Molecular Imaging of Cardiac Amyloidosis

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Transthyretin and light-chain amyloidosis are the 2 main causes of cardiac amyloidosis. Recent developments in molecular imaging have transformed our ability to diagnose transthyretin cardiac amyloidosis noninvasively and unmasked a hitherto unrecognized prevalence of the disease. This review summarizes the current and evolving imaging approaches, their molecular structural basis, and the gaps in imaging capabilities that have arisen as a result of parallel developments in pharmacotherapy delivering the first effective treatment options for this condition.

Key Words: molecular imaging; SPECT; ATTR; bone-seeking tracers; cardiac amyloidosis

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Cardiac amyloidosis is considered to be a rare disease, defined in the United States as a condition that affects fewer than 200,000 people (1). However, the recent adoption of noninvasive diagnostic approaches has unmasked a hitherto unrecognized community prevalence of wild-type amyloid transthyretin (ATTR) cardiac amyloidosis (CA), reversing prior impressions of its lower prevalence than amyloid light-chain (AL) amyloidosis (40.5 cases per million) (2). Quite serendipitously, parallel developments in pharmacotherapy have resulted in the recent approval by the U.S. Food and Drug Administration of 3 new agents for treating ATTR amyloidosis, with many clinical studies under way of new agents and expanded applications of approved ones. Taken together, these developments in diagnosis and therapy have transformed and invigorated the field of CA, which, until quite recently, relied solely on an invasive myocardial biopsy for diagnosis and faced therapeutic nihilism for ATTR amyloidosis. These developments have also spurred a renewed interest in the imaging of CA, with emphasis on diagnosis, differentiation of amyloid type, and monitoring of therapy.

THE MOLECULAR BASIS OF AMYLOIDOSIS

The term amyloid was coined by Schleiden and used by Virchow in the mid-19th century based on the property of staining with iodine (amyloid = starchlike) (3). However, today we know that amyloid is derived from the misfolding of proteins (not starch) into long, unbranched fibrils of approximately 10 nm in diameter and several micrometers in length with a specific β-sheet structure that is extremely stable and resistant to degradation by cells. Amyloid diseases are generally chronic and occur because of progressive extracellular accumulation of amyloid in tissues, resulting in impaired function of the affected organs.

More than 30 different specific amyloid diseases have been described, with organ tropism and clinical manifestations driven by the protein of origin. Examples include Alzheimer disease, β-2 microglobulin-related amyloidosis in chronic dialysis patients, and systemic amyloid A amyloidosis associated with chronic inflammation. Irrespective of the protein of origin, all amyloid is morphologically indistinguishable and characterized by an antiparallel β-sheet secondary structure (in which the N- and C- terminals are oriented in opposite directions), which gives it its characteristic imaging properties with Congo red (red deposits on light microscopy and apple-green birefringence under polarized-light microscopy) and thioflavin T (yellow-green fluorescence) (2). The heterogeneity of organ involvement and clinical syndromes, despite morphologic homogeneity, is one of the mysteries of this intriguing disease.

The majority (95%) of clinical CA is due to deposition of either transthyretin (TTR) or immunoglobulin AL–derived amyloid, which produce clinical diseases that differ vastly in their phenotypic expression, prognosis, and therapy. Rare cases of CA are derived from other precursor proteins, such as apolipoprotein A1. Amyloid fibril type is most specifically confirmed with immunohistochemistry or mass spectroscopy of biopsy material.

AL amyloidosis is the result of the unregulated proliferation of a single clone of plasma cells (a monoclonal gammopathy) resulting in the overproduction of immunglobulin ALs that transform into amyloid in tissues. Certain isotypes (λ) and variability subgroups (λV1) seem to confer a tendency to amyloidogenesis, since only a small proportion of patients with a plasma cell dyscrasia actually develop clinical amyloidosis (4). AL is a systemic disease with prognosis driven...
largely by the toxic–infiltrative cardiomyopathy (5). Hence, early diagnosis before advanced cardiac involvement that precludes chemotherapy is a key factor determining survival, potentially giving diagnostic imaging a key role in the evaluation of these patients. One of the critical steps in the evaluation (including imaging) of the patient with suspected CA is the identification of the amyloid type, since a mistaken diagnosis of ATTR for AL amyloidosis can have disastrous consequences.

