

# Somatostatin Receptor 2–Targeting Compounds

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The molecular imaging and treatment of neuroendocrine tumors (NETs) with radiolabeled somatostatin analogs represent a milestone in the development of theranostic compounds. Whole-body scintigraphy with <sup>111</sup>In-pentetreotide has revolutionized the diagnosis and staging of NETs and the evaluation of treatment outcomes. At present, diagnostic accuracy with positron-emitting radionuclides is greater than 90%. Peptide receptor radionuclide therapy (PRRT) has become a well-accepted treatment for patients with well-differentiated inoperable or metastatic NETs and disease progression after first-line treatment. Disease control rates (complete or partial remission or stable disease in patients with formerly progressive disease) of up to 95%, with a low incidence of long-term hematologic and renal toxicity, have been reported. In a recently published randomized trial, compared with intensified treatment of midgut NETs with long-acting and repeatable octreotide, PRRT reduced the hazard of disease progression and death by 79%. Upcoming developments in PRRT include the use of somatostatin receptor antagonists and  $\alpha$ -emitting radionuclides, which may further enhance treatment outcomes.

**Key Words:** theranostics; somatostatin receptor; peptide receptor radionuclide therapy; neuroendocrine tumors

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**T**he field of theranostics, comprising the combination of molecular imaging and molecular radiotherapy, exploits the receptor binding and internalization of the same peptides for imaging and therapy. In this field, the development of somatostatin receptor (SSTR)–targeting techniques represents a milestone. The somatostatin analog (SSA) octreotide was developed in the 1980s for its antiproliferative and hormone release–inhibiting effects in neuroendocrine tumors (NETs), such as pituitary tumors or the so-called carcinoids (1,2). Originally used to target a variety of neoplasms, SSTR–targeting theranostic compounds presently are widely used to image and treat well-differentiated gastroenteropancreatic and bronchopulmonary NETs.

## RECEPTOR TARGETING

### SSTR Expression

In humans, at least 5 subtypes of SSTRs can be found; subtype 2 (SSTR-2) is the predominant receptor in most NETs (3). In the

early era of SSTR scintigraphy, it was used for the detection of both primary tumors and previously undetected metastases of NETs and proved valuable in visualizing several other tumors as well as granulomatous diseases (4). The initial targets of <sup>111</sup>In-pentetreotide scintigraphy (4) were NETs (carcinoids, endocrine pancreatic tumors, pituitary tumors, paragangliomas, pheochromocytomas, medullary thyroid carcinomas, neuroblastomas, and small cell lung cancer) and other tumors and diseases (meningiomas, breast cancer, Merkel cell tumors, lymphomas, sarcoidosis, and tuberculosis).

There is a distinct correlation between tumor grade according to 2010 World Health Organization criteria and SSTR-2 expression (5). High levels of SSTRs are predominantly found in grade 1 and 2, well-differentiated and moderately differentiated NETs; with the loss of differentiation, the characteristic of SSTR expression is frequently lost, too. Hence, SSTR targeting for imaging or therapy is generally limited to lower-grade disease.

### Development of Targeted Compounds

In 1987, the first *in vivo* SSTR scintigraphy in humans was performed using <sup>123</sup>I-Tyr<sup>3</sup>-octreotide. Cumbersome and expensive, the radioiodine was replaced with chelated <sup>111</sup>In in 1989. This radiopharmaceutical was used for imaging and, starting in 1992, for therapy, making it the first SSTR–targeting theranostic compound (6). With the development of the chelator DOTA, SSAs could be stably linked to  $\beta$ -emitting radionuclides, which have favorable physical characteristics for use in therapy (Table 1).

Differences in chemical structure (as in the chelator, radioisotope, or SSA) imply differences in affinity and biodistribution (7–9). In an *in vitro* comparison, an increase in the affinity for SSTR-2 of up to 12-fold was seen with a single substitution (Table 2; Fig. 1) (10). The choice of the radioisotope is crucial for both diagnosis and therapy; the introduction of <sup>68</sup>Ga-labeled SSAs for PET has dramatically changed the diagnostic approach for NETs. From the therapeutic standpoint, despite no clinically demonstrated superiority in the affinity of <sup>90</sup>Y- versus <sup>177</sup>Lu-labeled SSAs, the greater manageability of the latter—from a dosimetric point of view—has made it the compound of choice for peptide receptor radionuclide therapy (PRRT). Given that <sup>90</sup>Y is a pure  $\beta$ -emitter, yttrium-based compounds cannot strictly be regarded as theranostic agents. Post-therapeutic imaging relies on the addition of a small amount of [<sup>111</sup>In-DOTA<sup>0</sup>-Tyr<sup>3</sup>]SSA to the administered therapeutic radiopharmaceutical. SSTR-2–targeting theranostic agents used in clinical practice are <sup>111</sup>In-pentetreotide ([<sup>111</sup>In-DTPA<sup>0</sup>]octreotide), [<sup>177</sup>Lu-DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotide, [<sup>177</sup>Lu-DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotate, [<sup>177</sup>Lu-DOTA<sup>0</sup>]lanreotide, [<sup>90</sup>Y-DOTA<sup>0</sup>-Tyr<sup>3</sup>]

