Initial Clinical Results of a Novel Immuno-PET Theranostic Probe in Human Epidermal Growth Factor Receptor 2–Negative Breast Cancer

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This prospective study evaluated the imaging performance of a novel pretargeting immunologic PET (immuno-PET) method in patients with human epidermal growth factor receptor 2 (HER2)–negative, carcinoembryonic antigen (CEA)–positive metastatic breast cancer, compared with CT, bone MRI, and ¹⁸F-FDG PET.

Methods: Twenty-three patients underwent whole-body immuno-PET after injection of 150 MBq of ⁶⁸Ga-IMP288, a histamine-succinyl-glycine peptide given after initial targeting of a trivalent anti-CEA, bispecific, antipeptide antibody. The gold standards were histology and imaging follow-up. Tumor SUVs (SUV_{max} and SUV_{mean}) were measured, and tumor burden was analyzed using total tumor volume and total lesion activity. **Results:** The total lesion sensitivity of immuno-PET and ¹⁸F-FDG PET were 94.7% (1,116/1,178) and 89.6% (1,056/1,178), respectively. Immuno-PET had a somewhat higher sensitivity than CT or ¹⁸F-FDG PET in lymph nodes (92.4% vs. 69.7% and 89.4%, respectively) and liver metastases (97.3% vs. 92.1% and 94.8%, respectively), whereas sensitivity was lower for lung metastases (48.3% vs. 100% and 75.9%, respectively). Immuno-PET showed higher sensitivity than MRI or ¹⁸F-FDG PET for bone lesions (95.8% vs. 90.7% and 89.3%, respectively). In contrast to ¹⁸F-FDG PET, immuno-PET disclosed brain metastases. Despite equivalent tumor SUV_{max}, SUV_{mean}, and total tumor volume, total lesion activity was significantly higher with immuno-PET than with ¹⁸F-FDG PET ($P = 0.009$). **Conclusion:** Immuno-PET using anti-CEA/anti-IMP288 bispecific antibody, followed by ⁶⁸Ga-IMP288, is a potentially sensitive theranostic imaging method for HER2-negative, CEA-positive metastatic breast cancer patients and warrants further research.

Key Words: breast cancer; immuno-PET; pretargeting; bispecific antibody; ⁶⁸Ga

J Nucl Med 2020; 61:1205–1211

DOI: 10.2967/jnumed.119.236000

Breast cancer (BC) is the most common cancer among women worldwide, and about 40% of all BC patients have a recurrence (1). Imaging plays an important role in BC management, and PET using ¹⁸F-FDG is recommended for relapse detection and therapy assessment (2). However, ¹⁸F-FDG is not an oncology-specific tracer and does not allow selection of patients for hormonal or targeted therapies. Recently, ¹⁸F-fluoroestradiol was approved for the characterization of estrogen receptor expression in known or suspected metastatic lesions of estrogen receptor–positive BC (3).

In the last decade, a promising option to improve diagnostic and theranostic imaging has been immunologic PET (immuno-PET), which combines the high sensitivity and quantitative capabilities of PET with the specificity and selectivity of a monoclonal antibody (mAb) against a given tumor cell-surface marker. This combination potentially provides a noninvasive, in vivo–quantifiable, 3-dimensional whole-body tumor biomarker expression cartography, permitting tumor detection and patient monitoring prospectively (4). In BC, clinical studies have reported a good performance for immunoconjugates targeting human epidermal growth factor receptor 2 (HER2) radiolabeled with PET emitters having different half-lives (⁶⁸Ga, ⁶⁴Cu, and ⁸⁹Zr) (5,6), and in preclinical studies, several biomarkers, such as syndecan-1, have been considered for triple-negative BC (7,8).

Carcinoembryonic antigen (CEA), specifically CEA cell-adhesion molecule-5, represents an attractive target in BC, with serum CEA positivity (i.e., ≥ 5 ug/L) being observed in 50%–60% of patients with metastatic disease (9), with a false-positive rate of 5% (10). A recent metaanalysis showed that elevated CEA was significantly associated with poorer disease-free survival and overall survival in BC, independent of tumor size, lymph node metastasis, or tumor grade and irrespective of estrogen receptor or HER2 status (11,12). Elevated CEA was more related to the non–triple-negative and nonluminal tumor type and older age (12). Moreover, most invasive lobular, tubular, and cribriform carcinomas were CEA-negative (72%). Conversely, 70% of invasive ductal carcinomas were CEA-positive (13).

