Somatostatin Receptor Scintigraphy: Blazing in Indium and Quenching in Gallium

Lisa Bodei

Molecular Imaging and Therapy Service, Department of Radiology, Memorial Sloan Kettering Cancer Center, New York, New York

In contemplating peer-reviewed information and the business of science divulgation, in which we all are immersed, one may wonder what legacy a very quoted article will hand down to posterity and what core of knowledge will stand the tests of time, of fashion, and of planned obsolescence of intellectual production (1).

Somatostatin receptor theranostics, as we know them today, need to be inscribed in the history of neuroendocrine tumors (NETs)—a relatively rare disease with high prevalence—which has been constellated with numerous fundamental discoveries, many of which awarded the Nobel Prize. Specifically, NET imaging

Received Jun. 18, 2020; revision accepted Jun. 26, 2020. For correspondence or reprints contact: Lisa Bodei, Molecular Imaging and

Therapy Service, Department of Radiology, Memorial Sloan Kettering Cancer Center, 1275 York Ave., New York, NY 10065.

E-mail: bodeil@mskcc.org

COPYRIGHT © 2020 by the Society of Nuclear Medicine and Molecular Imaging. DOI: 10.2967/inumed.120.251934

stems from the isolation of somatostatin from ovine hypothalamus by Paul Brazeau and Roger Guillemin in 1972 (later synthesized by Andrew V. Shally) and from the synthesis of octreotide, an analog capable of stability in plasma, in 1979. It then continues with the demonstration of overexpression of binding sites with high affinity for somatostatin in several NETs and culminates with the identification and cloning of the 5 somatostatin receptors in 1992 (2). In the same period, the group led by Eric P. Krenning at Erasmus University, Rotterdam, which included extraordinary teamwork by nuclear medicine and internal medicine physicians, endocrinologists, biologists, and radiochemists, with the collaboration of Jean-Claude Reubi, succeeded in radiolabeling octreotide. The idea at the basis of this endeavor was to exploit the internalizing and retention potential of the somatostatin analog octreotide to obtain in vivo binding and visualization of NETs. The obvious choice of radiolabel fell on the default imaging radionuclide at the time, ¹²³I, which could be directly labeled to the Tyr3-substituted octreotide through the chloramine method. The

Somatostatin Receptor Scintigraphy with Indium-111-DTPA-D-Phe-1-Octreotide in Man: Metabolism, Dosimetry and Comparison with Iodine-123-Tyr-3-Octreotide

E.P. Krenning, W.H. Bakker, P.P.M. Kooij, W.A.P. Breeman, H.Y. Oei, M. de Jong, J.C. Reubi, T.J. Visser, C. Bruns, D.J. Kwekkeboom, A.E.M. Reijs, P.M. van Hagen, J.W. Koper, and S.W.J. Lamberts

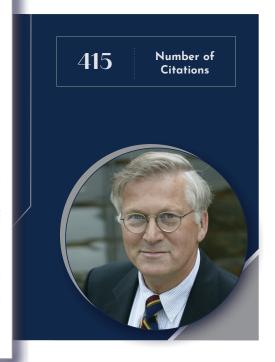
Departments of Nuclear Medicine and Internal Medicine III, University Hospital Dijkzigt, Rotterdam, The Netherlands; Sandoz Research Institute, Berne and Department of Endocrinology, Sandoz Pharma AG, Basel, Switzerland

Scintigraphy with 123 l-Tyr-3-octreotide has several major drawbacks as regards its metabolic behavior, its cumbersome preparation and the short physical half-life of the radionuclide.

The use of another radiolabeled analog of somatostatin, 111 In-DTPA-D-Phe-1-octreotide, has consequently been proposed. DTPA-D-Phe-1-octreotide can be radiolabeled with DTPA-D-Pne-1-octreotide can be radiotabeled with "In in an aneasy single-step procedure. DTPA-D-he-1-octreotide is cleared predominantly via the kidneys. Fecal excretion of radioactivity amounts to only a few percent of the administered radioactivity. For the radiation dose to normal tissues, the most important organs are the kidneys, the spleen, the urinary bladder, the liver and the remainder of the body. The calculated effective dose equivalent is 0.08 mSv/MBq. Optimal 111n-DTPA-D-Phe-1-octreotide scintigraphic imaging of various somatostatin receptor-positive tumors was obtained 24 hr after injection. In the six patients studied, tumor localization with ¹²³I-Tyr-3-octreotide and with ¹¹¹In-DTPA-D-Phe-1-octreotide were found to be similar. However, the norma pituitary is more frequently visualized with the latter radio-pharmaceutical. In conclusion, 111In-DTPA-D-Phe-1-octreotide appears to be a sensitive somatostatin receptor-positive tis sue-seeking radiopharmaceutical with some remarkable ad-vantages: easy preparation, general availability, appropriate half-life and absence of major interference in the upper ab dominal region, because of its renal clearance. Therefore, dominal region, because of its form the suitable for use in

carcinoids, islet cell tumors of the pancreas, paragangliomas and small-cell carcinomas of the lungs (1-4). Our experience points to several drawbacks of ¹²³I-Tyr-3-octreotide in its use for in vivo scintigraphy. First, the labeling of Tyr-3-octreotide with ¹²³I is cumbersome and requires special skills. Second, Na¹²³I of high specific activity (5,6) is expensive and hardly available worldwide. Third, the moment of the labeling and scanning procedures is dependent on the logistics of the production and delivery of Na¹²³I. Finally, substantial accumulation of radioactivity is seen in the intestines, since a major part of ¹²³I-Tyr-3-octreotide is rapidly cleared via the liver and biliary system. This makes the interpretation of planar and single-photon emission computed tomographic (SPECT) images of the upper abdomen difficult.

Part of these problems can be solved by replacing ¹²³I with ¹¹¹In, which also improves scintigraphy 24-48 h ra after application by virtue of its longer half-life. Binding of ¹¹¹In to the somatostatin analog octreotide has been carried out by complexing with a diethylenetriaminepentaacetic acid (DTPA) group coupled to the αNH₂-group of the N-terminal D-Phe residue (7). In rats, it appeared that ¹¹¹In-DTPA-D-Phe-I-octreotide: (1) is excreted via the kidneys, (2) shows only minor accumulation in the liver and (3) has an initial plasma half-life in the order



Bodei

first 10 patients studied with 123I-Tyr3-octreotide were announced in 1989 and constituted the first proof of principle of the value of somatostatin receptor imaging (3).