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**TABLE 1**  
Characteristics of Radionuclides Used in PRRT (7,49,52)

Radionuclide	Half-life	Type	Decay energy	Maximum tissue penetration
<sup>111</sup> In	2.8 d	Conversion	144–245 keV	0.2–0.55 mm
		Auger	0.5–25 keV	0.02–10 μm
		γ	171–245 keV	
<sup>90</sup> Y	2.7 d	β	2.3 MeV	12 mm
<sup>177</sup> Lu	6.7 d	β	0.5 MeV	2 mm
		γ	113–208 keV	
<sup>213</sup> Bi	46 min	α	5.5–5.9 MeV	100 μm
		β	1.0–1.4 MeV	
		γ	440 keV	

**TABLE 2**  
SSTR-2 Affinity (10)

Chemical	50% Inhibitory concentration (nM)*	SEM
Octreotide	2.0	0.7
Indium-labeled DTPA-octreotide	22	3.6
Yttrium-labeled DOTA-Tyr <sup>3</sup> -octreotide	11	1.7
Yttrium-labeled DOTA-Tyr <sup>3</sup> -octeotate	1.6	0.4
Gallium-labeled DOTA-Tyr <sup>3</sup> -octreotide	2.5	0.5
Gallium-labeled DOTA-Tyr <sup>3</sup> -octeotate	0.2	0.04

\*Lower values reflect greater affinity.

octreotide, [<sup>90</sup>Y-DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotate, [<sup>90</sup>Y-DOTA<sup>0</sup>]lanreotide, [<sup>213</sup>Bi-DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotide, and [<sup>177</sup>Lu-DOTA]JR11.

### SSTR IMAGING

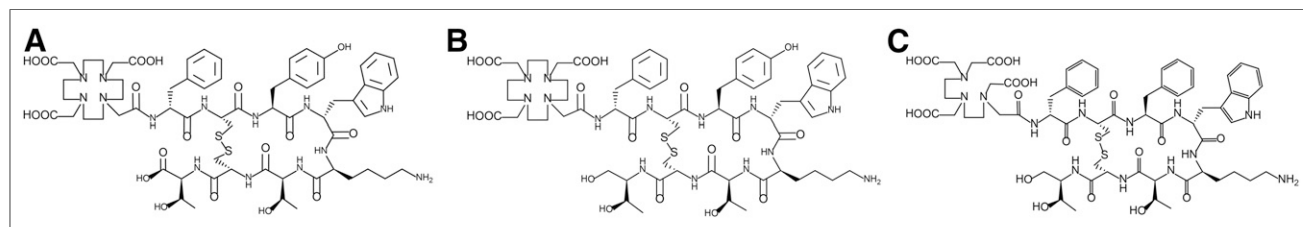
#### <sup>111</sup>In-Pentetreotide

Radiolabeled SSAs are used in localization, staging or restaging, and therapy selection; <sup>111</sup>In-pentetreotide was the first scintigraphic agent to be approved for NETs and has been the most widely studied (4). Compared with the sensitivity of available morphologic imaging, the sensitivity of <sup>111</sup>In-pentetreotide scintigraphy for gastroenteropancreatic and bronchopulmonary NETs and paragangliomas was well documented in the 1990s as being greater than 75% (11). At present, compared with CT or MRI, <sup>111</sup>In-pentetreotide is considered suboptimal, with a sensitivity of less than 60% (12). However, with

<sup>111</sup>In-pentetreotide whole-body scintigraphy, the therapeutic strategy is still modified in up to 50% of cases (13).

#### <sup>68</sup>Ga-SSA PET

In the last 15 y, molecular imaging of NETs has been revolutionized by the introduction of <sup>68</sup>Ga-SSAs for PET/CT (14). Numerous advantages, such as easy synthesis from a <sup>68</sup>Ge/Ga generator, higher spatial resolution (~4–5 mm), image quantification (SUV), favorable dosimetry, and the possibility of modifying clinical management in 36%–55% of cases, made <sup>68</sup>Ga-SSA PET/CT the current technique of choice (15,16). [<sup>68</sup>Ga-DOTA<sup>0</sup>-Tyr<sup>3</sup>]octeotate was recently approved by the U.S. Food and Drug Administration, and [<sup>68</sup>Ga-DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotide is expected to be approved by the European Medicines Agency soon.



**FIGURE 1.** Structural formulae of DOTA<sup>0</sup>-Tyr<sup>3</sup>-octreotate (A), DOTA<sup>0</sup>-Tyr<sup>3</sup>-octreotide (B), and DTPA<sup>0</sup>-octreotide (C), also known as DOTATATE, DOTATOC, and DTPA-OC or pentetreotide, respectively.

