
Effects of Hypercapnia on Myocardial Blood Flow in Healthy Human Subjects

Matthieu Pelletier-Galarneau¹⁻³, Robert A. deKemp^{2,3}, Chad R.R.N. Hunter^{2,3}, Ran Klein^{2,3}, Michael Klein⁴, Joel Ironstone⁴, Joseph A. Fisher⁵, and Terrence D. Ruddy¹⁻³

¹Division of Nuclear Medicine, The Ottawa Hospital, Ottawa, Ontario, Canada; ²Department of Medicine, University of Ottawa, Ottawa, Ontario, Canada; ³Division of Cardiology, University of Ottawa Heart Institute, Ottawa, Ontario, Canada; ⁴Thornhill Research Inc., Toronto, Ontario, Canada; and ⁵Department of Anesthesiology, University of Toronto, Ottawa, Ontario, Canada

Elevation of the end-tidal partial pressure of CO₂ (P_{ETCO₂}) increases cerebral and myocardial blood flow (MBF), suggesting that it may be a suitable alternative to pharmacologic stress or exercise for myocardial perfusion imaging. The purpose of this study was to document the pharmacodynamics of CO₂ for MBF using prospective end-tidal targeting to precisely control arterial P_{CO₂} and PET to measure the outcome variable, MBF. **Methods:** Ten healthy men underwent serial ⁸²Rb PET/CT imaging. Imaging was performed at rest and during 6-min hypercapnic plateaus (baseline; P_{ETCO₂} at 50, 55, and 60 mm Hg; repeat of P_{ETCO₂} at 60 mm Hg; and repeat of baseline). MBF was measured using ⁸²Rb injected 3 min after the beginning of hypercapnia and a 1-tissue-compartment model with flow-dependent extraction correction. Results were compared with those obtained during an adenosine stress test (140 µg/kg/min). **Results:** Baseline P_{ETCO₂} was 38.9 ± 0.8 (mean ± SD) mm Hg (range, 35–43 mm Hg). All P_{ETCO₂} targets were sustained, with SDs of less than 1.5 mm Hg. Heart rate, systolic blood pressure, rate × pressure product, and respiratory frequency increased with progressive hypercapnia. MBF increased significantly at each level of hypercapnia to 1.92-fold over baseline (0.86 ± 0.24 vs. 0.45 ± 0.08 mL/min/g; *P* = 0.002) at a P_{ETCO₂} of 60 mm Hg. MBF after the administration of adenosine was significantly greater than that with the maximal hypercapnic stimulus (2.00 vs. 0.86 mL/min/g; *P* < 0.0001). **Conclusion:** To our knowledge, this study is the first to assess the response of MBF to different levels of hypercapnia in healthy humans with PET. MBF increased with increasing levels of hypercapnia; MBF at a P_{ETCO₂} of 60 mm Hg was double that at baseline.

Key Words: hypercapnia; myocardial blood flow; adenosine; myocardial perfusion imaging; rubidium; end-tidal partial pressure of CO₂

J Nucl Med 2018; 59:100–106
DOI: 10.2967/jnumed.117.194308

Little is known about the effects of hypercapnia (arterial P_{CO₂} [P_{aCO₂}] of >45 mm Hg) on myocardial blood flow (MBF) in humans. The effects of P_{aCO₂} on MBF were first studied more than 100 y ago in dogs, in which coronary blood flow increased with hypercapnia (1,2). The authors attributed the increased MBF

to smooth muscle relaxation leading to decreased coronary resistance. These results were reproduced using different MBF measurement techniques in different animal models (3–6).

Yang et al. were the first to systematically examine the dose–response characteristics of controlled hypercapnia on absolute MBF in humans using precise CO₂ targeting (7). They studied the effect of a single 10-mm Hg increase in the end-tidal partial pressure of CO₂ (P_{ETCO₂}) on MBF in healthy humans using myocardial blood oxygen level–dependent MRI, a surrogate of MBF (7). They demonstrated that hypercapnia created a hyperemic response similar to that reported for adenosine infusion. Studies in canines demonstrated the dynamic response of MBF over a range of 30–60 mm Hg. More recently, Yang et al. showed no significant difference between the responses to hypercapnia and adenosine infusion in canines without coronary stenosis, with MBF doubling over that at baseline (8).

Other studies investigating the effects of hypercapnia on MBF in humans were performed with dissimilar protocols and produced variable results. None provided the dynamic range of response of MBF to graded P_{ETCO₂}. In this study, we assessed the relationship between graded levels of P_{ETCO₂} and increases in MBF, as measured using PET imaging. The dose–response relationship between P_{ETCO₂} and MBF is a prerequisite for using CO₂ as a noninvasive stimulus in the measurement of myocardial flow reserve (MFR).

MATERIALS AND METHODS

Study Population

Eleven healthy male volunteers, screened by a clinical history and physical examination, were recruited for this study. One recruited subject did not attend on imaging day and was therefore excluded. Participants had no history of obstructive coronary artery disease (CAD) and had no symptoms of CAD or heart failure. Participants provided written informed consent, and the study was approved by the Ottawa Health Science Network Research Ethics Board. Participants were instructed to avoid caffeinated drinks for 24 h before imaging.

PET Imaging

At present, MBF may be assessed noninvasively with several methods. Transthoracic echocardiography can estimate epicardial coronary artery flow reserve, and MRI can approximate global and regional flow reserve. Nevertheless, PET imaging remains the clinical gold standard to which other noninvasive techniques are compared and validated. The accuracy and repeatability of PET have made it the method of choice for quantifying absolute MBF (9).