The disadvantages of scintigraphic imaging with the iodinated octreotide derivative were, however, immediately apparent: the difficult radiolabeling procedure, the cost and scarce availability of ¹²³I worldwide, and, most importantly, the predominant biliary excretion, with substantial accumulation of the radiopeptide in the intestines, which complicated the interpretation of the very abdominal region involved by most NETs.

The group then developed an improved radiolabeled octreotide by replacing 123 I with 111 In. The different chemistry of this radionuclide required a chelating group, diethylenetriaminepentaacetic acid (DTPA), to be coupled to the amino-terminal of the first phenylalanine to form 111 In-DTPA-D-Phe1-octreotide, which would later be named 111In-pentetreotide, or OctreoScan (Mallinckrodt). The new compound exhibited a more hydrophilic profile, with predominant renal excretion and only minimal biliary excretion. The longer half-life of ¹¹¹In allowed for prolonged imaging times. The results for the first 26 patients studied by γ-camera scintigraphy, with a subgroup undergoing pharmacokinetics and dosimetry, were published in the seminal article "Somatostatin Receptor Scintigraphy with Indium-111-DTPA-D-Phe-1-Octreotide in Man: Metabolism, Dosimetry and Comparison with Iodine-123-Tyr-3-Octreotide" in 1992 (4). In these patients, 111In-DTPA-D-Phe1-octreotide provided optimal tumor visualization 24 h after injection, with comparable biodistribution (except for more intense uptake in the pituitary) and the capability for tumor localization to the original ¹²³I-Tyr³-octreotide but improved visualization of the abdomen and target-to-background ratios due to the longer half-life of ¹¹¹In. In addition, ¹¹¹In-DTPA-D-Phe¹-octreotide exhibited favorable pharmacokinetics (rapid plasma clearance and 85% urinary excretion at 24 h) and dosimetry (>70% of the effective-dose equivalent resulting from radioactivity in the kidneys, spleen, and liver). Multiphasic (4, 24, and 48 h) and SPECT imaging clearly improved the localization of lesions.

This 1992 work represented a milestone in NET management, was a paragon for the future development of theranostics based on radiolabeled receptor ligands, and marked the beginning of the era of somatostatin receptor imaging on a large scale. 111In-DTPA-D-Phe¹-octreotide was, in fact, the first imaging radiopharmaceutical to receive Food and Drug Administration approval, in 1994. It was extensively used worldwide for the following 20 years, allowing for the localization, staging, and restaging of NETs and their metastases, as well as selection for treatment with somatostatin

111In-DTPA-D-Phe1-octreotide had formidable success and opened the way to the radionuclide treatment of advanced NETs, which the author group developed as the next logical step of the in vivo localization by exploiting the Auger emission of 111In administered in high activities (5). This approach initiated one of the most fortunate examples of theranostics, and a few decades and a few radionuclides later, it led the same group of authors to develop yet another radiopharmaceutical, ¹⁷⁷Lu-DOTATATE, now approved for the treatment of metastatic or inoperable gastroenteropancreatic NETs (6).

However, just like our "Juggler of Day," 111 In-DTPA-D-Phe1octreotide also had a decline. Despite the improvement resulting from SPECT/CT hybrid imaging, the diffusion of PET/CT on a large scale rendered this tracer, which had been the workhorse of NET imaging for over 2 decades, gradually obsolete. Its resolution limit and the frequent suboptimal visualization of the abdomen, due to intestinal activity, which required biphasic imaging, laxatives, and hydration, could not compare with the progressively more sophisticated CT and MR images. The availability of ⁶⁸Ga-labeled compounds then marked the official decline of somatostatin receptor scintigraphy and the beginning of a new era. But this is a story for the next few decades...

DISCLOSURE

No potential conflict of interest relevant to this article was reported.

REFERENCES

- 1. Jacoby R. Social Amnesia: A Critique of Conformist Psychology from Adler to Laing. Boston, MA: Beacon Press; 1975.
- 2. Lamberts SW, van der Lely AJ, de Herder WW, Hofland LJ. Octreotide. N Engl J Med. 1996:334:246-254.
- 3. Krenning EP, Bakker WH, Breeman WA, et al. Localisation of endocrine-related tumours with radioiodinated analogue of somatostatin, Lancet, 1989:1:242-244.
- 4. Krenning EP, Bakker WH, Kooij PPM, et al. Somatostatin receptor scintigraphy with indium-111-DTPA-D-Phe-1-octreotide in man: metabolism, dosimetry and comparison with iodine-123-Tyr-3-octreotide. J Nucl Med. 1992;33:652-658.
- 5. Krenning EP, Kooij PP, Bakker WH, et al. Radiotherapy with a radiolabeled somatostatin analogue, [111In-DTPA-D-Phe1]-octreotide: a case history. Ann N Y Acad Sci. 1994;733:496-506.
- 6. Kwekkeboom DJ, de Herder WW, Kam BL, et al. Treatment with the radiolabeled somatostatin analog [177 Lu-DOTA 0 ,Tyr 3]octreotate: toxicity, efficacy, and survival. JClin Oncol. 2008;26:2124-2130.