**TABLE 3**  
Phase 2 Trials with Radiolabeled Somatostatin Analogs for Treatment of Advanced NETs

Study	Institution	Year	Radiopharmaceutical	Schedule	No. of patients	Tumor type	Tumor response					Baseline PD (%)		Criterion	Follow-up (mo) <sup>†</sup>	Survival <sup>‡</sup>	
							CR (%)	PR (%)	SD (%)	PD (%)	NA (%)	DCR (%) <sup>*</sup>	PD (%)			Overall	Progression-free
Anthony et al. (23)	Louisiana State University Medical Center	2002	[ <sup>111</sup> In-DTPA <sup>0</sup> -Tyr <sup>3</sup> ] octreotide	6.6 GBq × (mean) 3 cycles (range, 1–7 cycles)	27	GEP	0	7	78	11	4	85	NA	NA	18	NA	
Walcherr et al. (53)	University Hospital Basel	2001	[ <sup>90</sup> Y-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotide	6 GBq/m <sup>2</sup> × 4 cycles	41	NET	2	22	61	15	0	85	83	WHO	15	NR	
Walcherr et al. (54)	University Hospital Basel	2002	[ <sup>90</sup> Y-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotide	7.4 GBq/m <sup>2</sup> × 4 cycles	39	NET	5	18	69	8	0	92	100	WHO	NA	NA	
Bushnell et al. (40)	Multiple centers	2010	[ <sup>90</sup> Y-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotide	4.4 GBq × 3 cycles	90	GEP/B	0	4	70	12	14	74	100	SWOG	NA	27	
Cwikla et al. (38)	CCHMIAA	2010	[ <sup>90</sup> Y-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotide	3.7 GBq × 3 cycles	60	GEP	0	22	73	5	0	95	100	RECIST	NA	22	
Savelli et al. (55)	Brescia Civic Hospital	2012	[ <sup>90</sup> Y-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotide	2.56 GBq × 4 cycles	38	GEP	0	45	26	29	0	71	100	RECIST	NA	NA	
Kwekkeboom et al. (36)	Erasmus University Medical Center	2008	[ <sup>177</sup> Lu-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotate	7.4 Gbq × 4 cycles	310	GEP	2	28	51	20	0	81	43	SWOG	19	46	
Bodei et al. (37)	IEO	2011	[ <sup>177</sup> Lu-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotate	25.2–26.4 GBq × 4–6 cycles	51	SSTR+	2	27	53	18	0	82	76	RECIST	29	NR <sup>§</sup>	
Sansovini et al. (41)	IRST-IRCCS	2013	[ <sup>177</sup> Lu-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotate	17.8–25.5 GBq × 5 cycles	51	Pancreas	8	21	52	19	0	81	100	SWOG	NA	NR	
Paganelli et al. (56)	IRST-IRCCS	2014	[ <sup>177</sup> Lu-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotate	18.5–27.8 GBq × 5 cycles	43	GI	7	0	77	16	0	84	100	SWOG	38	NR	
Delpassand et al. (57)	EDNOC	2014	[ <sup>177</sup> Lu-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotate	7.4 GBq × (mean) 3 cycles	37	GEP	0	24	38	24	14	62	100	RECIST	14 <sup>  </sup>	NA	
Baum et al. (42)	Zentralklinik Bad Berka	2016	[ <sup>177</sup> Lu-DOTA <sup>0</sup> -Tyr <sup>3</sup> ] octreotide	7.0 GBq × 1–4 cycles	56	NET	16	18	32	34	0	66	100	RECIST	16 <sup>  </sup>	34	

\*Disease control rates in studies of Anthony et al. (23), Bushnell et al. (40), and Delpassand et al. (57) were 88%, 86%, and 72%, respectively, in evaluable patients.

<sup>†</sup>Reported as median unless otherwise indicated.

<sup>‡</sup>Reported as median months unless otherwise indicated. NR = not reached.

<sup>§</sup>Median time to progression was 36 months.

<sup>||</sup>Reported as mean.

CR = percentage of patients showing complete response; PR = percentage of patients showing partial response; SD = percentage of patients showing stable disease (including minor response, if reported); baseline PD = percentage of patients showing progressive disease before treatment; PD = percentage of patients showing progressive disease; NA = not available/assessable; DCR = disease control rate (percentage); FU = median follow-up (months) unless otherwise indicated; GEP = gastroenteropancreatic; WHO = World Health Organization; GEP/B = gastroenteropancreatic/bronchial; SWOG = Southwest Oncology Group; CCHMIAA = Central Clinical Hospital of Ministry of Internal Affairs and Administrations, Warsaw, Poland; IEO = Istituto Europeo di Oncologia, Milan, Italy; SSRT+ = (all) somatostatin receptor-positive tumors; IRST-IRCCS = Istituto Scientifico Romagnolo per lo Studio e la Cura dei Tumori-Istituto Di Ricovero e Cura a Carattere Scientifico, Meldola, Italy; GI = gastrointestinal; EDNOC = Excel Diagnostics and Nuclear Oncology Center, Houston, Texas.