All participants underwent a baseline rest ⁸²Rb PET scan to quantify MBF and low-dose CT for attenuation correction. After this baseline study, serial ⁸²Rb PET scans using 3 levels of P_{ETCO₂} (50, 55, and 60 mm Hg), a suitable surrogate of P_{aCO₂} (10), were acquired. Using

Received Apr. 4, 2017; revision accepted Jun. 8, 2017.
For correspondence or reprints contact: Terrence D. Ruddy, 40 Ruskin, Room H1220, Ottawa, Ontario, Canada K1Y4W7.
E-mail: truddy@ottawaheart.ca
Published online Jun. 15, 2017.
COPYRIGHT © 2018 by the Society of Nuclear Medicine and Molecular Imaging.

sequential gas delivery, P_{ETCO_2} levels were within ± 1 mm Hg of their arterial equivalents (11). The 60-mm Hg level scan was repeated after a rest period of at least 10 min. A second rest ^{82}Rb PET scan was performed after a return to normocapnia. For every level of P_{ETCO_2} , hypercapnia was maintained for 6 min. ^{82}Rb infusion and PET acquisition were started 3 min after the beginning of hypercapnia. This acquisition was followed by an adenosine stress ^{82}Rb PET scan at least 20 min later (Fig. 1). A total of 7 ^{82}Rb PET scans per participant were performed within 90 min.

Heart rate (HR) and blood pressure were measured 1 to 3 min before stress, immediately before tracer injection, and 3 to 5 min after injection (Spot Vital Signs Device; Welch Allyn). Additionally, during pharmacologic and hypercapnic stresses, HR was measured every minute using 12-lead echocardiography (CardioSoft Diagnostic System; GE). The rate \times pressure product (RPP) calculation was performed using the average HR and systolic blood pressure (SBP) at peak stress and after tracer infusion.

^{82}Rb PET imaging was performed according to the University of Ottawa Heart Institute standard clinical protocol previously described (12). In brief, participants were positioned in a 3-dimensional PET system (GE Discovery 690 PET/CT). The initial rest PET scan included low-dose CT for attenuation correction (fast helical, 1.5 s, 120 kVp with axial and angular mA modulation at a noise index of 50). After ^{82}Rb was administered at 10 MBq/kg intravenously over 30 s, PET imaging was performed over a period of 6 min. MBF was quantified using a 1-tissue-compartment model with flow-dependent extraction correction (12). Polar maps representing MBF were generated for each rest and stress state using validated software (FlowQuant; University of Ottawa Heart Institute) (13). Corrected MBF values were calculated using the population average rest RPP according to the following formula: corrected MBF = (MBF \times average RPP)/subject RPP. MFR were calculated by dividing MBF values by rest-MBF. All presented MBF and MFR were unadjusted except when noted otherwise.

Hypercapnia

Until recently, precise modulation of P_{aCO_2} and arterial oxygen tension (P_{aO_2}) levels could be achieved only by changing the concentrations of inspired gases. These fixed inspired gas methods are affected by variations in breathing patterns (frequency and tidal volume), leading to variability in end-tidal gas concentrations (14). A new approach of breath-by-breath control of arterial blood gases using sequential gas delivery (10,11,15) enabled the automated prospective control of arterial blood gases (16) independently of the subject's ventilation or pattern of breathing. Using sequential gas delivery, it is possible to maintain a euoxic-hypercapnic state at any level for several minutes with minimal discomfort for the subject (17)—conditions needed for the precise assessment of MBF using PET imaging.

P_{ETCO_2} and end-tidal O_2 partial pressure targets were achieved using prospective end-tidal gas targeting with previously described algorithms (11). These algorithms were applied by a computerized gas blender (RespirAct; Thornhill Research) that had PCO_2 and PO_2 sensors and that was connected to a sequential gas delivery breathing circuit (15). Pilot studies with the computerized gas blender showed that a PCO_2 of 60 mm Hg could be tolerated consistently for several minutes. Therefore, this level was chosen as the upper PCO_2 target for the study, so that a dose-response relationship could be discerned and provide an indication of whether the response of MBF to PCO_2 plateaued as PCO_2 approached 60 mm Hg.

For this study, 3 levels of P_{ETCO_2} (50, 55, and 60 mm Hg) were targeted and maintained over 6 min while euoxia was maintained (end-tidal O_2 partial pressure of 100 mm Hg). Tidal PCO_2 and PO_2 were monitored continuously and recorded. Before imaging, the participants' PCO_2 was increased to 60 mm Hg for 6 min to familiarize them with the sensation of hypercapnia and confirm their tolerance for the change. Baseline P_{ETCO_2} levels were estimated using average P_{ETCO_2} measurements after initial application of the mask and before hypercapnia. Baseline respiratory frequencies (f) were calculated after the participant was positioned under the camera while wearing the mask.

Study Tolerability

Subjects graded the discomfort associated with hypercapnia and adenosine on a subjective scale from 0 to 10, with 0 representing no discomfort and 10 representing intolerable discomfort.

Statistical Analysis

One-way repeated-measures ANOVAs were conducted before comparison of the different imaging states using the first baseline study and the first maximal hypercapnic stimulus (60 mm Hg). When data for a participant were missing, the entire dataset for that participant was excluded from repeated-measures ANOVAs. Post hoc analyses were conducted using paired Student t tests with Bonferroni-Holm sequential corrections for multiple comparisons with the dataset for all patients. A total of 8 comparisons were performed for the variables of interest (baseline vs. 50 mm Hg, baseline vs. 60 mm Hg, 50 mm Hg vs. 55 mm Hg, 55 mm Hg vs. 60 mm Hg, 60 mm Hg vs. repeat 60 mm Hg, 60 mm Hg vs. adenosine, baseline vs. repeat baseline, and repeat baseline vs. adenosine). Tolerability levels were compared using paired t tests. The average MFR was computed from the individual MFRs measured in each participant.

Unless otherwise specified, the results are presented as mean \pm SD. P values of less than 0.05 were considered statistically significant.

