

Labeled Leukocyte Imaging: Dawn of an Era

Christopher J. Palestro

Donald and Barbara Zucker School of Medicine, Hofstra University and Northwell Health, Hempstead, New York

More than one hundred years after the discovery of penicillin, infection remains a major threat to humankind. In 2017, more than 8 million deaths and 400,000 years of life lost were due to infection, making it first in morbidity and third in mortality among human diseases (1). Diagnosing infection is challenging, and imaging studies are often used for confirmation, localization, and assessment of its extension. For nearly 50 years, beginning with ^{67}Ga , molecular imaging has played an important role in the diagnosis of infection. Suboptimal imaging characteristics, the typical 48- to 72-h delay between administration and imaging, and the inability of ^{67}Ga to differentiate infection from inflammation, tumor, and trauma inspired the search for better agents (2).

In the mid 1970s, the demonstration by Thakur et al. (3) that autologous leukocytes labeled in vitro with ^{111}In -oxine could image infection in humans was a seminal event. The concept on which this test is based is simple: imaging of a physiologic process, the in vivo migration of radiolabeled leukocytes. Although the concept is simple, the execution is not and depends on several factors, including the imaging characteristics of the radionuclide, the label stability, and the labeled cell viability. Tritium, ^{32}P , and ^{51}Cr , which had been used to label leukocytes, provide data on leukocyte clearance from the blood but are not suitable for studying in vivo kinetics and distribution of leukocytes and cannot be used for localizing infection. ^{111}In has γ -emissions 174 keV and 247 keV, which are suitable for imaging. Its 67-h half-life is sufficiently long to image leukocyte migration and accumulation (3).

The label must be stable, and the labeled cells must be viable. Significant radionuclide elution from the cells would confound the interpretation of the test because it would not be possible to differentiate accumulation of labeled leukocytes from accumulation of other radiolabeled complexes or free radionuclide. Once inside the leukocyte, ^{111}In binds to intracellular components

Received May 15, 2020; revision accepted Jun. 4, 2020.

For correspondence or reprints contact: Christopher J. Palestro, Division of Nuclear Medicine and Molecular Imaging, Long Island Jewish Medical Center, 270-05 76th Ave., New Hyde Park, NY 11040.

E-mail: palestro@northwell.edu

COPYRIGHT © 2020 by the Society of Nuclear Medicine and Molecular Imaging.

DOI: 10.2967/jnumed.120.249797

Indium-111-Labeled Autologous Leukocytes in Man

Mathew L. Thakur, J. Peter Lavender, Rosemary N. Arnot, David J. Silvester,
and Anthony W. Segal

Hammersmith Hospital, London, England

Autologous leukocytes have been isolated, labeled with indium-111, and administered to 15 patients suspected of inflammatory disease. The stability of the label has been demonstrated and the in vivo kinetics and distribution of the labeled cells studied. The distribution is influenced by the type and viability of the cells separated by three different techniques. Generally, there was initial accumulation of radioactivity in the lungs; approximately half of this cleared in 15 min and the remainder slowly. Twenty-five to 50 percent of the radioactivity subsequently distributed in the spleen, liver, and bone marrow, and these did not show significant change with time up to 48 hr post injection. The In-111 radioactivity administered as labeled leukocytes free from erythrocytes cleared from the circulating blood with a half-time of 7.5 hr.

In three of 15 patients, the suspicion of inflammatory disease could not be confirmed, and in these a normal distribution of radioactivity was observed. In the remaining 12 patients, focal accumulation of radioactivity was detectable within 4 to 24 hr after administration, and subsequent confirmation of sepsis was obtained. From three such patients, samples of abscesses were recovered which showed markedly higher radioactivity than that in the same weight of blood.