Somatostatin Receptor Scintigraphy with Indium-111-DTPA-D-Phe-l-Octreotide in Man: Metabolism, Dosimetry and Comparison with Iodine-123-Tyr-3-Octreotide

E.P. Krenning, W.H. Bakker, P.P.M. Kooij, W.A.P. Breeman, H.Y. Oei, M. de Jong, J.C. Reubi, T.J. Visser, C. Bruns, D.J. Kwekkeboom, A.E.M. Reijs, P.M. van Hagen, J.W. Koper, and S.W.J. Lamberts

Departments of Nuclear Medicine and Internal Medicine III, University Hospital Dijkzigt, Rotterdam, The Netherlands; Sandoz Research Institute, Berne and Department of Endocrinology, Sandoz Pharma AG, Basel, Switzerland

Scintigraphy with 123I-Tyr-3-octreotide has several major drawbacks as regards its metabolic behavior, its cumbersome preparation and the short physical half-life of the radionuclide. The use of another radiolabeled analog of somatostatin, 111In-DTPA-D-Phe-1octreotide, has consequently been proposed. DTPA-D-Phe-1-octreotide can be radiolabeled with 111In in an easy single-step procedure. DTPA-D-Phe-1-octreotide is cleared predominantly via the kidneys. Fecal excretion of radioactivity amounts to only a few percent of the administered radioactivity. For the radiation dose to normal tissues, the most important organs are the kidneys, the spleen, the urinary bladder, the liver and the remainder of the body. The calculated effective dose equivalent is 0.08 mSv/MBq. Optimal 111In-DTPA-D-Phe-1-octreotide scintigraphic imaging of various somatostatin receptor-positive tumors was obtained 24 hr after injection. In the six patients studied, tumor localization with 123|-Tyr-3-octreotide and with 111In-DTPA-D-Phe-1-octreotide were found to be similar. However, the normal pituitary is more frequently visualized with the latter radiopharmaceutical. In conclusion. 111 In-DTPA-D-Phe-1-octreotide appears to be a sensitive somatostatin receptor-positive tissueseeking radiopharmaceutical with some remarkable advantages: easy preparation, general availability, appropriate half-life and absence of major interference in the upper abdominal region, because of its renal clearance. Therefore, 111In-DTPA-D-Phe-1-octreotide may be suitable for use in SPECT of the abdomen, which is important in the localization of small endocrine gastroenteropancreatic tumors.

J Nucl Med 1992; 33:652–658DOI: 10.2967/jnumed.120.251934a

e recently introduced the somatostatin analog Tyr-3-octreotide labeled with ¹²³I for the localization of primary and metastatic somatostatin receptor-rich tumors, such as carcinoids, islet cell tumors of the pancreas, paragangliomas and small-cell carcinomas of the lungs (1,4). Our experience points to several drawbacks of ¹²³I-Tyr-3-octreotide in its use for in vivo scintigraphy. First, the labeling of Tyr-3-octreotide with ¹²³I is cumbersome and requires special skills. Second, Na¹²³I of high specific activity (5,6) is expensive and hardly available worldwide. Third, the

Received Jul. 23, 1991; revision accepted Dec. 4, 1991.

moment of the labeling and scanning procedures is dependent on the logistics of the production and delivery of Na¹²³I. Finally, substantial accumulation of radioactivity is seen in the intestines, since a major part of ¹²³I-Tyr-3-octreotide is rapidly cleared via the liver and biliary system. This makes the interpretation of planar and single-photon emission computed tomographic (SPECT) images of the upper abdomen difficult.

Part of these problems can be solved by replacing 123 I with 111 In, which also improves scintigraphy 24–48 hr after application by virtue of its longer half-life. Binding of 111 In to the somatostatin analog octreotide has been carried out by complexing with a diethylenetriaminepentaacetic acid (DTPA) group coupled to the α NH₂-group of the N-terminal D-Phe residue (7). In rats, it appeared that 111 ln-DTPA-D-Phe-1-octreotide: (1) is excreted via the kidneys, (2) shows only minor accumulation in the liver and (3) has an initial plasma half-life in the order of minutes (8).

In this study, we report data concerning the metabolism of intravenously administered ¹¹¹In-DTPA-D-Phe-1-octreotide in man, as well as estimates of its radiation dose in principal organs and the effective dose equivalent. Also, scintigraphic images of various somatostatin receptor-positive tumors have been compared using both ¹²³I-Tyr-3-octreotide and ¹¹¹In-DTPA-D-Phe-I-octreotide as radiopharmaceuticals in the same patients.

MATERIALS AND METHODS

Radiopharmaceuticals

The somatostatin derivatives DTPA-D-Phe-1-octreotide (SDZ 215-811) and Tyr-3-octreotide (SDZ 204-090) were prepared by Sandoz (Basel, Switzerland). Indium-111-chloride ("ultra-pure") and Na¹²³I were obtained from Mallinckrodt Diagnostics (Petten, The Netherlands) and Medgenix (Fleurus, Belgium), respectively. Indium-111-chloride contained ^{114m}In to a limited extent (0.5 kBq ¹¹⁴In/MBq ¹¹¹In at calibration time). Radiolabeling of DTPA-D-Phe-1-octreotide with ¹¹¹In and of Tyr-3-octreotide with ¹²³I and quality control of the products were performed as described before (5–8). Depending on the interval between injection and scintigraphy, and whether SPECT was required, the administered radioactivity of ¹¹¹In-DTPA-D-Phe-1-octreotide and of ¹²³I-Tyr-3-octreotide ranged from 185 to 259 MBq and from 370 to 555 MBq, respectively, given as an intravenous bolus. The administered dose of the somatostatin analogs varied from 7 to 20 μg per injection.

Imaging

Planar and SPECT images were obtained with a large field of view gamma camera (Counterbalance 3700 and ROTA II, Siemens, Hoffman Estates, IL), equipped with a medium-energy parallel-hole collimator.

For reprints contact: E. P. Krenning, MD, PhD, Professor of Nuclear Medicine, University Hospital Dijkzigt, Department of Nuclear Medicine, Dr Mole waterplein 40, 3015 GD Rotterdam, The Netherlands.

COPYRIGHT © 2020 by the Society of Nuclear Medicine and Molecular Imaging.

The pulse-height analyzer windows were centered over both 111In photon peaks (172 keV and 245 keV) with a window width of 20%. Data from both windows were added to the acquisition frames. The camera was connected to a dedicated PDP 11/73 computer (Digital Equipment Corp., Maynard, MA) using the Gamma 11 and SPETS V 6.1 software (Nuclear Diagnostics, Stockholm, Sweden). The acquisition parameters for planar images were: (1) 128 × 128 word matrix, (2) images of head/neck: 300,000 preset counts (or max. 15 min) at 24 hr and 15 min preset time $(\approx 200,000 \text{ counts})$ at 48 hr after injection and (3) images of the rest of the body: 500,000 counts (or max. 15 min); for SPECT: (1) 60 projections, (2) 64 × 64 word matrix and (3) 60-sec acquisition time per projection. SPECT analysis was performed with a Wiener filter on original data. The filtered data were reconstructed with a ramp filter. If indicated, SPECT studies were performed 4 hr (head and neck) or 24 hr (remainder of the body) after injection of the radiopharmaceutical. Planar studies were carried out both after 24 hr and 48 hr (vide supra), and in a few cases also after 0.5 hr and 4 hr with the same protocol used in the 24-hr studies. Total-body scintigraphy (scintigraphy of extremities only if indicated) of every patient was performed at least once, usually 24 hr after injection.