The overall sensitivity of  $^{68}\text{Ga}$ -SSA PET/CT for NETs is greater than 90%, whereas specificity ranges from 92%–98%—better than that for CT scanning and  $^{111}\text{In}$ -pentetretotide scintigraphy, particularly for small tumors at a nodal or bone level (12,16,17). Unlike  $^{111}\text{In}$ -pentetretotide scintigraphy, [ $^{68}\text{Ga}$ -DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotate PET/CT has a consistent visual and semi-quantitative image interpretation among experienced and inexperienced readers. However, its use in theranostics to recommend or exclude PRRT requires practice and training (18).

### Receptor Quantification

The estimation of SSTR density is used to assess the viability of cold and radiolabeled SSA therapy as a prediction of the amount of (radio)pharmaceutical that will be concentrated at the tumor site and, hence, the possibility of a response (11). For  $^{111}\text{In}$ -pentetretotide, this estimate, expressed on the Krenning scale, is based on the relative uptake of the tumor compared with that of normal organs (liver or kidneys and spleen) on the planar image (11). The same concept can be applied to  $^{68}\text{Ga}$ -SSA PET/CT imaging (e.g., on the volumetric image). Furthermore, uptake can be objectively quantified as SUV, which strongly correlates (linearly under a threshold of 25) with the inhibitor constant on dynamic PET and, hence, SSTR levels (19–21). Although criteria have been validated for  $^{111}\text{In}$ -pentetretotide with the 4-point Krenning scale, there is no consensus on what should be considered sufficient uptake on  $^{68}\text{Ga}$ -SSA PET/CT. The findings of one study have suggested that tumor-to-spleen SUV ratio is superior to  $\text{SUV}_{\text{max}}$  in the early prediction of response (22).

### PRRT

#### Early Results

Starting in the early 1990s, high doses of  $^{111}\text{In}$ -pentetretotide were administered to patients who had high uptake on diagnostic scans to achieve therapeutic results. Doses of up to 18.5 GBq/cycle and, cumulatively, 160 GBq were administered, resulting in partial response rates of up to 7.5%. In addition, improvement of symptoms was reported in up to 62% of cases, and toxicity rates were acceptable (23–25). With the introduction of the chelator DOTA, SSAs could be linked to  $^{177}\text{Lu}$  and  $^{90}\text{Y}$ , increasing the dose to the tumor and making  $^{111}\text{In}$ -pentetretotide obsolete for therapeutic purposes. Here we provide an overview of the safety and efficacy of PRRT with  $\beta$ -emitters.

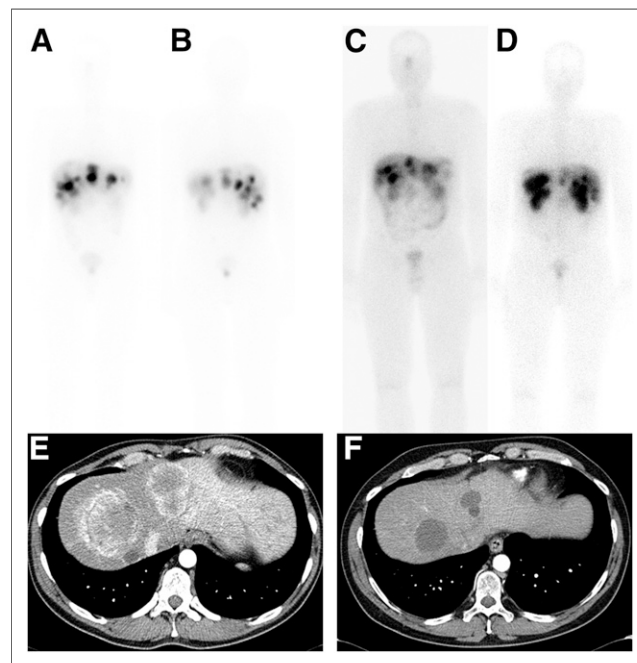
#### Safety

PRRT is generally very well tolerated. The full treatment is usually administered in 3 or 4 fractions at intervals of 6–12 wk, so as not to exceed single high doses to organs at risk. Acute toxicity, in the form of nausea (with or without vomiting), diarrhea, and abdominal pain, is mild and effectively treated with antiemetics and analgesics.

Because of transient bone marrow suppression, some subacute and usually self-limiting hematologic toxicity can be expected. In large trials, grade 3 or 4 hematologic toxicity has been reported in 3%–14% of the patients (26–28). Long-term myelotoxicity in the form of myelodysplastic syndrome or acute leukemia is a rare and severe adverse event associated with PRRT, occurring in 1%–2% of patients (29). Incidence rates are higher in patients who have been heavily pretreated with alkalinizing chemotherapeutics, probably reflecting the myelotoxic properties of these agents (30,31).

Long-term or persistent renal toxicity is considered rare. SSAs are partially reabsorbed in the proximal tubule cells; counteracting this reabsorption with the coinfusion of positively charged amino acids during treatment results in a mean dose reduction in the kidneys of 40% (32). Severe renal toxicity has been reported in 0%–9% of patients; high incidences have been reported in some trials with  $^{90}\text{Y}$  because of its biologic and physical properties and treatment in an early era without amino acid renal protection (26,27). With renal protection being the standard of care in international protocols, some radiation nephropathy is still considered normal. In 208 patients, an average annual decrease in creatinine clearance of  $3.4\% \pm 0.4\%$  was observed after [ $^{177}\text{Lu}$ -DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotate treatment (33).