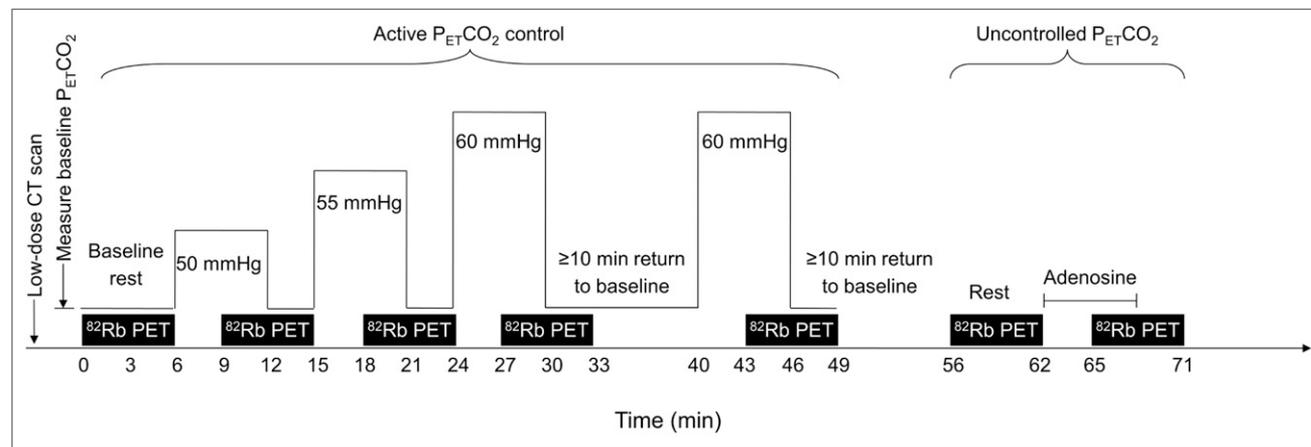


FIGURE 1. Sequence of imaging during single visit.

TABLE 1
Summary of Results

Parameter	Baseline (n = 10)	Repeat baseline (n = 10)	50 mm Hg (n = 10)	55 mm Hg (n = 9)	60 mm Hg (n = 9)	Repeat 60 mm Hg (n = 8)	Adenosine (n = 10)	P value determined by ANOVA*
P _{ETCO₂} (mm Hg)	38.9 ± 2.4	N/A	49.9 ± 0.1	54.8 ± 0.3	59.4 ± 1.2	59.7 ± 0.5	N/A	<0.0001
HR (beats/min)	57 ± 8	59 ± 8	62 ± 7	66 ± 9	68 ± 10	69 ± 8	76 ± 10	<0.001
f (breaths/min)	12 ± 4	N/A	17 ± 5	19 ± 4	22 ± 5	23 ± 6	N/A	<0.001
SBP (mm Hg)	109 ± 13	117 ± 13	119 ± 15	123 ± 11	131 ± 12	125 ± 8	121 ± 13	<0.001
DBP (mm Hg)	65 ± 11	71 ± 11	68 ± 12	70 ± 10	74 ± 13	71 ± 7	70 ± 12	0.196
RPP (bpm × mm Hg)	6,135 ± 1,650	6,900 ± 1,165	7,140 ± 1,495	7,487 ± 1,503	8,256 ± 1,493	7,666 ± 1,240	9,123 ± 1,632	0.001
MBF (mL/min/g)	0.45 ± 0.08	0.52 ± 0.10	0.62 ± 0.20	0.76 ± 0.21	0.86 ± 0.24	0.89 ± 0.24	2.00 ± 0.34	<0.0001
Corrected MBF (mL/min/g)	0.44 ± 0.045	0.47 ± 0.09	0.51 ± 0.15	0.57 ± 0.08	0.59 ± 0.15	0.64 ± 0.19	1.37 ± 0.31	<0.0001
MFR			1.37 ± 0.317	1.68 ± 0.33	1.97 ± 0.45	2.12 ± 0.48	4.53 ± 0.70	<0.0001

*Three participants for whom data were missing were excluded from repeated-measures ANOVA.

n = number of participants; N/A = not available.

Values (other than P values) are presented as mean ± SD.

Statistical analysis was performed using GraphPad Prism version 6.01 for Windows (GraphPad Software) and MedCalc for Windows, version 12.2 (MedCalc Software).

RESULTS

Population

The participants' age, weight, height, and body mass index were 30 ± 12 y, 84.8 ± 11.2 kg, 178 ± 8 cm, and 26.9 ± 4.3 kg/m², respectively. All participants had normal 12-lead echocardiography results before entering the study. The measured baseline P_{ETCO₂} was 38.9 ± 0.8 mm Hg (range, 35–43 mm Hg). For 1 participant, imaging at the 55-mm Hg level was not acquired for technical reasons; imaging at the 60-mm Hg level was not acquired for another participant because of an inability to maintain a stable P_{ETCO₂}. Data from these 2 participants were excluded from the repeated-measures ANOVAs. For 2 other participants, repeat 60-mm Hg acquisitions were not obtained.

Hypercapnic Stimulus

All P_{ETCO₂} targets were attained, with SDs of less than 1.5 mm Hg (Table 1). A representative P_{ETCO₂} tracing presented in Figure 2 shows a rapid increase in the P_{ETCO₂} to the target level and then a stable P_{ETCO₂} at all target levels.

Hemodynamic Parameters

HR, SBP, f, and RPP increased with progressive levels of hypercapnia (Table 1; Fig. 3). There was a moderate correlation between RPP and MBF (R² = 0.41; P < 0.0001) (Fig. 4A). Diastolic blood pressure (DBP) did not change (P = 0.2, as determined by ANOVA).