ATTR amyloidosis is derived from TTR, a carrier of thyroxine and retinol bound to retinol-binding protein that is produced largely by the liver and, in smaller amounts, by the retinal epithelium and choroid plexus. The transformation of this 127-amino-acid protein into amyloid is stimulated by either unknown mechanisms related to aging in wild-type ATTR or by at least 120 known point mutations in its gene located on the long arm of chromosome 18, resulting in single-amino-acid substitutions in variant ATTR. Specific mutations result in well-described but not exclusive cardiac, neuropathic, or mixed phenotypes. The most common TTR gene mutation worldwide is V30M. In the United States, the V122I mutation is the most prevalent and is carried by 3%–4% of the African American population (6). In contrast to AL amyloidosis, the cardiac consequences of ATTR appear to result mostly from infiltration of the myocardium, with less evidence of direct toxic effects.

Contemporary clinical data suggest that there is generally a lag period of 2 or more years between the patient’s first presentation to a physician and the diagnosis of CA (7). Recent educational efforts have focused on increasing awareness of red-flag clinical features. In this context, imaging plays a key role in the early and accurate diagnosis of CA.

### ESTABLISHED IMAGING APPROACHES

Contemporary imaging approaches are listed in Table 1 and illustrated in Figure 1. On echocardiography, the initial suspicion of CA is often spurred by the nonspecific finding of increased left ventricular thickness, which can be asymmetric in up to 20% of patients (8). Other findings on echocardiography include left and right atrial enlargement, increased thickness of the right ventricular walls and the interatrial septum, pericardial effusion, diastolic dysfunction, and a longitudinal strain abnormality with relative apical sparing (7). A previously described granular speckled appearance of the myocardium is now thought to be nonspecific for amyloidosis. Diagnostic confirmation of amyloidosis generally requires a tissue biopsy, and echocardiographic findings do not distinguish between ATTR and AL CA. Given the nonspecific nature of echocardiography findings, its diagnostic accuracy even in combination with electrocardiography findings is only 60% (9).

### Cardiac MR

In addition to the morphologic abnormalities seen on echocardiography, a unique abnormality of gadolinium kinetics, seen as an inability to null the myocardium on the inversion time mapping sequence along with early clearance of tracer from the blood pool, is typical. Native T1 time and the extracellular volume fraction are generally higher in amyloidosis than other commonly encountered conditions producing increased left ventricular thickness, and a high extracellular volume is associated with a particularly negative prognosis in amyloidosis (10). Cardiac MR features do not reliably distinguish between ATTR and AL CA, and the possibility of a negative cardiac MR result in biopsy-proven amyloidosis is reported to be up to 12% (9).

### TABLE 1

Radionuclide Tracers for Imaging of Amyloidosis

<table>
<thead>
<tr>
<th>Tracer Molecular target</th>
<th>Disease target</th>
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<tbody>
<tr>
<td><strong>SPECT tracers</strong></td>
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<tr>
<td>Bone-seeking radiotracers</td>
<td>Phosphate binders</td>
</tr>
<tr>
<td>99mTc-pyrophosphate</td>
<td></td>
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<tr>
<td>99mTc-3,3-diphosphono-1,2-propanodicarboxylic acid</td>
<td></td>
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<tr>
<td>99mTc-hydroxymethylene diphosphonate</td>
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<tr>
<td><strong>Other radiotracers</strong></td>
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<tr>
<td>123I-serum amyloid protein</td>
<td>Serum amyloid P</td>
</tr>
<tr>
<td>123I-aprotinin</td>
<td>Serum protease inhibitor, amyloid binder</td>
</tr>
<tr>
<td><strong>PET tracers</strong></td>
<td></td>
</tr>
<tr>
<td>Thioflavin-T derivatives</td>
<td>All amyloid fibrils (β-structure and side chains)</td>
</tr>
<tr>
<td>18F-florbetapir</td>
<td></td>
</tr>
<tr>
<td>18F-florbetaben</td>
<td></td>
</tr>
<tr>
<td>18F-flutemetamol</td>
<td></td>
</tr>
<tr>
<td>11C-Pittsburgh compound B</td>
<td></td>
</tr>
<tr>
<td>18F-sodium fluoride</td>
<td>Microcalcification</td>
</tr>
<tr>
<td>124I-m11-1F4 monoclonal antibody</td>
<td>Immunoglobulin ALs</td>
</tr>
</tbody>
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Radionuclide Scintigraphy with Bone-Seeking Tracers

A resurgence of interest and an improved understanding of the performance characteristics of myocardial scintigraphy with the use of 99mTc-labeled bone-seeking tracers, specifically Tc-pyrophosphate, 3,3-diphosphono-1,2-propanodicarboxylic acid (Tc-DPD), and Tc-hydroxymethylene diphosphonate have been transformative for the field of CA. A recent multinational study of 1,212 patients reported a high sensitivity and near-perfect specificity for ATTR amyloidosis. Notably, hereditary amyloidosis resulting from some mutations result in type B amyloid fibrils, which are less avid for BST. On the other hand, prior reports suggest that some patients with AL may show BST uptake. Differentiation of ATTR from AL amyloidosis is a critical step in evaluation of patients with suspected CA. Wt = wild-type.