J Nucl Med 18: 1012-1019, 1977

The behavior of neutrophils within the body and their localization in pathologic processes has been a subject of study since the late 19th century (1,2). Over the past two decades, the use of radioactive tracers has provided an excellent means for measuring the rate of disappearance of labeled cells from the circulation. Until recently, however, (3,4) the radioactive tracers for white cells have been limited to tritium (H-3 , $T_{1/2} = 12.4$ yr), phosphorus-32 (P-32 , $T_{1/2} = 14.3$ d) and chromium-51 (320 keV 7%, $T_{1/2} = 28$ d). Owing to the unsuitability for external

cles, approximately 40% of the activity is intracellular but the remainder represents particles that are absorbed non-specifically on the cell membrane and cannot be eliminated easily (7).

Indium-111 chelated with 8-hydroxyquinoline (oxine) has been found to be the most efficient agent of several radioactive particles (8) and soluble agents (9) that have been investigated for the labeling of leukocytes. Indium-111 has favorable gamma emissions for external detection (84% at 173 keV, and 94% at 240 keV), and its half-life of 67 hr is

358

Number of Citations



with minimal elution; up to 90% of the activity is retained intracellularly at 22 h. Finally, the labeled leukocytes must be viable; leukocyte viability was about 75% (3). To put the work of Thakur et al. (3) into proper perspective, in the decades since its publication, among the several radionuclides (including positron emitters) investigated for labeling leukocytes, only ^{99m}Tc has yielded a clinically useful agent (2).

Compared with the ideal molecular infection imaging agent, which should be safe, available, rapidly completed, and sensitive and specific for infection, labeled leukocyte imaging has several weaknesses. The in vitro labeling process requires handling of human blood products, with its associated hazards for personnel and patients. The procedure is labor-intensive and time-consuming and can be performed only by trained individuals. Consequently, in most institutions, the test is available only during routine working hours.

The issue of sensitivity and specificity is not straightforward. Although labeled leukocyte imaging is used for infection imaging, it is in reality host-response imaging, in which the presence of infection is implied by patterns of labeled leukocyte accumulation. In the usual clinical scenario, most leukocytes labeled are neutrophils; the test is therefore most sensitive for detecting those infections in which the primary cellular response is neutrophilic, such as bacterial infections. The procedure is less sensitive for detecting those infections in which the predominant cellular response is not neutrophilic, that is, tuberculosis and some opportunistic infections (2).

Labeled leukocyte imaging is specific for leukocyte-mediated inflammatory processes but is not specific for infection and will be positive in any inflammatory process mediated by neutrophils. For example, the test cannot reliably differentiate septic arthritis from an active inflammatory process such as rheumatoid arthritis (2).

Why then does labeled leukocyte imaging—a procedure that is not without some risk, is not ubiquitously available, and is not specific for infection—still have a preeminent position in molecular imaging of infection? The answer is that, to date, no

better agent has come along. Numerous attempts have been made to develop in vivo leukocyte labeling methods using antigranulocyte antibodies, antibody fragments, and peptides, none of which have stood the test of time. Generally they are sensitive, but specificity is variable and they have their own safety issues, including human antimurine antibody development precluding repeat use, and severe adverse events occurring shortly after administration. Attempts to develop infection-specific agents such as radiolabeled antibiotics, vitamins, and antimicrobial peptides have had only modest success, and none have ever made it into routine nuclear medicine practice (2).

The one agent that has gained widespread acceptance for imaging of infection is ^{18}F -FDG. Though not specific, the test is rapidly completed and provides high-resolution images. Furthermore, ^{18}F -FDG is especially useful for those indications for which labeled leukocyte imaging is of limited value, such as tuberculosis, spondylodiscitis, and fever of unknown origin. ^{18}F -FDG, however, is a complement to, not a replacement for, labeled leukocyte imaging (2).

More than 4 decades after its introduction, in vitro labeled leukocyte imaging still occupies a preeminent role in molecular imaging of infection. Considering the rapidity with which modern-day diagnostic imaging tests are developed, this is a remarkable feat.

DISCLOSURE

No potential conflict of interest relevant to this article was reported.