Measurement of ¹¹¹In Radioactivity in Blood, Urine and Feces

Radioactivity in blood, urine and feces was measured with an LKB-1282-Compugamma system or a GeLi-detector equipped with a multichannel analyser (Series 40, Canberra).

Blood samples were collected directly before injection and 2, 5, 10, 20 and 40 min and 1, 4, 20 and 48 hr after injection. Urine was collected from the time of injection in two 3-hr intervals and thereafter in intervals of 6 hr until 48 hr after injection. If feasible, feces was collected until 72 hr after injection. The chemical status of the radionuclide in blood and urine was analyzed as a function of time by using the SEP-PAK Cl8, HPLC and gel filtration techniques as described previously (5). The nature of peptide- bound radioactivity in blood and urine was tested by investigation of specific binding to somatostatin receptors on rat brain cortex cell membranes as described previously (9).

Patients

The data described in this paper were derived from patients who had been referred for a variety of potentially somatostatin receptor-positive tumors. All patients gave informed consent to participate in the study, which had been approved by the ethics committee of our hospital. Kinetic studies with ¹¹¹In-DTPA-D- Phe-l-octreotide by means of gamma camera scintigraphy were performed in 26 patients. Additionally, plasma, urine and feces samples were obtained from 9, 10, and 4 patients, respectively. In another six patients we were able to perform somatostatin-analog scintigraphy with ¹²³I-Tyr-3-octreotide as well as with ¹¹¹In-DTPA-D-Phe-l-octreotide with a maximum interval of 3 mo.

Dosimetry

For the estimation of the radiation dose, the MIRDOSE version 2 program (10) and ICRP publication 53 (11) were used. The uptakes in the most important source organs, the kidneys, the spleen, the liver, the urinary bladder and the remainder of the body, were determined as a function of time. Radioactivity in the kidneys, liver and spleen was calculated as described before (6). Radioactivity in the urinary bladder was calculated using the measured radioactivity excreted in the urine and assuming a bladder voiding interval of 3.5 hr (11). Radioactivity in the remainder of the body (expressed as percentage of the administered radioactivity) as a function of time was determined as 100% minus the % uptake in kidneys, liver, spleen, urinary bladder and excreted radioactivity in urine and feces.

In eight patients, the uptake in the kidneys, spleen and liver was measured with the gamma camera 0.5, 4, 24 and 48 hr after injection of ¹¹¹In-DTPA-D-Phe-1-octreotide. The results obtained in two patients could not be used because of an extensive overlap of the right kidney and the liver. In the six remaining patients, the excreted radioactivity in the urine (until 48 hr after ¹¹¹In-DTPA-D-Phe-1-octreotide injection) was measured as well. In four patients, the fecal excretion was also determined until 72 hr after injection. The calculation of the radiation dose in the gastrointestinal tract was performed using the mean radioactivity detected in the feces of these four patients. The dose estimates of the various organs and the effective dose equivalent were calculated for each of the six patients individually.

In order to perform dosimetry when only gamma camera measurements were available 24 and 48 hr after ¹¹¹In-DTPA-D-Phe-l-octreotide injection, a model was developed. In this model the residence times for the kidneys, spleen and liver were calculated for each individual patient on the basis of the organ uptakes after 24 and 48 hr. For the urinary bladder contents and the remainder of the body measurements, mean residence times were used. With this model, it was possible to calculate the dose estimates and the effective dose equivalent in another 18 patients. Apart from the organs mentioned above, radioactivity was seen in both the thyroid and pituitary glands in most patients.

RESULTS

Metabolism

The average plasma radioactivity decreased rapidly after injection of $^{111}\text{In-DTPA-D-Phe-1-octreotide}$ in nine patients. Assuming a plasma volume of 3 liters, the mean radioactivity in the blood circulation was calculated to decrease within 10 min to 33% \pm 7% (s.d.) of the injected amount. In five patients, the chemical status of the radionuclide in the plasma was investigated as a function of time. In Figure 1, the time course of total and peptide-bound radioactivity in the plasma of these five patients are presented up to 20 hr after injection of $^{111}\text{In-DTPA-D-Phe-1-octreotide}$. During the first 4 hr, plasma radio-activity was mainly

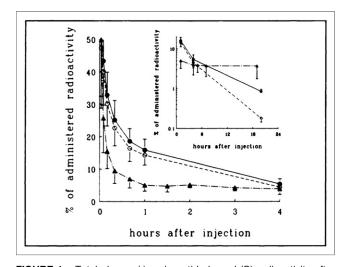


FIGURE 1. Total plasma (•) and peptide-bound (O) radioactivity after administration of $^{111}\text{ln-DTPA-D-Phe-1-octreotide}$ in five patients. Results are compared with total plasma (Δ) radioactivity after administration of $^{123}\text{l-Tyr-3-octreotide}$ in seven patients taken from reference 6. Data are expressed as mean \pm s.d. percentage of the administered radioactivity. Note the logarithmic scale of values in the ordinate of the inset.

peptide-bound in the form of intact ¹¹¹In-DTPA-D-Phe-l-octreotide as demonstrated by HPLC. Only the last two points of the observation period showed proportionally increasing amounts of nonpeptide-bound radioactivity, eluting in the void volume.