Received Sep. 6, 2019; revision accepted Dec. 4, 2019.

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Published online Mar. 13, 2020.

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In the 1970s and later, the potential of cancer imaging using ^{131}I - and $^{99\text{m}}\text{Tc}$ -labeled anti-CEA antibodies was reported for CEA-positive tumors, including BC, but the method was limited by the poor resolution of SPECT and the high blood activity observed with directly radiolabeled mAbs and fragments (14–17). Anti-CEA targeted therapies are still under development, and labetuzumab govitecan (anti-CEA cell-adhesion molecule-5/SN-38 antibody–drug conjugate) recently has shown promising efficacy with manageable toxicity in metastatic colorectal cancer patients (18). Durable objective responses in highly pretreated metastatic triple-negative BC patients, as well as in estrogen receptor–positive, HER2-negative BC, were reported with an antibody–drug conjugate that targets the human trophoblast cell-surface antigen-2, suggesting the advance of a targeted drug conjugate platform for solid tumors (19,20).

Pretargeting refers to a system of improved image contrast and high sensitivity obtained by first administering an anti-CEA bispecific antibody (BsmAb), followed by haptens labeled with ^{111}In or ^{131}I (21–23). TF2 is an engineered trivalent BsmAb composed of a humanized anti–histamine-succinylglycine antigen-binding fragment derived from the murine 679 antibody and 2 humanized anti-CEA antigen-binding fragments (trivalency) of the humanized hMN-14 antibody, formed into a 157-kD protein by the dock-and-lock procedure (24). The ^{68}Ga -IMP288 bivalent peptide pretargeted with TF2 has demonstrated promising immuno-PET performance in a pilot optimization clinical trial (25). The current prospective study was designed to assess the initial sensitivity, safety, and preferred conditions of pretargeting immuno-PET using the BsmAb TF2 and the ^{68}Ga -IMP288 peptide, in comparison to ^{18}F -FDG PET and conventional imaging in relapsed HER2-negative BC patients.

MATERIALS AND METHODS

Population

Patients at least 18 y old, with progressive HER2-negative metastatic BC after standard treatments and presenting with a CEA serum level of at least 10 ng/mL and at least 1 lesion 10 mm or larger on conventional imaging (CT, MRI, and bone scans), were eligible. The other inclusion criteria were a Karnofsky performance status of at least 70, a minimum life expectancy of 6 mo, a creatinine level of no more than 2.5 times the normal level, and normal serum human antimouse antibody and human antihuman antibody (HAHA) titers. Women of child-bearing potential had to have a negative pregnancy test result. The exclusion criteria were pregnancy and breast feeding, any serious active disease or comorbid medical condition, any history of another cancer during the last 5 y, with the exception of nonmelanoma skin tumors or stage 0 (in situ) cervical carcinoma, and a known hypersensitivity to antibodies or proteins. In the 4 wk preceding immuno-PET, a staging workup that included a complete history, physical examination, CEA, and cancer antigen 15-3 serum level measurements (with biomarker doubling time determination when possible) were performed. The patients underwent morphologic imaging (thoracic-abdominal-pelvic contrast-enhanced CT for visceral lesion evaluation and pelvic-spine MRI for bone evaluation) and ^{18}F -FDG PET. The trial, sponsored by Nantes University Hospital, was approved by the responsible ethics committee (Comités de Protection des Personnes) and registered at ClinicalTrials.gov (NCT01730612), and all patients gave written informed consent. Enrollment was at the Institut de Cancérologie de l'Ouest, associated with Nantes University Hospital of the health

cooperation group of Institut Régional de la Cancérologie Nantes Atlantique.

Investigational Products and Study Design

The reagents were prepared for human use by Immunomedics, Inc., with a sufficient quantity being produced to complete a small feasibility and safety study. A 1.85-GBq (at calibration time) pharmaceutical-grade ^{68}Ga generator (Eckert-Ziegler) was used. ^{68}Ga -IMP288 was obtained with a specific activity of 40–100 MBq/nmol and a radiochemical purity of more than 95%.