MBF and MFR

MBF increased 92% with the maximal hypercapnic stimulus (0.86 ± 0.24 vs. 0.45 ± 0.08 mL/min/g; P = 0.002) (Fig. 5). Increases in P_{ETCO₂} and MFR were significantly correlated (R² = 0.43; P < 0.0001) (Fig. 4B). The dose–response relationship between MBF and P_{ETCO₂} levels is shown in Figure 4C. MBF was significantly greater at 50 mm Hg than at baseline (P = 0.030),

at 55 mm Hg than at 50 mm Hg (P = 0.014), at 60 mm Hg than at 55 mm Hg (P = 0.011), and at 60 mm Hg than at baseline (P = 0.002). There was a similar dose–response relationship for corrected MBF (Fig. 4C). Corrected MBF values were significantly lower than noncorrected values at 50 mm Hg (P = 0.032), 55 mm Hg (P = 0.0174), and 60 mm Hg (P = 0.002). MFR was significantly higher at 55 mm Hg than at 50 mm Hg (1.70 vs. 1.38; P = 0.005) and significantly higher at 60 mm Hg than at 55 mm Hg (1.91 vs. 1.70; P = 0.005).

Multiple regression analyses were conducted to examine the relationship between MBF and 2 potential predictors, P_{ETCO₂} and RPP. The multiple regression model with 2 predictor variables produced an R² of 0.55, an F_{2,35} of 21.23, and a P value of less than 0.001. Both P_{ETCO₂} (β = 0.01285; P = 0.0022) and RPP (β = 0.00006; P = 0.0014) had significant positive regression weights.

Adenosine

Adenosine increased MBF in all subjects, to 2.00 ± 0.34 mL/min/g, resulting in an MFR of 4.53 ± 0.70 (range, 3.56–5.73). The adenosine-induced increase in MBF was significantly greater than that elicited by the maximal hypercapnic stimulus of 60 mm Hg (2.00 vs. 0.86 mL/min/g; P < 0.0001).

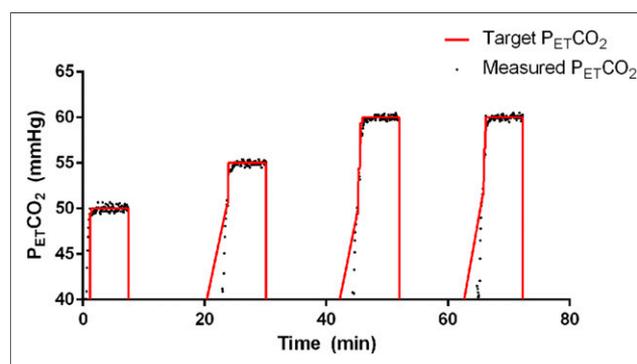


FIGURE 2. Representative curve of target P_{ETCO₂} (red line) and measured P_{ETCO₂} (black dots) in participant at different levels of hypercapnia.

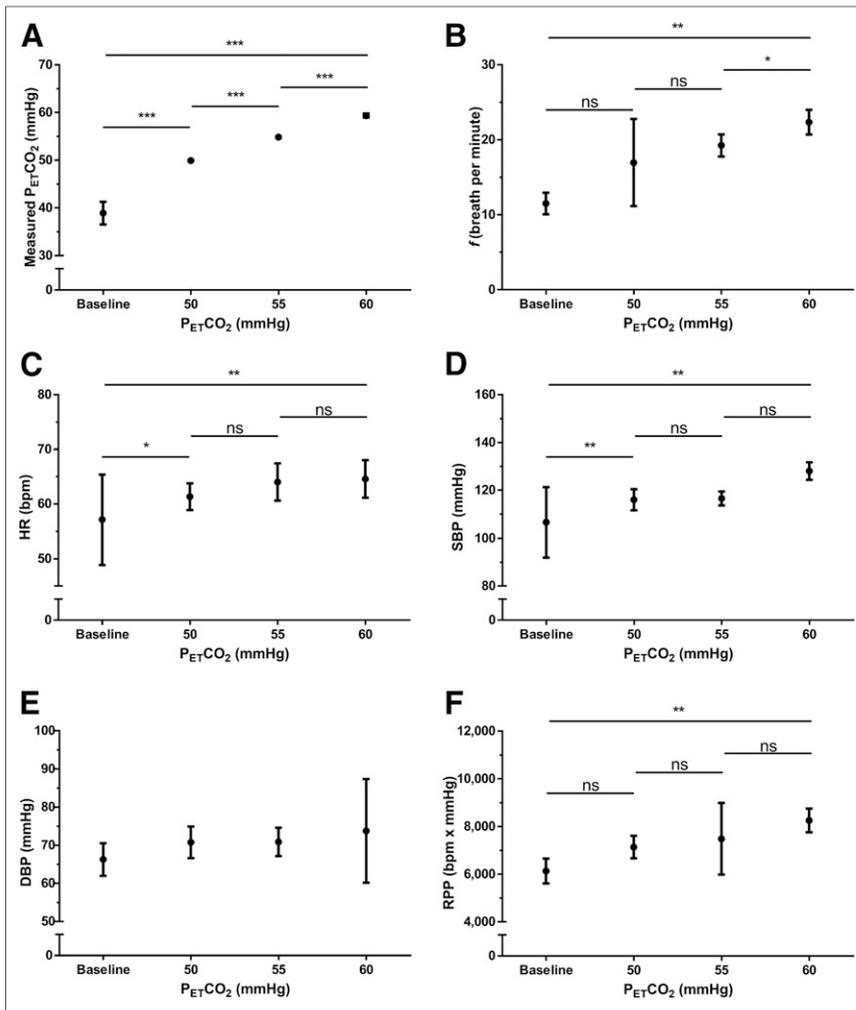


FIGURE 3. Measured P_{ETCO_2} (A), f (B), HR (C), SBP (D), DBP (E), and RPP (F) of all participants at baseline and at different levels of hypercapnia. Results are presented as mean \pm SD. P values of ≥ 0.05 were not significant (ns). * $P < 0.05$. ** $P < 0.01$. *** $P < 0.0001$.