Urinary excretion of radioactivity was measured in ten patients. In five of these patients, the chemical status of the radionuclide in the urine was also investigated. Figure 2 shows the rapid excretion of the administered radioactivity via the urine from about 25% after 3 hr, 50% after 6 hr, 85% after 24 hr to over 90% after 48 hr for this group of five patients. Figure 2 also shows that the excreted radioactivity was mainly peptide-bound in these patients. SEP-PAK and HPLC analyses of urine and plasma samples as functions of time after injection demonstrated that peptide-bound radioactivity predominantly consisted of ¹¹¹In-DTPA-D-Phe-1-octreotide during the first hours after injection. Twenty-four hours after injection, in addition to ¹¹¹In-D-Phe-1-octreotide and peptide-bound degradation products, a major part of urinary radioactivity eluted from the HPLC in the void volume (Fig. 3). Feces, collected until 72 hr after injection of 111In-DTPA-D-Phe-l-octreotide from four patients with normal intestinal function, contained less than 2% of the administered radioactivity. The SEP-PAK C18 and HPLCpurified radiolabeled peptide component in plasma and urine showed the same biological activity as the radiopharmaceutical itself, as indicated by its specific binding to somatostatin receptors on rat brain cortex cell membranes (data not shown).

Dosimetry

The uptake of radioactivity in the liver, spleen and kidneys was measured with the gamma camera in six patients. If either the thyroid or the pituitary, or both, were clearly distinguishable from the surrounding tissue, the uptake in these organs was measured as well. Other organs (e.g., the intestines) showed low accumulation of radioactivity and were, therefore, disregarded in the gamma camera measurements. The time courses of radioactivity in the liver, spleen and kidneys showed close similarities between individual patients (data not shown). The results of the uptake

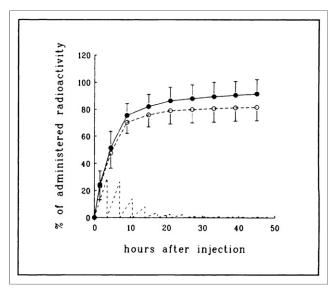


FIGURE 2. Cumulative total (•) and peptide-bound (O) ¹¹¹In urinary excretion (n = 5) and mean urinary bladder radioactivity voiding (—) (n = 5) after intravenous injection of ¹¹¹In-DTPA-D-Phe-1-octreotide. Data are expressed as mean \pm s.d. percentage of the administered radioactivity.

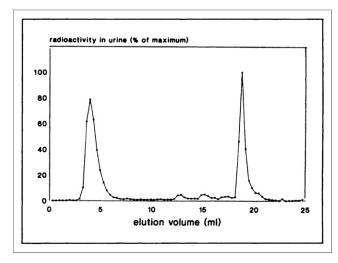


FIGURE 3. Typical HPLC-elution profile of human urine, collected 24 hr after intravenous injection of ¹¹¹ln-DTPA-D-Phe-1-octreotide. At a retention volume of about 4 ml, non-peptide-bound ¹¹¹ln, such as ¹¹¹ln-DTPA, is eluted at 12–18 ml ¹¹¹ln containing degradation products and at about 19 ml the original radioligand.

measurements are given in Table 1. Five thyroid and two pituitaries in the group of six patients could be completely distinguished from the surrounding background (vide infra). The radioactivity in the thyroid (maximal 0.03%) and the pituitary (maximal 0.003%) varied strongly between individual patients. Frequently, these organs were only visible on the 24-hr images. Therefore, the residence time was calculated assuming that the effective half-life equals the physical half-life. The absorbed doses for the thyroid and the pituitary (12,13) corresponding to the maximum uptake measured in the group of six patients, are given in Table 2. Radioactivity excreted in the urine was used for calculation of the residence time of the urinary bladder contents for all patients and

TABLE 1

Radioactivity in Selected Organs and Remainder of the Body (mean ± s.d.) as a Function of Time After Intravenous Administration of ¹¹¹In-DTPA-D-Phe1-Octreotide in Man on the Basis of Gamma Camera Measurements and Calculations Expressed as Percentage of the Administered Radioactivity

		%Update						
Time (hr)	n	Liver	Spleen	Kidney	Remainder of the body*			
0.5	6	2.8 ± 1.2	2.0 ± 0.9	6.8 ± 1.3	84.2 ± 7.7			
4	6	1.9 ± 0.4	2.4 ± 0.9	7.2 ± 1.8	46.7 ± 17.9			
24	6	2.2 ± 0.6	2.6 ± 1.1	6.3 ± 2.1	8.7 ± 9.7			
48	6	1.8 ± 0.4	1.8 ± 0.6	5.0 ± 2.0	7.2 ± 8.8			
24	18	3.0 ± 1.3	2.9 ± 1.6	4.8 ± 1.7				
48	18	2.5 ± 1.0	2.1 ± 1.2	3.5 ± 1.3				

^{*}The radioactivity in the remainder of the body is the administered radioactivity (100%) minus the %radioactivity in liver, spleen, kidneys, urinary bladder, urine and feces.

showed the same course as the mean depicted in Figure 2. For all patients, the radioactivity in the feces was taken to be the mean of the radioactivity measured in the four patients (0.5% and 1.7% after 24 and 48 hr, respectively). Even assuming that these maximum values occurred in the same patient, the contribution of the thyroid, the pituitary and the feces to the effective dose equivalent was less than 5% and could therefore be neglected. The dose estimates of the various organs and the effective dose equivalent were calculated for these six patients (data not shown).

The contribution to the absorbed dose caused by the \$^{114m}In\$ contamination was calculated on the basis of the information obtained from the manufacturer and was found to be less than 0.5% of the dose from \$^{111}In\$. In practice, the radionuclide was administered before the calibration time, thus lowering even more the contribution of ^{114m}In to the radiation dose.