TF2 diluted in 250 mL of 0.9% NaCl, and ^{68}Ga -IMP288 diluted in 50 mL of 0.9% NaCl, were administered by intravenous infusion over 30 min. The patients were premedicated with oral antihistamine the day before TF2 infusion and with intravenous antihistamine (5 mg, dexchlorpheniramine) and corticosteroid (500 mg, hydrocortisone hemisuccinate) 5 min before infusion of 60–120 nmol of TF2, and 3–6 nmol of ^{68}Ga -IMP288 were administered 24–30 h later. Four different conditions of pretargeting (molar doses of BsmAb and peptide and delay) were assessed for pharmacokinetics optimization purposes. Because there were no statistically significant differences among the cohorts, the results for the 4 cohorts were combined to compare the sensitivity of immuno-PET with that of ^{18}F -FDG PET or conventional imaging.

Safety was assessed through vital signs, physical examination, and adverse events. HAHA titers were determined by Immunomedics at 3 or 6 mo, and up to 9 mo in some cases, using an enzyme-linked immunosorbent assay (abnormal when ≥ 50 ng/mL) (25).

Immuno-PET Imaging

PET/CT was performed using a 4-ring Siemens Biograph mCT system 60 and 120 min after injection of 150 MBq of ^{68}Ga -IMP288 and reconstructed using 3-dimensional ordinary Poisson ordered-subsets expectation maximization with point-spread-function correction and time-of-flight mode (3 iterations, 21 subsets, gaussian postfiltering of 2 mm in full width at half maximum, $4 \times 4 \times 2$ mm voxels). Whole-body acquisitions were performed under spontaneous breathing for 2.5 min per bed position. CT was obtained using variable mAs, 120 kVp, and a pitch of 1 without contrast enhancement. Acquisitions were performed from the top of the head to mid thigh (6–8 steps per patient).

Qualitative Imaging Analysis

Immuno-PET abnormal uptake was defined visually as a focal increase in uptake higher than the surrounding background level. If more than 10 lesions per bone or more than 10 lesions for the entire liver were counted, the lesion number was capped at 10. With masking to the other diagnostic results, CT and bone MR images were analyzed by the consensus of 2 radiologists with expertise in oncology, and ^{18}F -FDG PET and immuno-PET images were analyzed by the consensus of 2 nuclear medicine physicians with expertise in immunotargeting and PET. For both ethical and practical reasons, not every suspected lesion was evaluated by histology. Complementary imaging, primarily MRI, was performed to assess the most important lesions suspected by immuno-PET and not detected by the initial work-up (CT and bone MRI). The gold standard was therefore determined on the basis of histology and imaging follow-up. Indeed, ^{18}F -FDG PET, CT, pelvic-spinal MRI, and any additional imaging were performed 3 mo after immuno-PET to confirm the abnormalities.

True-positive (TP) results were abnormal image findings confirmed by histology or detected by at least 1 other imaging modality and confirmed by follow-up. A negative finding on an imaging method was considered to be false-negative (FN) if positive on 1 other imaging method plus histopathology or by 1 other imaging method

and confirmed by follow-up. Percentage sensitivity $[(TP/TP + FN) \times 100]$ on a lesion basis was calculated for each imaging modality.

Semiquantitative Immuno-PET and ^{18}F -FDG PET Analyses

With a threshold of 40%, tumor SUV_{max} and SUV_{mean} were measured on tumor foci for immuno-PET and ^{18}F -FDG PET, and the partial-volume effect was corrected. Tumor burden was analyzed using total tumor volume (the sum in cubic centimeters of the volume of each positive lesion) and total lesion activity (tumor volume $\times \text{SUV}_{\text{mean}}$) obtained automatically with Oncoplanet software (version 2.0 RC; Dosisoft).

Statistical Analysis

The continuous variables (tumor SUV_{max} , tumor SUV_{mean} , total tumor volume, and total lesion activity) were described by median and interquartile range (IQR). Variable comparisons between ^{18}F -FDG PET and immuno-PET were performed by means of the Wilcoxon signed-rank test. All comparisons were 2-sided, with a significance limit of less than 5%. All calculations were made using the Stata SE 13.1 statistical tool (StataCorp LP).

RESULTS

Patient Characteristics and Adverse Events

The demographics of the 23 metastatic HER2-negative BC patients, all women, are presented in Tables 1 and 2. Two patients

TABLE 1
Patient Characteristics, Part 1

Characteristic	Data
Total number of patients	23
Median age (y)	61 (range, 37–80)
Initial tumor classification (n)	
T1	7
T2	14
T4	2
Initial stage UICC (n)	
Stage I	6
Stage IIA	10
Stage I/IB	5
Stage IIIB	2
Histology (n)	
Ductal	20
Lobular	3
Hormone receptor status (n)	
ER+ and PR+	14
ER+ and PR–	7
ER– and PR+	1
ER– and PR–	1
Histologic grade (n)	
I	1
II	19
III	3
Premenopausal status (n)	2
Postmenopausal status (n)	21

ER = estrogen receptor; PR = progesterone receptor.