Dosimetry has been proposed to optimize PRRT. Dose thresholds extrapolated from external-beam radiotherapy were initially proposed for the kidneys and bone marrow. The subsequent introduction of the “biologically effective dose” concept, at least for the kidneys, provided a better dose–effect correlation (34). However, although renal dosimetry is more reliable, bone marrow dosimetry is still in need of fine-tuning. Because threshold doses for renal toxicity are infrequently reached or are largely exceeded with standard  $^{177}\text{Lu}$ -based PRRT, and because long-term follow-up of large patient cohorts has indicated a very low risk of severe renal toxicity, routine dosimetry stratification of PRRT candidates is worth exploring in nonstandard treatments, such as retreatments (or salvage treatments) (26,35,36). Given that the bone marrow is



**FIGURE 2.** (A–D). Anterior (A and C) and posterior (B and D) planar whole-body scintigrams obtained 24 h after first (A and B) and fourth (C and D) treatments with [ $^{177}\text{Lu}$ -DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotate in 43-y-old patient with grade 2 (Ki-67, 15%) rectal NET with metastases to liver. After treatment (C and D), physiologic uptake was more pronounced in pituitary gland, kidneys, spleen, and bowels. This decrease in tumor-to-organ ratio may indicate favorable outcome. (E and F) Contrast-enhanced CT of abdomen before (E) and 2 mo after (F) treatment. In addition to decrease in size after treatment, pretherapeutic arterial enhancement was lost and lesions became hypodense, indicating therapeutic efficacy.

the major organ at risk for toxicity after  $^{177}\text{Lu}$ -based PRRT, refined dosimetric methods, possibly based on microdosimetry, are warranted.

### Efficacy

Objective response rates (complete response plus partial response) ranged from 4% to 45% in available phase 2 trials and might be limited in several trials (Table 3). However, analysis of survival data showed that survival in patients with a partial response might be similar to that in patients with stable disease after PRRT (36–39). With these findings in mind, for the evaluation of a response, one could consider disease control rates (complete response, partial response, and stable disease), which ranged from 62% to 95% in trials including only patients with progressive disease at baseline (38–42). Figure 2 shows the efficacy of PRRT in a patient with rectal NET with metastases to the liver.

Irrespective of radiographic outcome, PRRT can improve patients' quality of life. The secretion of bioactive hormones by NETs can result in severe symptoms and life-threatening crises. Small intestinal NETs are mainly associated with the carcinoid syndrome, which includes flushing, diarrhea, bronchospasm, tachycardia, and anxiety. PRRT has been shown to improve patients' quality of life and performance and to decrease a range of symptoms, such as fatigue, nausea, pain, dyspnea, insomnia, and diarrhea (43). In patients with functional pancreatic NETs, PRRT proved very successful in achieving durable control of severe hypoglycemia in insulinomas (44).

In January 2017, numerous phase 2 trials and case series were validated with the publication of the interim analysis of the Neuroendocrine Tumor Therapy (NETTER-1) trial (45). This

phase 3 trial included randomization between treatment with [ $^{177}\text{Lu}$ -DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotate ( $4 \times 7.4$  GBq) and treatment with high-dose, long-acting, repeatable octreotide in 229 patients with advanced midgut NETs and disease progression after first-line SSA therapy. The 3 most important findings were that for the PRRT arm, the median progression-free survival was not reached (vs. 8.4 mo for the control arm), the hazard ratio for disease progression or death was 0.21, and the objective tumor response was 18% (vs. 3% for the control arm).

Unlabeled SSAs are the first line of treatment in advanced NETs because of their highly favorable toxicity profile (46). However, after failure of this treatment, therapeutic options are limited. Previous randomized trials demonstrated the efficacy of everolimus and sunitinib (Table 4) and led to the implementation of these targeted drugs. With the publication of the NETTER-1 trial, PRRT has been added to the standard of care. Further randomized trials are needed to compare the efficacy and the toxicity profiles of PRRT and targeted therapies to clarify the best sequencing of the treatment algorithm.

### FUTURE DEVELOPMENTS

Advances in SSTR-2 targeting were obtained with the SSTR antagonist [ $^{177}\text{Lu}$ -DOTA]JR11. In vitro and in vivo animal studies demonstrated significant increases in (membrane-bound) tumor uptake, tumor radiation dose, and durable double-strand DNA breaks with the SSTR antagonist [ $^{177}\text{Lu}$ -DOTA]JR11 compared with the SSTR agonist [ $^{177}\text{Lu}$ -DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotate (47). In 4 patients, [ $^{177}\text{Lu}$ -DOTA]JR11 resulted in increased residence times, tumor uptake and, hence, tumor dose compared with [ $^{177}\text{Lu}$ -DOTA<sup>0</sup>-Tyr<sup>3</sup>]octreotate (48). Twelve months