On the basis of the data obtained in the six patients, it appears that more than 70% of the effective dose equivalent results from radioactivity accumulated in the kidneys, spleen and liver. The uptake of radioactivity in these organs showed much greater individual variation in these six patients than the radioactivity

TABLE 2

Dose Estimates After Intravenous Administration of 111 In-DTPA-D-Phe-1-Octreotide in Man on the Basis of Gamma Camera Measurements (n = 24), Urinary Excretion Measurements (n = 10) and Fecal Excretion Measurements (n = 4)

Target organ	Absorbed dose (mGy/MBq)	Range (mGy/MBq)
Kidneys	0.45 ^{MD}	0.19–0.80
Liver	0.07 ^{MD}	0.04-0.15
Spleen	0.32 ^{MD}	0.10-0.66
Gonads	0.019 ^{MD}	0.015-0.026
Red marrow	0.020 ^{MD}	0.016-0.026
Urinary bladder wall	0.18 ^{MN}	n.a.*
GI tract		
Small intestinal wall	0.03 ^{MN,†}	n.a.*
ULI wall	0.04 ^{MN,†}	n.a.*
LLI wall	0.06 ^{MN,†}	n.a.*
Thyroid gland	0.04 ^{MX}	
Pituitary	0.11 ^{MX}	
Median effective dose	Range (mSv/MBq)	
0.08		0.05-0.12

The input to the small intestines is taken to be the same as the fecal excretion during 72 hr (n=4), viz 1.7%. For the thyroid and the pituitary (n=6), the radiation dose corresponds to the maximum organ uptake measured in these patients.

excreted in the urine and the calculated uptake in the remainder of the body. Therefore, in the model employed, the uptake of radioactivity in the liver, kidneys and spleen is based on individual gamma camera measurements. Table 1 shows that in the group of six patients, radioactivity in the spleen increases slightly from 0.5 to 24 hr and decreases afterwards in all patients. In the liver, the radioactivity decreases rapidly during the first hour, thereafter an increase is measured until 24 hr, followed by a decrease. For the calculation of the residence time in the model, the radioactivity in the liver and the spleen was assumed to remain constant from 0 to 24 hr and to decrease after 24 hr monoexponentially with time. Radioactivity in the kidneys shows a steady decrease (Table 1) starting shortly after the injection of the radiopharmaceutical. For this reason, the calculation of the kidney residence time was based on a monoexponential curve. The initial uptake in the kidneys was calculated by extrapolating the uptakes at 24 and 48 hr to t = 0.

The periodic accumulation and discharge of radioactivity in the urinary bladder was calculated from the mean collected urinary radioactivity in 10 patients. There is no significant difference in the cumulative urinary radioactivity between these 10 patients and the 5 patients presented in Figure 2. The radioactivity in the remainder of the body was calculated by subtraction of the mean uptakes in the kidneys (n = 6), spleen (n = 6), liver (n = 6), urinary bladder contents (n = 10) and mean excreted radioactivity in urine (n = 10) and feces (n = 4) from the administered amount. The residence time in the remainder of the body could be determined by fitting a monoexponential curve to the radioactivity course after injection. The six patients were recalculated using the model described. There were no statistically significant differences in the dose estimates of the various organs and in the effective dose equivalent in comparison to the direct estimates for each individual (data not shown). Therefore, application of the model appeared to be valid. In 18 patients, the uptake in the kidneys, spleen and liver was measured after 24 and 48 hr (Table 1). These data did not differ significantly from the values obtained at the same time points in the six patients. Consequently, the model was applied to the additional 18 patients. The final dosimetric results for the complete group of 24 patients are shown in Table 2.

Iodine-123-Tyr-3-Octreotide and ¹¹¹In-DTP A-D-Phe-1-Octreotide Scintigraphy: Comparison in Localization of Tumor Tissue

A comparison between scintigraphy with ¹²³I-Tyr-3-octreotide and scintigraphy with 111In-DTPA-D-Phe-l-octreotide, performed in an interval of less than 3 mo for the same patients, is given in Table 3. In four of six patients (Patients 1-3, and 6) the results of the subsequent scintigrams were identical. In Patient 2, ¹²³I-Tyr-3octreotide scintigraphy showed accumulation of radioactivity in the region of the gallbladder, which is not unusual for this radiopharmaceutical, considering its extensive biliary clearance. However, the same was observed with 111In-DTPA-D-Phe-l-octreotide, which is unusual, since this radiopharmaceutical is largely excreted by the kidneys. SPECT with ¹¹¹In-DTPA-D-Phe-1-octreotide demonstrated a tumor located ventral to the gallbladder. In Patient 3, SPECT images after 111In-DTPA-D-Phe-1-octreotide injection revealed the localization of the insulinoma, medial and anterior to the spleen and left kidney respectively (Fig. 4). However, this localization had not been recognized on planar images with both radiopharmaceuticals. In Patient 4, presumed carcinoid deposits were more numerous using ¹¹¹1n-DTPA-D-Phe-1-octreotide scintigraphy.

MD denotes median.

^{MN}denotes mean.

 $^{^{\}mbox{\scriptsize MX}}\mbox{denotes}$ calculations for maximum uptake.

^{*}Not applicable due to the model.

[†]ICRP 30 GI model used.

TABLE 3
Patient Data and Results of Somatostatin-Receptor Scintigraphy with ¹²³I-Tyr-3-Octreotide and ¹¹¹In-DTPA-D-Phe-1-Octreotide

				Interval between	Abnormal sites of radioactive accumulation	
Patient no.	Sex	Age	Tumor type	scans	¹²³ l-octreotide	¹¹¹ In-octreotide
1	F	66	Small-cell lung cancer	5 wk	Right lung, lower lobe	Right lung, lower lobe
2	М	65	Gastrinoma	7 wk	Gallbladder region	Gallbladder region (see text)
3	F	59	Insulinoma	1 wk	None (SPECT not done)	None (SPECT positive, Fig. 4)
4	F	50	Carcinoid	10 wk	Left supraclavicular lymph node, liver	Left+right supraclavicular lymph node, abdomen, chest, liver
5	F	64	Carcinoid	5 wk	3 caudal abdominal sites, 1 cranial abdominal site	3 caudal abdominal sites, liver (gallbladder removed; see text)
6	F	28	Pheochromocytoma	4 wk	Lower left abdomen	Lower left abdomen

Tumor progression in the relatively long interval between the two scintigrams cannot be excluded, however. In Patient 5, the liver showed an irregular uptake pattern of ¹¹¹In-DTPA-D-Phe-1-octreotide, whereas a homogeneous distribution was seen with ¹²³I-Tyr³-3-octreo-tide scintigraphy. The cranial abdominal site of radionuclide accumulation observed with ¹²³I-Tyr-3-octreotide was not seen on the subsequent ¹¹¹In-DTPA-D-Phe-1-octreotide scintigram. This site very likely represented abnormal uptake in the gallbladder and the hepatoduodenal ligament, which had been surgically removed between the two scintigrams and were proven to be massively infiltrated by a carcinoid tumor.