TABLE 2
Patient Characteristics, Part 2

Characteristic	Data
Primary tumor treatment (n)	
Surgery	
Yes	21
No	2
Radiotherapy	
Yes	20
No	3
Adjuvant chemotherapy	
Yes	22
No	1
Adjuvant hormone therapy	
Yes	22
No	1
Number of earlier regimens (chemo- or hormone therapy)	
1	6
2	1
3	5
>3	11
Median CEA serum level ($\mu\text{g/L}$)	73.45 (range, 35.1–111.8)
Median cancer antigen 15-3 serum level (kU/L)	112.6 (range, 12.4–3,000)
Median CEA doubling time (mo)	4.95 (range, 0.3–330.1)
Median cancer antigen 15-3 doubling time (mo)	4.75 (range, 0.0–94.1)

benefited from a second immuno-PET coupled with a second ^{18}F -FDG PET, accounting for 25 immuno-PET and ^{18}F -FDG PET studies in the 23 patients.

No patient experienced an anaphylactic reaction during or after the TF2 BsmAb or hapten infusions. HAHA levels were abnormal in 4 of 25 (16%) patients but normalized within 9 mo.

Imaging Results and Sensitivity

As shown in a previous study with the same radiopharmaceutical (25), the physiologic distribution of the tracer showed moderate liver and spleen uptake, renal elimination, and presence in the heart and blood vessels due to the circulation of the radiopharmaceutical.

In total, 1,178 lesions were confirmed according to the gold standard: 66 in lymph nodes, 29 in the lungs, 153 in the liver, 919 in bone or bone marrow, and 11 in other sites (1 cutaneous, 3 adrenal, 1 ovarian, 1 mammary, and 5 cerebral). All patients had evidence of metastatic disease on immuno-PET, ^{18}F -FDG PET, CT, and bone MRI (Table 3).

In total, 1,116 foci were detected by immuno-PET, all confirmed as TP according to the gold standard, and 62 lesions were missed (FN lesions), predominantly in lung and bone. All immuno-PET lesions were seen at 60 as well as 120 min. ^{18}F -FDG PET, CT, and bone MRI detected 1,056, 216, and 412 TP lesions,

TABLE 3
Sensitivity of Immuno-PET, ^{18}F -FDG PET, and Conventional Imaging

Location	Immuno-PET	CT	^{18}F -FDG PET	Axial bone MRI
Overall	1,116 of 1,178 (94.7%)	Not applicable	1,056 of 1,178 (89.6%)	Not applicable
Lymph nodes	61 of 66 (92.4%)	46 of 66 (69.7%)	59 of 66 (89.4%)	Not applicable
Bone	881 of 919 (95.8%)	Not applicable	821 of 919 (89.3%)	412 of 454 (90.7%)
Liver	149 of 153 (97.3%)	141 of 153 (92.1%)	145 of 153 (94.8%)	Not applicable
Lung	14 of 29 (48.3%)	29 of 29 (100%)	22 of 29 (75.9%)	Not applicable

respectively, and presented 122, 35, and 42 FN results, respectively (Table 3). The mammary and cutaneous sites were detected by CT, ^{18}F -FDG PET and immuno-PET, whereas CT missed the adrenal lesions and ^{18}F -FDG PET missed the ovarian lesion. Interestingly, immuno-PET detected 5 brain lesions confirmed by brain MRI, whereas brain imaging was difficult by ^{18}F -FDG PET (4 FN), because of physiologic cerebral ^{18}F -FDG uptake (Fig. 1).

Immuno-PET showed a somewhat higher overall sensitivity (94.7%) than ^{18}F -FDG PET (89.6%). Regarding the different metastatic sites, immuno-PET also had a higher sensitivity than CT and ^{18}F -FDG PET for lymph node (92.4% vs. 69.7% and 89.4%, respectively) and liver (97.3% vs. 92.1% and 94.8%, respectively), whereas sensitivity was lower for lung metastases (48.3% vs. 100% and 75.9%, respectively). Immuno-PET had a slightly higher sensitivity for bone evaluation than MRI or ^{18}F -FDG PET (95.8% vs. 90.7% and 89.3%, respectively) (Fig. 2).