**TABLE 4**  
Randomized Controlled Trials for Treatment of Advanced NETs (64)

Study	Trial name	Year	Tumor type	Intervention	Control	Median PFS (mo)	HR (95% CI)
Rinke et al. (58)	PROMID	2009	Midgut	Octreotide LAR (30 mg/4 wk)	Placebo	14 vs. 6	0.34 (0.20–0.59)
Caplin et al. (59)	CLARINET	2014	Pancreatic, midgut, hindgut	Lanreotide (120 mg/4 wk)	Placebo	NR at 24 vs. 18	0.47 (0.30–0.73)
Pavel et al. (60)	RADIANT-2	2011	NET + carcinoid syndrome	Everolimus (10 mg/d)*	Placebo*	16 vs. 11	0.77 (0.59–1.00)
Yao et al. (61)	RADIANT-3	2011	Pancreatic	Everolimus (10 mg/d)*	Placebo*	11 vs. 5	0.35 (0.27–0.45)
Yao et al. (62)	RADIANT-4	2016	Nonfunctioning lung/gastrointestinal tract	Everolimus (10 mg/d)*	Placebo*	11 vs. 4	0.48 (0.35–0.67)
Raymond et al. (63)		2011	Pancreatic	Sunitinib (37.5 mg/d)*	Placebo*	11 vs. 6	0.42 (0.26–0.66)
Strosberg et al. (45)	NETTER-1	2017	Midgut	[ $^{177}\text{Lu}$ -DOTA <sup>0</sup> -Tyr <sup>3</sup> ]octreotate (7.4 GBq $\times$ 4 cycles)	Octreotide LAR, 60 mg/mo	NR vs. 8	0.21 (0.13–0.34)

\*With continuation of somatostatin analog therapy.

PFS = progression-free survival (intervention vs. control); HR = hazard ratio for disease progression and (disease-related) death; LAR = long-acting and repeatable; NR = not reached.

after treatment, 2 patients showed a partial response, 1 showed stable disease, and 1 showed progressive disease, all without long-term renal or hematologic toxicity.

Studies have been performed with  $\alpha$ -emitting [ $^{213}\text{Bi-DOTA}^0\text{-Tyr}^3$ ]octreotide. Theoretically, the high-energy particles can increase double-strand DNA breaks, and the limited tissue range (2 cell diameters) can decrease collateral damage (Table 1). In 7 patients with  $\beta$ -emitting PRRT-refractory NETs, 13.3–20.8 GBq of [ $^{213}\text{Bi-DOTA}^0\text{-Tyr}^3$ ]octreotide was administered in 2–5 dose-escalating cycles (49). The patients had predominantly progressive liver disease, and treatment was administered intraarterially. Treatment was effective in all patients; at the time of analysis, the response in the liver had been maintained for 12–34 mo. Subacute toxicity was limited, but 1 (chemotherapy-naïve) patient developed myelodysplastic syndrome 24 mo after treatment. In addition, the  $\gamma$ -emitting capacities of  $^{213}\text{Bi}$  decay were used for posttherapeutic scintigraphy.

As changes in disease management are often based on morphologic and molecular imaging, the development of a new PET tracer—[ $^{64}\text{Cu-DOTA}^0\text{-Tyr}^3$ ]octreotate—may further enhance therapeutic decision making. Compared with  $^{68}\text{Ga-SSA}$  PET tracers, this tracer has a longer half-life and lower positron decay energy, theoretically leading to a favorable tumor-to-background ratio and better detection of small lesions (50). In a direct comparison, the patient-based sensitivities were equal, but [ $^{64}\text{Cu-DOTA}^0\text{-Tyr}^3$ ]octreotate had a higher lesion detection rate and was superior at identifying affected organs (51).

On a biochemical level, adding the combination of the DNA methyltransferase inhibitor 5-aza-2'-deoxycytidine and the histone deacetylase inhibitor valproic acid may be beneficial. A study of human pancreatic NET cell lines showed epidrug-induced upregulation of SSTR-2 with increased uptake of radiolabeled octreotide, which may be useful in patients with low levels of SSTR-2 expression (52).

## CONCLUSION

From the first in vivo SSTR scintigraphy with  $^{123}\text{I-Tyr}^3$ -octreotide in 1987 to the first phase 3 trial with [ $^{177}\text{Lu-DOTA}^0\text{-Tyr}^3$ ]octreotate, 3 decades of development have inseparably linked SSTR-targeting theranostic agents to NETs. PRRT is on the verge of becoming the standard of care for patients with well-differentiated inoperable or metastatic NETs and disease progression after first-line SSA therapy. Several preclinical and clinical developments, such as the use of SSA antagonists and  $\alpha$ -emitting radionuclides, stand to enhance the diagnostic and therapeutic properties of SSTR-targeting theranostic agents and can enter phase 1 or 2 trials. At this time, there is a need for further randomized trials to identify the optimal, multidisciplinary sequencing of long-lasting treatments for these patients.

## DISCLOSURE

No potential conflict of interest relevant to this article was reported.