Apart from the marked differences in hepatic and renal clearances (Fig. 5), tissue accumulation of both radiopharmaceuticals was similar except for the pituitary, which was nearly always visible with ¹¹¹In-DTPA-D-Phe-1-octreotide in contrast to ¹²³I-Tyr-3-octreotide scintigraphy. However, discrimination between accumulation of ¹¹¹In-DTPA-D-Phe-1-octreotide in the pituitary and in the surrounding tissues is often impossible, especially caudally. Consequently, only the contour of the cranial part of the pituitary can be distinguished since in the region of the brain the

S | L K S

FIGURE 4. Indium-111-DTPA-D-Phe-1-octreotide 24 hr SPECT image of Patient 3 with a solitary insulinoma in the tail of the pancreas, which is shown medial and anterior to the spleen and left kidney. The line on the reference image (left) indicates the position of the transversal slice (right). K = Kidney, K = Kidney,

(background) radioactivity is relatively very low because of the blood-brain barrier. For both radiopharmaceuticals, radioactivity was always observed in the thyroid, liver, spleen and kidneys, the urinary bladder and in the intestinal tract, although in the last organ ¹¹¹In-DTPA-D-Phe-1-octreotide was much less visible.

DISCUSSION

In spite of a lower affinity of ¹¹¹In-DTPA-D-Phe-1-octreotide for the rat brain somatostatin receptor compared to that of ¹²³I-Tyr-3-octreotide (7), the indiumlabeled compound visualized somatostatin receptor-positive animal tumors more efficiently in vivo, probably due to its different metabolic behavior (8). These metabolic properties turned out to be similar in man; our study shows that after intravenous administration ¹¹¹In-DTPA-D-Phe-1-octreotide is rapidly cleared from the circulation via the kidneys. However, its initial disappearance from the circulation is considerably slower compared to that of ¹²³I-Tyr-3-octreotide (Fig. 1) (6).

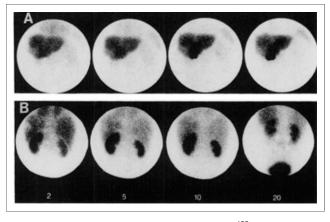


FIGURE 5. Sequential anterior abdominal views of ¹²³I-Tyr-3-octreotide scintigraphy (A) showing rapid hepatobiliary clearance, and posterior abdominal views of ¹¹¹In-DTPA-D-Phe-1-octreotide scintigraphy (B) showing rapid renal clearance. Note the rapidly decreasing blood-pool radioactivity over the heart with both radiopharmaceuticals. The 20-min image of the latter scintigrams also shows a low-grade accumulation of radioactivity in the lower part of the vertebral column due to a chondrosarcoma.

This slower initial clearance, combined with the longer physical half-life of 111 In ($t_{1/2} = 2.8$ days for m In versus 13.2 hr for 123 I) results in a longer residence time of the radiopharmaceutical in the tissues. The presence of a lower radioactivity in the remainder of the body 24 hr after injection of 111In-DTPA-D-Phe-1-octreotide leads to a lower background radioactivity (Fig. 1) (6). The higher background radioactivity with 123I-Tyr-3-octreotide is due to its much higher circulating levels of degradation products than is the case with 111In-DTPA-D-Phe-1-octreotide. Therefore, 111In-DTPA-D-Phe-1-octreotide is a more suitable radioligand to localize somatostatin receptor-rich tissues (vide infra). Furthermore, using ¹¹¹In-DTPA-D-Phe-1-octreotide, interpretation of scintigrams of the abdominal region is less affected by intestinal background radioactivity. This contrasts remarkably with 123I-Tyr-3-octreotide scintigraphy, because the hepatobiliary clearance of this compound results in a high hepatic and intestinal accumulation of radioactivity, which is hardly overcome with laxatives.

Analysis of the chemical status of plasma radioactivity during the first 4 hr after injection shows mainly peptide- bound ¹¹¹In in the form of the original ¹¹¹In-DTPA-D-Phe-1-octreotide. Similarly, analysis of radioactivity in the urine shows predominantly intact ¹¹¹In-DTPA-D-Phe-1-octreotide during the first hours after injection. Furthermore, peptide-bound radioactivity in plasma and urine has somatostatin receptor-binding properties as demonstrated by specific binding to rat brain cortex cell membranes. Degradation of ¹¹¹In-DTPA-D-Phe-1-octreotide was observed only in plasma and urine samples obtained more than 4 hr after intravenous injection of the radio-pharmaceutical, when circulating radioactivity amounted to less than 10% of the administered radioactivity. Ultimately, degradation to ¹¹¹In-labeled products such as ¹¹¹In-DTPA is suggested by the appearance of the peak in the void volume of HPLC analysis.

Accumulation of radioactivity after intravenous administration of ¹¹¹In-DTPA-D-Phe-l-octreotide in man is observed in the pituitary and thyroid gland, the spleen, liver, kidneys and the urinary bladder. Imaging of the gallbladder is occasionally seen, whereas the presence of intestinal radioactivity (mainly in the colon at 24 hr) depends on the simultaneous use of laxatives. The relatively low clearance of the radioligand via the hepatobiliary system favors its use in SPECT of the abdomen, which is strongly indicated in the localization of small endocrine pancreatic tumors. The mechanism of the thyroid gland imaging is still unclear. With radiolabeled somatostatin analogue autoradiography, we could not find somatostatin receptors in normal thyroid gland and differentiated thyroid carcinoma (papillary cancer) tissue slices (14,15). However, a high percentage of malignant parafollicular thyroid tumors are somatostatin receptor-positive by both autoradiography and scintigraphy (14,15). It is remarkable that in the two cases with Graves' hyperthyroidism investigated so far, accumulation of radioactivity in the thyroid was increased (unpublished data). The presence of lymphocytes [which can be somatostatin receptor-positive (16)] in the thyroid could explain this observation. Autoradiography of normal spleen tissue revealed the presence of somatostatin receptors (unpublished); however, the exact cell type bearing the somatostatin receptor has not been identified yet. Patients on octreotide treatment show a diminished accumulation of radioligand in the spleen (unpublished), compatible with occupancy of spleen somatostatin receptors by the unlabeled octreotide.