The median size of liver, lung, and lymph node lesions were 24 mm (range, 10–103 mm), 10.5 mm (range, 6–19 mm), and 28 mm (range, 9–43 mm), respectively. Of all the lesions identified by ^{18}F -FDG and immuno-PET, 3 (a lymph node lesion, a bone lesion, and a cutaneous lesion) were evaluated by immunohistochemistry. Diffuse and heterogeneous staining (1+ to 3+ intensity) was seen in over 50% of the carcinoma cells (example of lymph node staining is shown in Fig. 3).

PET Quantitative Metrics

For determination of immuno-PET SUV, total tumor volume, and total lesion activity, only the 15 patients injected with optimized pretargeting parameters according to the study of Bodet-Milin et al. were considered, because these cohorts represent optimal pretargeting conditions that would be applied if the technique were used in clinical practice (i.e., a BsMab-to-peptide mole ratio of 20 or 40 and a 30-h pretargeting delay) (25). Tumor uptake did not significantly differ between immuno-PET

and ^{18}F -FDG PET, with a median SUV_{max} of 23.83 (IQR, 9.09–44.65) and 15.87 (IQR, 11.70–18.87), respectively ($P = 0.088$), and a median SUV_{mean} of 4.90 (IQR, 3.00–8.16) and 4.71 (IQR, 3.49–5.62), respectively ($P = 0.125$) (Table 4). Tumor burden evaluated by functional volumes was equivalent between the 2 PET methods, with median total tumor volume equal to 294 (IQR, 159–558) for immuno-PET and 299 (IQR, 139–410) for ^{18}F -FDG PET ($P = 0.256$), whereas tumor activity total lesion activity was significantly higher with immuno-PET (median of 2,123; IQR, 995–5,304) than with ^{18}F -FDG PET (median of 1,197; IQR, 355–2,433; $P = 0.009$). When considering the entire patient population, as described in Figure 4, a correlation was found between the SUV_{max} of the most intense lesion in immuno-PET and the serum CEA level ($P = 0.0396$), whereas no correlation was found between ^{18}F -FDG PET tumor SUV_{max} and the CEA serum level.

DISCUSSION

CEA represents a potential target for antibody-based imaging or therapy in several solid tumors, including estrogen receptor–positive, HER2-negative, and triple-negative BC (14–18,26). Moreover, the discrepancy between serum CEA levels and CEA tissue expression in patients with BC is well known. Whereas immunohistochemistry shows positive CEA expression in 70%–90%, serum CEA levels are often within the reference range (16). Further, immunohistochemistry is often performed only on a tumor biopsy sample (i.e., a small fragment of the disease), whereas an anti-CEA PET imaging study will allow an evaluation of CEA expression in tumors disseminated throughout the whole body. Indeed, it is possible that an examination of the heterogeneity of CEA expression in the disseminated tumors, as elucidated by imaging, may improve the prediction of antitumor responses. This initial study is the first demonstrating the excellent sensitivity of anti-CEA pretargeted immuno-PET in HER2-negative metastatic BC, suggesting its potential for tumor imaging or theranostic approaches in this subtype of BC or other CEA-expressing tumors.

In this study, pretargeted immuno-PET achieved a 94.7% overall sensitivity, with a somewhat better sensitivity than morphologic imaging or ^{18}F -FDG PET for lymph node, liver, and bone examinations in BC patients with metastatic disease. As described in the literature, the number of FN results by ^{18}F -FDG PET may be accentuated by most patients having a hormonal status of estrogen receptor–positive or progesterone receptor–positive (2). However, in contrast to ^{18}F -FDG PET, immuno-PET detected brain metastases, potentially providing improved patient management due to the possibility of theranostic targeting of cerebral dissemination. Disappointingly, immuno-PET was less effective than CT or ^{18}F -FDG PET for

FIGURE 1. (A and B) Pretargeted immuno-PET with TF2 and ^{68}Ga -IMP288 peptide reveals 2 unknown brain metastases (right frontal and right frontoparietal, arrows). (C) T1-weighted gadolinium MR brain image confirms both asymptomatic lesions (arrows). (D) ^{18}F -FDG PET image shows only 1 metastasis (right frontal lesion, arrow).