Reproducibility

The interval between the first and second 60-mm Hg acquisitions averaged 24.8 ± 8.3 min (range, 12–41 min). MBF at the repeat 60-mm Hg stimulus was not significantly different from at the initial 60-mm Hg one (0.87 vs. 0.86 mL/min/g; $P = 0.051$) (Fig. 6A). This finding remained true when correction for RPP was used (0.64 vs. 0.59 mL/min/g; $P = 0.434$). Finally, MFR was, on average, marginally greater at the repeat 60-mm Hg level than at the initial 60-mm Hg stress (2.12 vs. 1.97; $P = 0.019$) (Fig. 6B).

Study Tolerability

For 7 subjects, hypercapnia was more tolerable than adenosine, whereas for 2 subjects, adenosine was more tolerable than hypercapnia. In 1 subject, both stimuli were equally tolerable. Average tolerability scores for hypercapnic and adenosine stimuli did not significantly differ (4.5 ± 2.0 vs. 6.0 ± 2.5 ; $P = 0.15$).

DISCUSSION

To our knowledge, this study in healthy young men is the first documentation of a dose–response curve for CO_2 and MBF with

PET. MBF increased progressively with hypercapnia, reaching a doubling of the baseline MBF at a P_{ETCO_2} of 60 mm Hg. Although some animal studies showed no change in MBF with hypercapnia (18,19), our results are consistent with those of other animal studies reporting an increase in MBF with hypercapnia (7,8,20,21). Beaudin et al. (22), using cardiac MRI, demonstrated an increase of 34% in coronary sinus flow with a $Paco_2$ of 45 mm Hg. Tzou et al. (23) demonstrated that hypercapnia increased by approximately 40% the coronary blood flow velocity of the left anterior descending coronary artery, as measured with transthoracic Doppler echocardiography. Furthermore, this increase in blood flow velocity was greater at higher levels of inspired CO_2 . Similar results were obtained with invasive measurements of coronary sinus flow (24).

However, other studies with Doppler echocardiography and PET showed little or no increase in MBF with hypercapnia (25,26). In these studies, the application of fixed inspired CO_2 concentrations elicited unreliable increases in $Paco_2$ (14); indeed, when measured, $Paco_2$ had barely changed, if at all (25). However, these studies in humans were hampered by the inability to attain and maintain the specific Pco_2 levels needed to generate an MBF– Pco_2 relationship or to separate the direct effects of CO_2 from the secondary effects of increases in myocardial work due to hypercapnia-induced increases in HR and SBP (22,24,26). In our young subjects, these secondary effects were mild but contributed to the increase in MBF (Fig. 4).

We also found that hypercapnia elicited reproducible changes in MBF with a repeat P_{ETCO_2} stimulus of 60 mm Hg. In contrast, the pharmacokinetics of intravenous agents resulted in unpredictable blood levels and time courses of the effects (see Fierstra et al. (10) for a discussion). Hypercapnia in the range of 60 mm Hg is high but not outside the range commonly encountered in adults in the community. For example, in people with obstructive sleep apnea, such levels occur as often as 5–20 times/h all night (27). Hypercapnia does not cause irreversible adverse effects, even in patients with underlying morbidities (28–30). A P_{ETCO_2} of 60 mm Hg was well tolerated by all subjects in the present study. As such, hypercapnia may be considered a candidate in the search for an MBF stimulus that is noninvasive, tolerable, safe, rapidly reversible and, most importantly, reproducible—in terms of both stimulus and response.

An increase in RPP, a proxy of cardiac work, with hypercapnia is associated with increased myocardial work and oxygen consumption (31), leading to increased MBF. In the present study, hypercapnia increased both HR and SBP, consistent with previous results in humans (32,33). After correction for RPP, there was still a significant increase in MBF. This result was confirmed by multiple variable regression analyses demonstrating independent

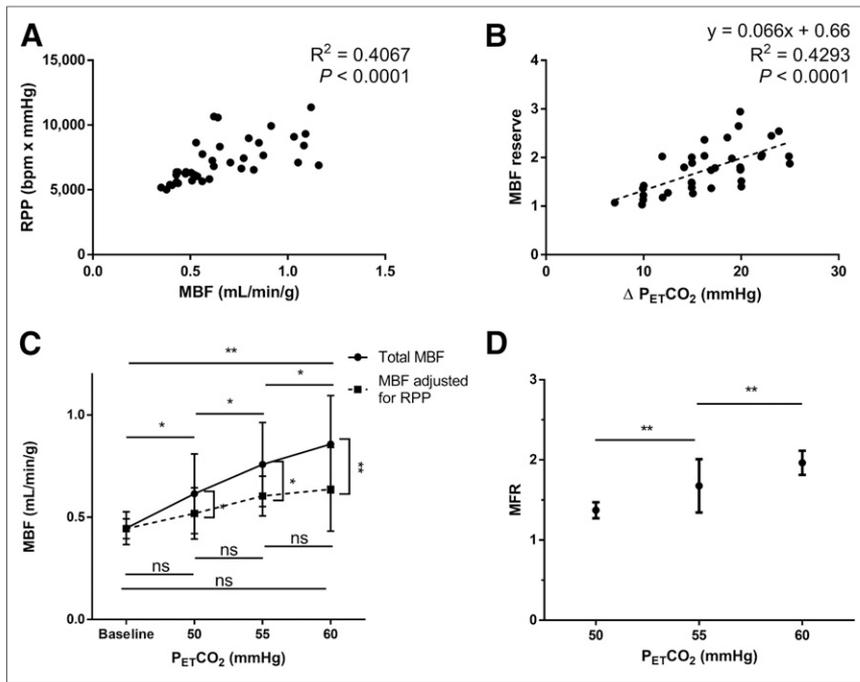


FIGURE 4. (A) MBF vs. RPP. (B) MFR vs. increase in P_{ETCO_2} . (C) Relationships between MBF and P_{ETCO_2} and between adjusted MBF and P_{ETCO_2} . (D) Relationship between MFR and hypercapnia. Results are presented as mean \pm SD. * $P < 0.05$. ** $P < 0.01$.

