192 input function with a triexponentially decaying function and then fitted a PD-L1-positive tumor time-activity curve by using a 3-compartment 3-parameter kinetic model (data extracted from Figs. 3 and 4 in Huisman with the Web-Plot-Digitizer software; $R^2 = 0.996$ and 0.998 , respectively) (1,3). Estimates of K_i and k_b were provided, leading to the computation of V_T as $K_i/k_b = 4.7 \text{ mL}\cdot\text{cm}^{-3}$. This analysis also allowed us to perform both decay-corrected tissue- and decay-uncorrected trapped-tracer time-activity curves (supplemental data, available at <http://jnm.snmjournals.org>).

Let us first consider the rate of decay-corrected trapped tracer per tissue volume unit (at steady state): $dC_{\text{trapped}}(t)/dt = K_i \times C_{\text{plasma}}(t) - k_b \times C_{\text{trapped}}(t)$. At peak time of decay-corrected C_{trapped} time-activity curve, $dC_{\text{trapped}}(t)/dt = 0$ and then $C_{\text{trapped}}(t_{\text{peak}})/C_{\text{plasma}}(t_{\text{peak}}) = K_i/k_b = V_T$. Assuming $C_{\text{tissue}}(t_{\text{peak}}) \approx C_{\text{trapped}}(t_{\text{peak}})$ (i.e., neglecting free tracer in blood and interstitial volume), t_{peak} was estimated to be 87 min from decay-corrected C_{tissue} time-activity curve, leading to $C_{\text{tissue}}/C_{\text{plasma}} = 4.5 \text{ mL}\cdot\text{cm}^{-3}$ (versus $K_i/k_b = 4.7 \text{ mL}\cdot\text{cm}^{-3}$). Second, considering decay-uncorrected data, the differential equation becomes $dC_{\text{trapped}}(t)/dt = K_i \times C_{\text{plasma}}(t) - k_b \times C_{\text{trapped}}(t) - \lambda \times C_{\text{trapped}}(t)$, where λ is the ^{18}F physical-decay-rate constant. As a consequence, at peak time of decay-uncorrected- C_{trapped} time-activity curve, $C_{\text{trapped}}(t_{\text{peak}})/C_{\text{plasma}}(t_{\text{peak}}) = K_i/(k_b + \lambda)$. The ratio $K_i/(k_b + \lambda)$ was calculated as $2.1 \text{ mL}\cdot\text{cm}^{-3}$,

whereas, at decay-uncorrected- C_{trapped} t_{peak} of 53 min after injection, the ratio $C_{\text{tissue}}/C_{\text{plasma}}$ (that may involve decay correction or not) was found to be $2.2 \text{ mL}\cdot\text{cm}^{-3}$.

We therefore suggest that the $\text{SUV}/C_{\text{plasma}}$ ratio (or, equivalently, the $C_{\text{tissue}}/C_{\text{plasma}}$ ratio) is actually correlated with $V_T = K_i/k_b$ when assessed within 85–90 min after injection. However, the authors acknowledged that their results were only valid within 50–80 min after injection (1). Furthermore, we suggest that the $\text{SUV}/C_{\text{plasma}}$ ratio assessed within 50–55 min after injection should be correlated with $K_i/(k_b + \lambda)$, instead of K_i/k_b (1). This alternative ratio reports on what is actually occurring at decay-uncorrected- C_{trapped} t_{peak} , that is, an equilibrium between uptake and release plus physical decay. It is worth noting, regarding the part of postinjection time in its measurement uncertainty, a +13% increase occurs in the 50–55 min time range, whereas, for comparison, C_{tissue} alone, and, hence, SUV_{BW} , shows a +3% increase.

In conclusion, investigating potential clinical biomarkers and relevant simplified metrics is of utmost importance for selecting NSCLC patients who could benefit from immune checkpoint-inhibitor treatment. In ^{18}F -BMS-986192 PET imaging, Huisman et al. convincingly showed that SUV_{BW} at 60 min after injection, may be a relevant simplified parameter to quantify tumor uptake for baseline PET studies. We additionally suggest that the ratio $\text{SUV}_{\text{BW}}/C_{\text{plasma}}$ might be probed as a complementary possible simplified parameter, that is correlated with $K_i/(k_b + \lambda)$ within 50–55 min after injection.

DISCLOSURE

No potential conflict of interest relevant to this article was reported.

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