FIGURE 2. (A–C) In patient 1, pretargeted immuno-PET with TF2 and ^{68}Ga -IMP288 peptide images show 2 vertebral metastases (L1 and T9, arrows) (A), ^{18}F -FDG PET discloses no vertebral abnormalities (B), and vertebral MRI confirms both lesions (blue arrows) and discloses another (red arrow) at T8 (C). (D–F) In patient 2, CT shows suspected liver lesion (D) and pretargeted immuno-PET with TF2 and ^{68}Ga -IMP288 peptide reveals high uptake by liver lesion (arrow) (E), which was not seen by ^{18}F -FDG PET (F).

lung metastasis detection, whereas the sensitivity of CT and ^{18}F -FDG PET was consistent with that reported previously (27,28). ^{68}Ga -PET has a larger range than ^{18}F (6.10 vs. 0.54 mm in lung tissue, respectively), reducing PET image resolution, whereas the spontaneous-breathing PET/CT acquisition associated with a partial volume may affect PET performance with ^{68}Ga more than that with ^{18}F for detection of small lung lesions (median value of 10.5 mm in this cohort) (29). Moreover, Laessig et al. reported a decreased expression of CEA in lung metastases, compared with liver and bone metastases (30), as may explain the poorer performance of immuno-PET in this region. However, labeling of the peptide with ^{18}F , as described previously, may improve the detection of small lung lesions (31)

FIGURE 3. A 45-mm left axillary lymphadenopathy excision in patient with initial left ductal BC, 5% progesterone receptor, 50% estrogen receptor, and HER2-negative. (A) Carcinoma proliferation surrounded by fibrous capsule (hematoxylin, eosin, and saffron, $\times 200$). (B) Diffuse and heterogeneous (+ to +++) CEA membrane expression in over 50% of carcinoma cells (immunohistochemical study, $\times 200$).

without altering radiation exposure, since ^{68}Ga and ^{18}F have similar exposure rates ($1.79 \cdot 10^{-4} \text{ mSv} \cdot \text{m}^2 \cdot \text{MBq}^{-1} \cdot \text{h}^{-1}$ for ^{68}Ga and $1.87 \cdot 10^{-4} \text{ mSv} \cdot \text{m}^2 \cdot \text{MBq}^{-1} \cdot \text{h}^{-1}$ for ^{18}F).

CEA, especially CEA cell-adhesion molecule-5, functions as a cell-adhesion molecule during tumor invasion of the lymphatic lumen, possibly explaining the high sensitivity for lymph node detection (32). Indeed, immunohistochemistry confirmed a strong cellular CEA expression in a lymph node showing high uptake in 1 case. Finally, even if imaging specificity requiring histologic confirmation of false-positive lesions was not determined in this cohort of patients with diffuse disease, immuno-PET is expected to have a higher specificity than ^{18}F -FDG PET. This expectation needs further study.

Despite the absence of a significant difference between immuno-PET and ^{18}F -FDG PET for tumor SUV_{max} and SUV_{mean} , which reflect the activity of the most intense lesion, the total lesion activity, reflecting the tumor volume and uptake intensity of all whole-body foci, was significantly higher for immuno-PET than for ^{18}F -FDG PET. The total tumor volume, being comparable for the 2 PET methods, suggests that using CEA targeting will result in a higher tumor intensity with whole-body imaging than when using a glucose metabolism tracer, such as in ^{18}F -FDG PET.

This initial study also confirmed that intravenous injections of corticosteroids and antihistamines before TF2 and IMP-288 peptide infusions may induce transient immunodepression, limiting immediate and delayed immune reactions. The 16% HAHA response rate resolved at 9 mo in all cases and was similar to that reported by Bodet-Milin et al. previously and lower than that reported by Schoffelen et al. (52%) (25,33).