99mTc-pyrophosphate uptake in some patients with AL amyloidosis have prompted a strong recommendation for mandatory serum studies (serum-free AL assay and immunofixation electrophoresis) in all patients undergoing scintigraphy (13). A quantitative heart-to-contralateral ratio has been proposed for 99mTc-pyrophosphate, with values greater than 1.5 being highly specific for ATTR (14). Grades ≥ 2 are considered positive. Figure 2 shows a typical example of a positive 99mTc-pyrophosphate scan. Prior reports of

scans, is not useful for amyloidosis imaging because of poor sensitivity. 99mTc-pyrophosphate scintigraphy is a relatively simple test that is widely available and applicable in a broad range of patients, including those with relative contraindications to other imaging modalities, such as atrial fibrillation, implanted cardiac devices, contrast allergy, renal dysfunction, and extreme obesity. The combination of high clinical efficacy (diagnostic accuracy) and clinical effectiveness (applicability) make it highly useful for screening.

Interpretation is based on a semi-quantitative visual grading of tracer uptake, first proposed by Perugini (grade 0 = no myocardial uptake, grade 1 = myocardial < rib uptake, grade 2 = myocardial equal to ribs, and grade 3 = myocardial > rib uptake, usually with diminished bone uptake) (14). Grades ≥ 2 are considered positive. Figure 2 shows a typical example of a positive 99mTc-pyrophosphate scan. Prior reports of

EVOLVING IMAGING APPROACHES

Several PET tracers have been studied for CA, including the thioflavin T analogs 11C-Pittsburgh compound B (18),

<table>
<thead>
<tr>
<th>Protein source</th>
<th>Normal structure</th>
<th>Misfolding</th>
<th>Amyloid formation</th>
<th>Tissue disposition</th>
<th>Imaging scintigraphy</th>
<th>PET imaging</th>
</tr>
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<tbody>
<tr>
<td>ATTR</td>
<td></td>
<td></td>
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<td></td>
<td>Echo</td>
<td>CMR</td>
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<tr>
<td>AL</td>
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18F-florbetapir (19), 18F-florbetaben (20), and 18F-flutemetamol (21). These agents were all developed for imaging cerebral Aβ-amyloid in Alzheimer disease and are taken up by myocardial amyloid. They do not, however, distinguish between ATTR and AL. A major impetus for the development of amyloid PET tracers has been the expectation of more accurate quantification abilities than are possible with SPECT. In preliminary studies, quantitative metrics show higher myocardial uptake in patients with AL or ATTR CA than in controls (19,22), in AL patients than in ATTR patients (23), and in chemotherapy-naïve patients than in postchemotherapy patients (24). 18F-NaF, an agent used to image bone metastases in several types of cancer, targets microcalcifications. Small studies suggest utility for the diagnosis of ATTR CA, with quantitative parameters able to distinguish patients with ATTR CA from AL CA and controls (25). The affinity of 18F-NaF for ATTR CA corroborates the calcium hypothesis to explain the binding of bone-seeking tracers to ATTR. Serum amyloid P component is a circulating plasma protein that has high affinity for amyloid fibrils and is a nonfibrillar component of all types of amyloid. Theoretically, SAP targeted imaging should be specific for tissue amyloid. However, the SPECT tracer 123I-SAP was found to be less useful for cardiac imaging because of high blood-pool activity and poor count statistics in the myocardium (26). Another SPECT tracer, 99mTc-aprotinin, targets this serum protease inhibitor found in amyloid deposits. However, its utility is also limited by poor-quality images (27). Similarly, a murine monoclonal antibody developed against ALs and labeled with iodine was found to have limited use in CA (28).