The rapid appearance of intact ¹¹¹In-DTPA-D-Phe-l-octreotide in the urine indicates an effective renal clearance of this radio-

pharmaceutical. By contrast, ¹²³I-Tyr-3-octreotide is rapidly cleared by the liver and little of it is excreted intact into the urine. The different metabolism of ¹¹¹In-DTPA-D-Phe-1-octreotide compared with ¹²³I-Tyr-3-octreotide indicates that the modification of octreotide with the ¹¹¹In-DTPA group inhibits hepatic clearance and/or facilitates renal clearance. Rat liver perfusion studies have indeed shown that ¹¹¹In-DTPA-D-Phe-1-octreotide is cleared much more slowly by the liver than ¹²³I-Tyr-3-octreotide (unpublished data). It is unknown whether the effect of the ¹¹¹In-DTPA group on the metabolic routing of peptides is a general phenomenon. The relatively long residence time of ¹¹¹In-DTPA-D-Phe-1-octreotide in the kidneys suggests that following glomerular filtration part of the label is reabsorbed in the tubules (8).

The higher sensitivity of ¹¹¹In-DTPA-D-Phe-1-octreotide compared to ¹²³I-Tyr-3-octreotide in localizing the pituitary is remarkable, since in vitro studies have shown a higher affinity of the radioiodinated ligand for somatostatin receptors in rat brain (7). However, its actual affinity for somatostatin receptors on the pituitary is at present unknown.

For several somatostatin receptor-positive tumors, ¹¹¹In-DTPA-D-Phe-1-octreotide shares with ¹²³I-Tyr-3-octreo-tide the advantage of providing a more sensitive imaging technique when compared to currently available diagnostic procedures, e.g., CT, ultrasound and MRI. Furthermore, the future availability of both DTPA-D-Phe-1-octreotide and pure ¹¹¹InCl₃, will make an easy single-step labeling procedure possible and, hence, scintigraphy of somatostatin receptor-positive tumors generally available. The effective dose equivalent, although higher than that of ¹²³I-Tyr-3-octreotide, is comparable with values for other ¹¹¹In-labeled radiopharmaceuticals (*11*) and is acceptable in view of the clinical indications.

ACKNOWLEDGMENTS

The authors thank Ina Loeve, Marianne Goemaat-Visser, Marcel van der Pluijm and Michiel de Roo for their expert technical assistance and Michael Stabin for providing the S-value tables for ^{114m}In.

REFERENCES

- Krenning EP, Bakker WH, Breeman WAP, et al. Localisation of endocrinerelated tumours with radioiodinated analogue of somatostatin. *Lancet* 1989;1: 242-244
- Lamberts SWJ, Bakker WH, Reubi J-C, Krenning EP. Somatostatin-receptor imaging in the localization of endocrine tumors. N Engl J Med 1990;323: 1246–1249.
- Lamberts SWJ, Hofland LJ, van Koetsveld PM, et al. Parallel in vivo and in vitro detection of functional somatostatin receptors in human endocrine pancreatic tumors: consequences with regard to diagnosis, localization, and therapy. J Clin Endocrinol Metab 1990;71:566–574.
- Kwekkeboom DJ, Krenning EP, Bakker WH, et al. Radioiodinated somatostatin analog scintigraphy in small-cell lung cancer. J Nucl Med 1991; 32: 1845–1848.
- Bakker WH, Krenning EP, Breeman WAP, et al. Receptor scintigraphy with a radioiodinated somatostatin analogue: radiolabeling, purification, biologic activity and in vivo application in animals. J Nucl Med 1990;31: 1501–1509.
- Bakker WH, Krenning EP, Breeman WAP, et al. In vivo use of a radioiodinated somatostatin analogue: dynamics, metabolism and binding to somatostatin receptor positive tumors in man. J Nucl Med 1991;32: 1184–1189.
- Bakker WH, Albert R, Bruns C, et al. [111In-DTPA-D-Phe1]-octreotide, a
 potential radiopharmaceutical for imaging of somatostatin receptor-positive
 tumors: synthesis, radiolabeling and in vitro validation. *Life Sci* 1991; 49:
 1583–1591.

- Bakker WH, Krenning EP, Reubi JC, et al. In vivo application of [111In- DTPA-D-Phe¹]-octreotide for detection of somatostatin receptor-positive tumors in rats. Life Sci 1991;49:1593–1601.
- Reubi JC, Maurer R, Klijn JGM, et al. High incidence of somatostatin receptors in human meningiomas: biochemical characterization. J Clin Endocrinol Metab 1986;63:433–438
- Watson EE, Stabin M, Bolch WE. Documentation package for MIRDOSE (version 2). 1988, Oak Ridge Associated Universities.
- International Commission on Radiological Protection. Radiation dose to patients from radiopharmaceuticals. ICRP Publication 53, Oxford: Pergamon Press; 1988.
- Siegel JA, Stabin MG. Absorbed fractions for electrons and bèta particles in small spheres (Abstract). J Nucl Med 1988,29:803.
- Ellett WH, Humes RM. Absorbed fractions for small volumes containing photon-emitting radioactivity. M1RD Pamphlet NO. 8. J Nucl Med 1971; 12(suppl 5):25–32.
- Reubi JC, Modgliani E, Calmettes C, et al. In vitro and in vivo identification of somatostatin receptors in medullary thyroid carcinomas, pheochromocytomas and paragangliomas. In: Calmettes C, Guliana JM, eds. *Medullary thyroid car*cinoma. Colloque INSERM/John Libey Eurotext Ltd.; 1991;211:85–87.
- Reubi JC, Chayvialle JA, Franc B, et al. Somatostatin receptors and somatostatin content in medullary thyroid carcinomas. *Lab Invest* 1991; 64:567–573.
- Sreedharan SP, Kodama KT, Peterson KE, Goetzl EJ. Distinct subsets of somatostatin receptors on cultured human lymphocytes. *J Biol Chem* 1989; 264: 949–953.