Finally, immuno-PET with the 2-step pretargeting method described herein enables the use of short-half-life PET emitters, such as ^{68}Ga or ^{18}F , allowing image acquisition only a few hours after peptide injection, whereas using an intact IgG instead of an antigen-binding fragment requires imaging a few days after the mAb injection for optimal contrast (because of the delayed immunoglobulin tumor targeting and clearance from the circulation). Intact IgG requires radiolabeling with long-half-life radionuclides, such as ^{89}Zr or ^{124}I , which are less favorable in terms of dosimetry. However, pretargeting approaches require optimization studies to determine the best BsmAb and peptide molar dose ratio and pretargeting delay. The TF2/ ^{68}Ga -IMP288 system used here has already been evaluated in a study of medullary thyroid cancer patients, concluding that a 120-nmol BsmAb dose, a molar ratio of 20, and a 30-h pretargeting delay represent optimal conditions (25). The present study on BC patients confirmed the increased tumor uptake and contrast using 120 nmol of BsmAb and a 30-h pretargeting delay, compared with a 60-nmol BsmAb dose or a 24-h pretargeting delay (Supplemental Table 1; supplemental materials are available at <http://jnm.snmjournals.org>), but this finding requires confirmation in a larger study.

TABLE 4
PET Semiquantitative Analyses

Parameter	Immuno-PET		¹⁸ F-FDG PET		P
	Median	IQR	Median	IQR	
Tumor SUV _{max}	23.83	9.09–44.65	15.87	11.70–18.87	0.088
Tumor SUV _{mean}	4.90	3.00–8.16	4.71	3.49–5.62	0.125
Total tumor volume	294	159–558	299	139–410	0.256
Total lesion activity	2,123	995–5,304	1,197	355–2,433	0.009

CONCLUSION

This initial feasibility study demonstrated that anti-CEA immuno-PET using a pretargeted trivalent, BsmAb, and then a ⁶⁸Ga-IMP288 small peptide is a feasible and safe procedure for detecting metastatic lesions in HER2-negative metastatic BC patients. Immuno-PET targeting CEA showed a higher overall sensitivity than ¹⁸F-FDG PET for disclosing metastases, including dissemination in the brain, and thus offers the possibility of an in vivo evaluation of CEA expression and disclosure of whole-body tumor burden, including lesions not accessible to biopsy. Immuno-PET is a potential theranostic molecular imaging technique to select patients for an antibody-based individualized therapy because of metastatic tumor heterogeneity and the discrepancy between serum levels and tissue expression of the antigen in patients.

DISCLOSURE

David Goldenberg and Robert Sharkey own Immunomedics stock or stock options, and Dr. Goldenberg has royalty-bearing patented inventions. David Goldenberg is the founder and retired chief scientific officer of Immunomedics, Inc., and the founder and retired chairman of IBC Pharmaceuticals, Inc. Robert Sharkey is a consultant to Immunomedics, Inc. The proprietary TF2 and IMP288 reagents were provided by Immunomedics, Inc., and IBC Pharmaceuticals, Inc. This work was supported in part by grants from the University Hospital of Nantes and Fondation Avenir in 2010, the French National Agency for Research, called “Investissements d’Avenir” IRON Labex ANR-11-LABX-0018-01 and ArronaxPlus Equipex ANR-11-EQPX-0004 and by a grant, INCa-DGOS-Inserm_12558 (SIRIC ILIAD). This work was presented

in part at the 2014 European Nuclear Medicine Congress in Göteborg, Sweden; at the 2014 San Antonio Breast Cancer Symposium in San Antonio, Texas; and at the 2015 annual meeting of the American Society of Clinical Oncology in Chicago, Illinois. No other potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

We thank the patients and their families for their participation, and Sabina Belcheva and the nuclear medicine technologists at the Institut de Cancérologie de l’Ouest for their contributions.

KEY POINTS

QUESTION: Is anti-CEA pretargeting immuno-PET using the BsmAb TF2 and the ⁶⁸Ga-IMP288 peptide a potential theranostic molecular imaging technique to select HER2-negative BC patients for an antibody-based individualized therapy?

PERTINENT FINDINGS: In a prospective clinical trial comparing the sensitivity of anti-CEA pretargeting immuno-PET with ¹⁸F-FDG PET for 23 HER2-negative metastatic BC patients, we showed a higher overall sensitivity for immuno-PET (94.7%) than for ¹⁸F-FDG PET (89.6%) for disclosing metastases, particularly in the brain. The possibility is thus offered of an in vivo evaluation of CEA tumor expression, including lesions not accessible to biopsy.

IMPLICATIONS FOR PATIENT CARE: Anti-CEA pretargeting immuno-PET appears to be an efficient theranostic molecular imaging technique for an antibody-based individualized therapy to select patients unable to receive trastuzumab.

FIGURE 4. Correlation between SUV_{max} of most intense lesion in immuno-PET and serum CEA level (µg/L).

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