REFERENCES

1. Ordonez AA, Sellmyer MA, Gowrishankar G, et al. Molecular imaging of bacterial infections: overcoming the barriers to clinical translation. *Sci Transl Med*. 2019;11:eaax8251.
2. Palestro CJ. Molecular imaging of infection: the first 50 years. *Semin Nucl Med*. 2020;50:23–34.
3. Thakur ML, Lavender JP, Arnot RN, Silvester DJ, Segal AW. Indium-111-labeled autologous leukocytes in man. *J Nucl Med*. 1977;18:1014–1021.

4. Dancy JT, Deubelbeiss KA, Harkar LA, et al: Neutrophil kinetics in man. *J Clin Inv* 58: 705-715, 1976
5. Burleson RL, Holman BL, Tow DE: Scintigraphic demonstrations of abscesses with radioactive labeled leukocytes. *Surg Gynecol Obstet* 14: 379-382, 1975
6. Anderson BR, English D, Akalin HE, et al: Inflammatory lesions localised with ^{99m}Tc labeled leukocytes. *Arch Intern Med* 135: 1067-1071, 1975
7. Thakur ML, Coleman RE, Welch MJ: Indium-111 labeled leukocytes for the localization of abscesses: Preparation, analysis, tissue distribution and comparison with gallium-67 citrate in dogs. *J Lab Clin Med* 89: 217-228, 1977
8. McAfee JG, Thakur ML: Survey of radioactive agents for in vitro labeling of phagocytic leukocytes. I. Soluble agents. *J Nucl Med* 17: 480-487, 1976
9. McAfee JG, Thakur ML: Survey of radioactive agents for in vitro labeling of phagocytic leukocytes. II. Particulate agents. *J Nucl Med* 17: 488-493, 1976
10. Thakur ML, Segal AW, Welch MJ, et al: Indium-111 labeled cellular blood compounds: Mechanism of labeling and intracellular localization in human neutrophils. *J Nucl Med*: in press
11. Thakur ML, Coleman RE, Mayhall GC, et al: Preparation and evaluations of ¹¹¹In-labeled leukocytes as an abscess imaging agent in dogs. *Radiology* 119: 731-732, 1976
12. Segal AW, Thakur ML, Arnot RN, et al: Indium-111 labeled leukocytes for localization of abscesses. *Lancet* 13: 1056-1058, 1976
13. Bøyum A: Separation of leukocytes from blood and bone masses. *Scan J Clin Lab Invest*: Suppl No 97, 77-89, 1968
14. Park BH, Fikrig SM, Smithwick EM: Infections and nitroblue tetrazolium reduction by neutrophils. A diagnostic acid. *Lancet* 2: 532-534, 1968
15. Williams ED, Merrick MV, Lavender JP: The distribution and dosimetry of ¹¹¹In bleomycin in man. *Brit J Radiol* 48: 275-278, 1975
16. Snyder WS, Ford MR, Warner GG, et al: Estimates of absorbed fractions for monoenergetic photon sources uniformly distributed in various organs of a heterogeneous phantom. MIRD Pamphlet No 5, *J Nucl Med* 10: Suppl No 3, 1969
17. Weisberger AS, Heinle RW, Storaasli JP, et al: Transfusion of leukocytes labeled with radioactive phosphorous. *J Clin Invest* 29: 336-341, 1950
18. English D, Anderson BR: *J Immunol Methods* 5: 249-252, 1974
19. Herzenberg LA, Sweet RG, Herzenberg LA: Fluorescence-activated cell sorting. *Sci Am* 234: 108-117, 1976
20. Thakur ML, Welch MJ, Joist JH, et al: Indium-111 labeled platelets: Studies on preparation and evaluations of in vitro and in vivo functions. *Thromb Res* 9: 345-357, 1976
21. Rannie GH, Thakur ML, Ford WL: An experimental comparison of radioactive labels with potential application to lymphocyte migration studies in patients. *Clin Exp Immunol*: in press