contributions of RPP and P_{ETCO_2} . These data suggest that the increase in MBF can be partially, but not completely, attributed to increased cardiac work. If the increase in MBF due to increased myocardial work is assumed to be proportional to the increase in RPP, then approximately half of the increase in MBF was due to increased cardiac work. The other half likely was related to the direct vasodilatation effect of hypercapnia. Indeed, prior studies demonstrated that hypercapnia dilates the coronary arteries, a phenomenon mediated by endothelium-derived nitric oxide, adrenergic stimulus, and other mechanisms yet to be determined (3,34–36). At similar P_{ETCO_2} levels in healthy young subjects, Claassen et al. (37) observed a 23% increase in the mean arterial pressure with an SBP of up to 160 mm Hg, which would have increased RPP considerably more than the conditions in the present study and which would have provided greater stress. Like them, we found

hypertensive responses plateauing as P_{ETCO_2} approached 60 mm Hg.

In all subjects, adenosine injection increased MBF more than the maximal hypercapnic stimulus, with an average MFR of 4.53 for adenosine stress. On the one hand, this finding is consistent with previously published results reporting an MFR of 4 to 5 after adenosine stress and shows that our subjects had normal or above-normal coronary flow reserve (38). On the other hand, this result differs from the recently published observations of Yang et al. (8), who evaluated changes in MBF with hypercapnia (P_{ACO_2} increased by 25 mm Hg) and adenosine stresses in canines without coronary stenosis, with non-flow-limiting coronary stenosis, and after caffeine administration. In canines without coronary stenosis, increases in MBF did not differ in response to hypercapnia and adenosine, doubling relative to the baseline level (8). Similar results were observed using myocardial blood oxygen level-dependent MRI (7).

We observed similar MFR levels with hypercapnia, but MFR was greater with adenosine. These findings could have been

related to the fact that the effects of adenosine on MBF are species-dependent and a standard dose of adenosine (140 μ g/kg/min) is insufficient to produce maximal hyperemia in canines (39). Nevertheless, the increases in MBF with hypercapnia were similar in magnitude to previously reported increases during exercise stress (1.8–2.0) (38,40) and with dipyridamole (41,42). Our subject population overlaps the coronary stress test population, some of whom also are young and have normal coronary anatomy. A definitive answer as to what extent hypercapnia can substitute for exercise and dipyridamole would require head-to-head comparisons of exercise versus hypercapnia and dipyridamole versus hypercapnia in a coronary stress test population.

Finally, we showed that the effect of hypercapnia on MBF was reproducible; MBF and corrected MBF with the first and repeat 60-mm Hg stimuli did not differ. The finding that MFR was 8%

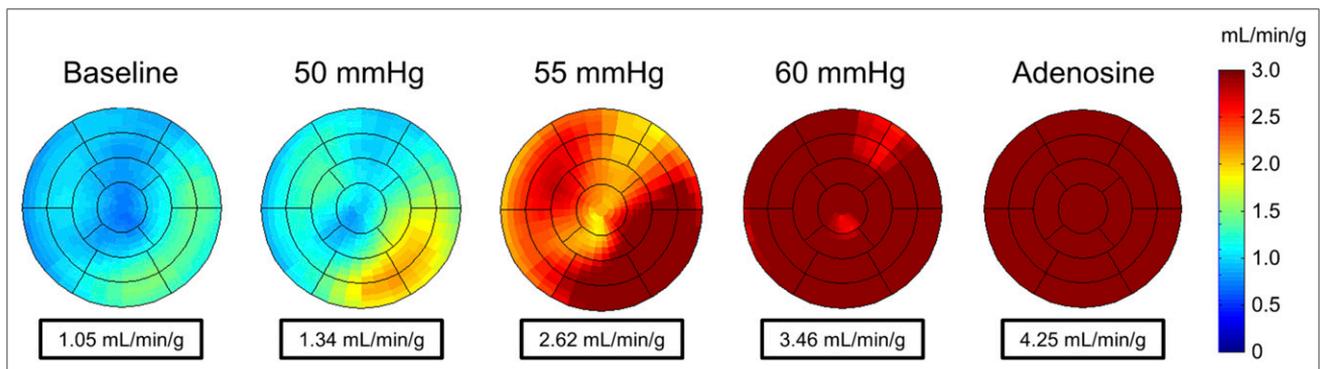


FIGURE 5. MBF polar maps of representative subject, showing global MBF at baseline, during hypercapnic stimuli, and during adenosine stress.