BINDING MECHANISMS: INSIGHTS FROM STRUCTURE–FUNCTION RELATIONSHIPS

The structure of TTR-derived amyloid fibrils was recently revealed using cryoelectron microscopy (29). Perturbations in the structural conformation of the precursor protein are at the center of amyloidogenesis. Interestingly, at least 2 identified rescue mutations, T119M and R104H, stabilize TTR in patients who are compound heterozygotes for a pathogenic mutation and prevent expression of disease (30). Despite the heterogeneity of the mechanisms leading to amyloid formation (aging, gene mutation, proteolytic remodeling), the amyloid fibril is characterized by relative structural homogeneity. Early studies exploring the binding mechanism of thioflavin T suggested that the surface of the cross β structures forms the binding site, and the orientation of the side chains may be the common pattern that determines the affinity of thioflavin T to a broad range of amyloid fibrils (31). The binding mechanism of thioflavin T is shared by analogous tracers, including 11C-Pittsburgh compound B and the 18F-tracers (Fig. 4). The SPECT and PET tracers bind by different, not yet fully elucidated, mechanisms. It would seem logical to attribute the tropism of the
SPECT bone-seeking tracers to calcium binding; however, there are only sparse clinical data to support this notion. To specifically explore tracer binding of the bone-seeking tracers, Stats et al. performed a unique study of endomyocardial biopsies from 8 ATTR and AL amyloidosis patients (13). Using specific stains and immunohistochemistry, they reported a greater density of microcalcification in biopsies from ATTR patients and a greater density of macrophages in biopsies from AL patients, supporting a calcium-based binding mechanism of bone tracer uptake in ATTR. In this regard, it is relevant that 99mTc-pyrophosphate was introduced as a tracer for the imaging of subacute myocardial infarction, where its uptake correlates with calcium deposits in the infarcted myocardium (32), and that 18F-NaF is a tracer that specifically binds to sites of active microcalcification. These facts further support a calcium hypothesis to explain the binding of these tracers. Of clinical relevance, some of the cases of AL in the study of Stats et al. demonstrated densities of microcalcification comparable to ATTR cases, consistent with the clinical finding of 99mTc-DPD/99mTc-pyrophosphate positivity in some patients with AL amyloidosis.

The mechanistic basis of increased calcification in ATTR is unknown. The density of calcification in the study of Stats et al. was unrelated to age, cardiac function, renal function, or serum calcium. The authors postulate that the denser microcalcification in ATTR may be a reflection of chronicity compared with the typical time course of AL. The finding of a higher density of macrophages in biopsies from patients with localized AL is consistent with prior studies showing 18F-FDG uptake in these patients (33).

Although amyloid fibrils derived from different precursor proteins are mostly structurally homogeneous, recent reports using Western blot analysis have indicated the existence of 2 types of amyloid fibrils derived from TTR. Type A fibrils consist of a mixture of C-terminal fragments (N-terminally truncated) and full-length TTR and are the predominant type in wild-type ATTR and most hATTR. Type B fibrils consist only of full-length TTR and are found in some patients with V30M ATTR and have been reported with the T114C mutation (34). The fibril type in V30M determines the clinical phenotype. Patients with type A are characterized by late-onset (age > 50 y) disease with a mixture of neuropathy and cardiomyopathy. Patients with type B have early-onset disease that has a predominantly neuropathic phenotype. Geographic clustering occurs, with early-onset (third decade) disease prevalent in Portugal, Japan, and Brazil, and a mixture of early- and late-onset disease (fifth to sixth decades) common in Sweden (35). Individual patients with hATTR display only one type of fibril, which does not change over time (36). Type A fibrils are less avid for Congo red than type B fibrils, a fact that may explain the lower yield of subcutaneous-fat-pad biopsies in wild-type ATTR (35).

Pilebro et al. reported that patients with type B fibrils were 99mTc-DPD–negative, despite echocardiographic and, in some patients, cardiac MR evidence of cardiac involvement (37). A low sensitivity for 99mTc-DPD has also been reported for ATTR because of the P64L mutation (38). Thus, calcification alone cannot fully explain the affinity of bone-seeking tracers to ATTR, and direct binding to fibrils cannot be excluded at this time. However, these small studies did not confirm cardiac involvement by biopsy and, therefore, are far from conclusive.

**KNOWLEDGE GAPS AND FUTURE DIRECTIONS**

With the advent of effective treatment options for ATTR amyloidosis, a critical need has emerged for an imaging test to monitor disease activity by serial imaging. All current imaging approaches have limitations in this regard, including scintigraphy with bone-seeking tracers and cardiac MR (39,40). With its superior ability to quantify tracer uptake, PET-based tracers are being actively investigated for serial imaging. An improved understanding of structure–function relationships and binding mechanism will help in efforts to develop new amyloid tracers.

Despite the transformative effects of scintigraphy on our ability to diagnose ATTR CA, the global experience with these tracers for the diagnosis of CA is still relatively small. Its performance characteristics in the community setting (in contrast to specialist centers with highly selected referral populations) remain unexplored. Conditions other than CA resulting in positive scintigraphy are being identified (e.g., hydroxychloroquine cardiotoxicity) (41). Binding mechanisms and the effect of fibril type on tracer affinity need to be clarified. Thus, whereas scintigraphy with the 99mTc bone-seeking tracers is a remarkably useful test for patients with suspected amyloidosis, vigilance is required to identify and further evaluate cases in which there is discordance between clinical and imaging findings.

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REFERENCES