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## REFERENCES

1. Kvols LK, Moertel CG, O'Connell MJ, Schutt AJ, Rubin J, Hahn RG. Treatment of the malignant carcinoid syndrome: evaluation of a long-acting somatostatin analogue. *N Engl J Med*. 1986;315:663–666.
2. Lamberts SW, Uitterlinden P, Verschoor L, van Dongen KJ, del Pozo E. Long-term treatment of acromegaly with the somatostatin analogue SMS 201-995. *N Engl J Med*. 1985;313:1576–1580.
3. Reubi JC, Waser B, Schaer JC, Laissue JA. Somatostatin receptor sst1-sst5 expression in normal and neoplastic human tissues using receptor autoradiography with subtype-selective ligands. *Eur J Nucl Med*. 2001;28:836–846.
4. Krenning EP, Kwekkeboom DJ, Bakker WH, et al. Somatostatin receptor scintigraphy with [ $^{111}\text{In-DTPA-D-Phe}^1$ ] and [ $^{123}\text{I-Tyr}^3$ ]octreotide: the Rotterdam experience with more than 1000 patients. *Eur J Nucl Med*. 1993;20:716–731.
5. Mizutani G, Nakanishi Y, Watanabe N, et al. Expression of somatostatin receptor (SSTR) subtypes (SSTR-1, 2A, 3, 4 and 5) in neuroendocrine tumors using real-time RT-PCR method and immunohistochemistry. *Acta Histochem Cytochem*. 2012;45:167–176.
6. Krenning EP, Bakker WH, Kooij PP, et al. Somatostatin receptor scintigraphy with indium-111-DTPA-D-Phe-1-octreotide in man: metabolism, dosimetry and comparison with iodine-123-Tyr-3-octreotide. *J Nucl Med*. 1992;33:652–658.
7. Kwekkeboom DJ, Bakker WH, Kooij PP, et al. [ $^{177}\text{Lu-DOTA}^0\text{Tyr}^3$ ]octreotate: comparison with [ $^{111}\text{In-DTPA}^0$ ]octreotide in patients. *Eur J Nucl Med*. 2001;28:1319–1325.
8. Forrer F, Uusijarvi H, Waldherr C, et al. A comparison of  $^{111}\text{In-DOTATOC}$  and  $^{111}\text{In-DOTATATE}$ : biodistribution and dosimetry in the same patients with metastatic neuroendocrine tumours. *Eur J Nucl Med Mol Imaging*. 2004;31:1257–1262.
9. Esser JP, Krenning EP, Teunissen JJ, et al. Comparison of [ $^{177}\text{Lu-DOTA}^0\text{Tyr}^3$ ]octreotate and [ $^{177}\text{Lu-DOTA}^0\text{Tyr}^3$ ]octreotide: which peptide is preferable for PRRT? *Eur J Nucl Med Mol Imaging*. 2006;33:1346–1351.
10. Reubi JC, Schar JC, Waser B, et al. Affinity profiles for human somatostatin receptor subtypes SST1-SST5 of somatostatin radiotracers selected for scintigraphic and radiotherapeutic use. *Eur J Nucl Med*. 2000;27:273–282.
11. Kwekkeboom DJ, Kam BL, van Essen M, et al. Somatostatin-receptor-based imaging and therapy of gastroenteropancreatic neuroendocrine tumors. *Endocr Relat Cancer*. 2010;17:R53–R73.
12. Gabriel M, Decristoforo C, Kendler D, et al.  $^{68}\text{Ga-DOTA-Tyr}^3$ -octreotide PET in neuroendocrine tumors: comparison with somatostatin receptor scintigraphy and CT. *J Nucl Med*. 2007;48:508–518.
13. de Mestier L, Dromain C, d'Assignies G, et al. Evaluating digestive neuroendocrine tumor progression and therapeutic responses in the era of targeted therapies: state of the art. *Endocr Relat Cancer*. 2014;21:R105–R120.
14. Treglia G, Castaldi P, Rindi G, Giordano A, Rufini V. Diagnostic performance of gallium-68 somatostatin receptor PET and PET/CT in patients with thoracic and gastroenteropancreatic neuroendocrine tumours: a meta-analysis. *Endocrine*. 2012;42:80–87.
15. Deppen SA, Blume J, Bobbey AJ, et al.  $^{68}\text{Ga-DOTATATE}$  compared with  $^{111}\text{In-DTPA-octreotide}$  and conventional imaging for pulmonary and gastroenteropancreatic neuroendocrine tumors: a systematic review and meta-analysis. *J Nucl Med*. 2016;57:872–878.
16. Ambrosini V, Campana D, Bodei L, et al.  $^{68}\text{Ga-DOTANOC}$  PET/CT clinical impact in patients with neuroendocrine tumors. *J Nucl Med*. 2010;51:669–673.
17. Srirajakanthan R, Kayani I, Quigley AM, Soh J, Caplin ME, Bomanji J. The role of  $^{68}\text{Ga-DOTATATE}$  PET in patients with neuroendocrine tumors and negative or equivocal findings on  $^{111}\text{In-DTPA-octreotide}$  scintigraphy. *J Nucl Med*. 2010;51:875–882.
18. Fendler WP, Barrio M, Spick C, et al.  $^{68}\text{Ga-DOTATATE}$  PET/CT interobserver agreement for neuroendocrine tumor assessments: results from a prospective study on 50 patients. *J Nucl Med*. 2017;58:307–311.
19. Van Binnebeek S, Koole M, Terwinghe C, et al. Dynamic  $^{68}\text{Ga-DOTATOC}$  PET/CT and static image in NET patients: correlation of parameters during PRRT. *Nuklearmedizin*. 2016;55:104–114.
20. Velikyan I, Sundin A, Sorensen J, et al. Quantitative and qualitative intrapatient comparison of  $^{68}\text{Ga-DOTATOC}$  and  $^{68}\text{Ga-DOTATATE}$ : net uptake rate for accurate quantification. *J Nucl Med*. 2014;55:204–210.
21. Kratochwil C, Stefanova M, Mavriopoulou E, et al. SUV of [ $^{68}\text{Ga}$ ]DOTATOC-PET/CT predicts response probability of PRRT in neuroendocrine tumors. *Mol Imaging Biol*. 2015;17:313–318.
22. Haug AR, Auernhammer CJ, Wangler B, et al.  $^{68}\text{Ga-DOTATATE}$  PET/CT for the early prediction of response to somatostatin receptor-mediated radionuclide therapy in patients with well-differentiated neuroendocrine tumors. *J Nucl Med*. 2010;51:1349–1356.