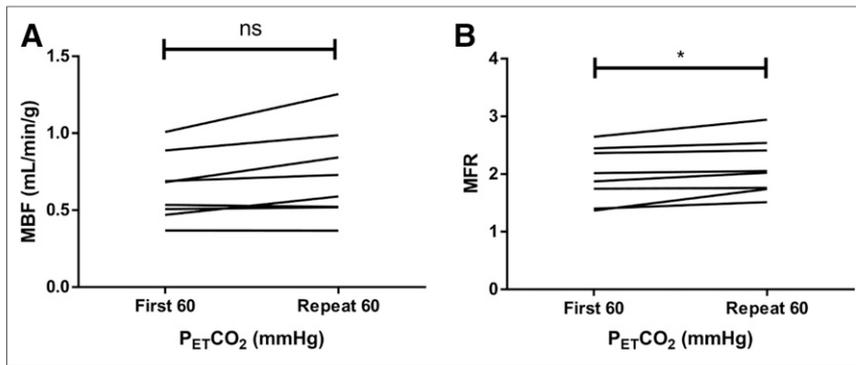


FIGURE 6. Average MBF (A) and MFR (B) during initial and repeat 60-mm Hg P_{ETCO_2} stress. Results are presented as mean \pm SD. P values of ≥ 0.05 were not significant. * $P < 0.05$.

greater on repeat measurements may have been due to a combination of test-retest variability and the effects of the intervening rest period of 12–41 min with free breathing of room air, whereas the initial 60-mm Hg acquisition was performed soon after the 50- and 55-mm Hg acquisitions. Indeed, this finding may have been partly attributable to the fact that the MBF response to an increase in P_{aCO_2} decreased during prolonged (> 10 min) hypercapnia (3).

The main limitation of this study pertained to the population studied. Our subjects were all healthy male volunteers with low resting MBF (female subjects were not purposely excluded; all of the volunteers were male). Whether similar results would be obtained with different populations, such as patients with CAD and subjects with high resting MBF, remains to be determined.

CONCLUSION

To our knowledge, this study is the first to assess changes in MBF in response to different levels of hypercapnia in humans with PET. MBF increased at each incremental P_{ETCO_2} level up to 60 mm Hg. Approximately half of this increase could be attributed to increased cardiac work, as indicated by the RPP. With the maximal hypercapnic stimulus of 60 mm Hg, MBF doubled—comparable to the increase in MBF obtained with exercise stress. Larger increases in RPP due to hypercapnia may increase MBF further.

Hypercapnia has additional characteristics to commend it for clinical investigation: it is readily available, inexpensive, and an intrinsic molecule that can be precisely, consistently, and reproducibly administered by automated delivery systems. It is well tolerated at the effective partial pressure of 60 mm Hg, and it is safe even at multiple times its effective dose, giving it a large safety margin as a drug. The rapid onset and offset of P_{CO_2} levels and the consistency of MBF with repeated hypercapnia tests commend it for repeated tests in a single session, as is sometimes required in clinical practice. To assess the potential clinical impact of hypercapnia in the diagnosis of CAD, further investigations comparing MBF with hypercapnia versus pharmacologic stress or exercise in patients with CAD are warranted.

DISCLOSURE

No potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

We thank Farrah Ahmed, Brian Marvin, and Marlie Poirier of the University of Ottawa Heart Institute, Ottawa, Ontario, Canada, for their contribution in the collection of data. We thank Steve Iscoe for editorial comments.

REFERENCES

- Barcroft J, Dixon WE. The gaseous metabolism of the mammalian heart: part I. *J Physiol (Lond)*. 1907;35:182–204.
- Markwalder J, Starling EH. A note on some factors which determine the blood-flow through the coronary circulation. *J Physiol (Lond)*. 1913;47:275–285.
- Ledingham IM, McBride TI, Parratt JR, Vance JP. The effect of hypercapnia on myocardial blood flow and metabolism. *J Physiol (Lond)*. 1970;210:87–105.
- Alexander CS, Liu SM. Effect of hypercapnia and hypocapnia on myocardial blood flow and performance in anesthetized dogs. *Cardiovasc Res*. 1976;10:341–348.
- Hoffman WE, Albrecht RF, Ripper R, Jonjev ZS. Brain compared to heart tissue oxygen pressure during changes in arterial carbon dioxide in the dog. *J Neurosurg Anesthesiol*. 2001;13:303–309.
- Feigl EO. Coronary physiology. *Physiol Rev*. 1983;63:1–205.
- Yang HJ, Yumul R, Tang R, et al. Assessment of myocardial reactivity to controlled hypercapnia with free-breathing T2-prepared cardiac blood oxygen level-dependent MR imaging. *Radiology*. 2014;272:397–406.
- Yang HJ, Dey D, Sykes J, et al. Arterial CO_2 as a potent coronary vasodilator: a preclinical PET/MR validation study with implications for cardiac stress testing. *J Nucl Med*. 2017;58:953–960.
- Schelbert HR. Positron emission tomography measurements of myocardial blood flow: assessing coronary circulatory function and clinical implications. *Heart*. 2012;98:592–600.
- Fierstra J, Sobczyk O, Battisti-Charbonney A, et al. Measuring cerebrovascular reactivity: what stimulus to use? *J Physiol (Lond)*. 2013;591:5809–5821.
- Slessarev M, Han J, Mardimae A, et al. Prospective targeting and control of end-tidal CO_2 and O_2 concentrations. *J Physiol (Lond)*. 2007;581:1207–1219.
- Efseaff M, Klein R, Ziadi MC, Beanlands RS, deKemp RA. Short-term repeatability of resting myocardial blood flow measurements using rubidium-82 PET imaging. *J Nucl Cardiol*. 2012;19:997–1006.
- Dekemp RA, Declerck J, Klein R, et al. Multisoftware reproducibility study of stress and rest myocardial blood flow assessed with 3D dynamic PET/CT and a 1-tissue-compartment model of ^{82}Rb kinetics. *J Nucl Med*. 2013;54:571–577.
- Fisher JA. The CO_2 stimulus for cerebrovascular reactivity: fixing inspired concentrations vs. targeting end-tidal partial pressures. *J Cereb Blood Flow Metab*. 2016;36:1004–1011.
- Fisher JA, Iscoe S, Duffin J. Sequential gas delivery provides precise control of alveolar gas exchange. *Respir Physiol Neurobiol*. 2016;225:60–69.
- Ito S, Mardimae A, Han J, et al. Non-invasive prospective targeting of arterial $P(CO_2)$ in subjects at rest. *J Physiol (Lond)*. 2008;586:3675–3682.
- Spano VR, Mandell DM, Poubanc J, et al. CO_2 blood oxygen level-dependent MR mapping of cerebrovascular reserve in a clinical population: safety, tolerability, and technical feasibility. *Radiology*. 2013;266:592–598.
- Powers ER, Bannerman KS, Fitz-James I, Cannon PJ. Effect of elevations of coronary artery partial pressure of carbon dioxide (P_{CO_2}) on coronary blood flow. *J Am Coll Cardiol*. 1986;8:1175–1181.
- Case RB, Greenberg H. The response of canine coronary vascular resistance to local alterations in coronary arterial P_{CO_2} . *Circ Res*. 1976;39:558–566.
- Love WD, Tyler MD, Abraham RE, Munford RS. Effects of O_2 , CO_2 , and drugs on estimating coronary blood flow from RB86 clearance. *Am J Physiol*. 1965;208:1206–1210.
- Feinberg H, Gerola A, Katz LN. Effect of changes in blood CO_2 level on coronary flow and myocardial O_2 consumption. *Am J Physiol*. 1960;199:349–354.
- Beaudin AE, Brugniaux JV, Vöhringer M, et al. Cerebral and myocardial blood flow responses to hypercapnia and hypoxia in humans. *Am J Physiol Heart Circ Physiol*. 2011;301:H1678–H1686.

23. Tzou WS, Korcarz CE, Aeschlimann SE, Morgan BJ, Skatrud JB, Stein JH. Coronary flow velocity changes in response to hypercapnia: assessment by transthoracic Doppler echocardiography. *J Am Soc Echocardiogr.* 2007;20:421–426.
24. Kazmaier S, Weyland A, Buhre W, et al. Effects of respiratory alkalosis and acidosis on myocardial blood flow and metabolism in patients with coronary artery disease. *Anesthesiology.* 1998;89:831–837.
25. Momen A, Mascarenhas V, Gahremanpour A, et al. Coronary blood flow responses to physiological stress in humans. *Am J Physiol Heart Circ Physiol.* 2009;296:H854–H861.
26. Yokoyama I, Inoue Y, Kinoshita T, Itoh H, Kanno I, Iida H. Heart and brain circulation and CO₂ in healthy men. *Acta Physiol (Oxf).* 2008;193:303–308.
27. Chin K, Hirai M, Kuriyama T, et al. Changes in the arterial Pco₂ during a single night's sleep in patients with obstructive sleep apnea. *Intern Med.* 1997;36:454–460.
28. Feihl F, Perret C. Permissive hypercapnia: how permissive should we be? *Am J Respir Crit Care Med.* 1994;150:1722–1737.
29. Potkin RT, Swenson ER. Resuscitation from severe acute hypercapnia: determinants of tolerance and survival. *Chest.* 1992;102:1742–1745.
30. Ayas N, Bergstrom LR, Schwab TR, Narr BJ. Unrecognized severe postoperative hypercapnia: a case of apneic oxygenation. *Mayo Clin Proc.* 1998;73:51–54.
31. Gobel FL, Norstrom LA, Nelson RR, Jorgensen CR, Wang Y. The rate-pressure product as an index of myocardial oxygen consumption during exercise in patients with angina pectoris. *Circulation.* 1978;57:549–556.
32. Kiely DG, Cargill RI, Lipworth BJ. Effects of hypercapnia on hemodynamic, inotropic, lusitropic, and electrophysiologic indices in humans. *Chest.* 1996;109:1215–1221.
33. Battisti-Charbonney A, Fisher JA, Duffin J. Respiratory, cerebrovascular and cardiovascular responses to isocapnic hypoxia. *Respir Physiol Neurobiol.* 2011;179:259–268.
34. Weinberger SE, Schwartzstein RM, Weiss JW. Hypercapnia. *N Engl J Med.* 1989;321:1223–1231.
35. Fierstra J, Maclean DB, Fisher JA, et al. Surgical revascularization reverses cerebral cortical thinning in patients with severe cerebrovascular steno-occlusive disease. *Stroke.* 2011;42:1631–1637.
36. Guensch DP, Fischer K, Flewitt JA, et al. Breathing manoeuvre-dependent changes in myocardial oxygenation in healthy humans. *Eur Heart J Cardiovasc Imaging.* 2014;15:409–414.
37. Claassen JAHR, Zhang R, Fu Q, Witkowski S, Levine BD. Transcranial Doppler estimation of cerebral blood flow and cerebrovascular conductance during modified rebreathing. *J Appl Physiol.* 2007;102:870–877.
38. Wyss CA, Koepfli P, Mikolajczyk K, Burger C, von Schulthess GK, Kaufmann PA. Bicycle exercise stress in PET for assessment of coronary flow reserve: repeatability and comparison with adenosine stress. *J Nucl Med.* 2003;44:146–154.
39. Jeremias A, Filardo SD, Whitbourn RJ, et al. Effects of intravenous and intracoronary adenosine 5'-triphosphate as compared with adenosine on coronary flow and pressure dynamics. *Circulation.* 2000;101:318–323.
40. Krivokapich J, Smith GT, Huang SC, et al. ¹³N ammonia myocardial imaging at rest and with exercise in normal volunteers: quantification of absolute myocardial perfusion with dynamic positron emission tomography. *Circulation.* 1989;80:1328–1337.
41. Renaud JM, DaSilva JN, Beanlands RS, DeKemp RA. Characterizing the normal range of myocardial blood flow with ⁸²rubidium and ¹³N-ammonia PET imaging. *J Nucl Cardiol.* 2013;20:578–591.
42. Ziadi MC, Dekemp RA, Williams KA, et al. Impaired myocardial flow reserve on rubidium-82 positron emission tomography imaging predicts adverse outcomes in patients assessed for myocardial ischemia. *J Am Coll Cardiol.* 2011;58:740–748.