23. Anthony LB, Woltering EA, Espenan GD, Cronin MD, Maloney TJ, McCarthy KE. Indium-111-pentetreotide prolongs survival in gastroenteropancreatic malignancies. *Semin Nucl Med.* 2002;32:123–132.
24. Delpassand ES, Samarghandi A, Mourta JS, et al. Long-term survival, toxicity profile, and role of F-18 FDG PET/CT scan in patients with progressive neuroendocrine tumors following peptide receptor radionuclide therapy with high activity In-111 pentetreotide. *Theranostics.* 2012;2:472–480.
25. Valkema R, De Jong M, Bakker WH, et al. Phase I study of peptide receptor radionuclide therapy with [In-DTPA]octreotide: the Rotterdam experience. *Semin Nucl Med.* 2002;32:110–122.
26. Bodei L, Kidd M, Paganelli G, et al. Long-term tolerability of PRRT in 807 patients with neuroendocrine tumours: the value and limitations of clinical factors. *Eur J Nucl Med Mol Imaging.* 2015;42:5–19.
27. Imhof A, Brunner P, Marinček N, et al. Response, survival, and long-term toxicity after therapy with the radiolabeled somatostatin analogue [<sup>90</sup>Y-DOTA]-TOC in metastasized neuroendocrine cancers. *J Clin Oncol.* 2011;29:2416–2423.
28. Bergsma H, Konijnenberg MW, Kam BL, et al. Subacute haematotoxicity after PRRT with <sup>177</sup>Lu-DOTA-octreotate: prognostic factors, incidence and course. *Eur J Nucl Med Mol Imaging.* 2016;43:453–463.
29. Kesavan M, Turner JH. Myelotoxicity of peptide receptor radionuclide therapy of neuroendocrine tumors: a decade of experience. *Cancer Biother Radiopharm.* 2016;31:189–198.
30. Brireau B, Hentic O, Lebtahi R, et al. High risk of myelodysplastic syndrome and acute myeloid leukemia after <sup>177</sup>Lu-octreotate PRRT in NET patients heavily pretreated with alkylating chemotherapy. *Endocr Relat Cancer.* 2016;23:L17–L23.
31. Bodei L, Modlin IM, Luster M, et al. Myeloid neoplasms after chemotherapy and PRRT: myth and reality. *Endocr Relat Cancer.* 2016;23:C1–C7.
32. Rolleman EJ, Valkema R, de Jong M, Kooij PP, Krenning EP. Safe and effective inhibition of renal uptake of radiolabelled octreotide by a combination of lysine and arginine. *Eur J Nucl Med Mol Imaging.* 2003;30:9–15.
33. Bergsma H, Konijnenberg MW, van der Zwan WA, et al. Nephrotoxicity after PRRT with <sup>177</sup>Lu-DOTA-octreotate. *Eur J Nucl Med Mol Imaging.* 2016;43:1802–1811.
34. Cremonesi M, Botta F, Di Dia A, et al. Dosimetry for treatment with radiolabelled somatostatin analogues: a review. *Q J Nucl Med Mol Imaging.* 2010;54:37–51.
35. Sandström M, Garske-Roman U, Granberg D, et al. Individualized dosimetry of kidney and bone marrow in patients undergoing <sup>177</sup>Lu-DOTA-octreotate treatment. *J Nucl Med.* 2013;54:33–41.
36. Kwekkeboom DJ, de Herder WW, Kam BL, et al. Treatment with the radiolabeled somatostatin analog [<sup>177</sup>Lu-DOTA<sup>0</sup>,Tyr<sup>3</sup>]octreotate: toxicity, efficacy, and survival. *J Clin Oncol.* 2008;26:2124–2130.
37. Bodei L, Cremonesi M, Grana CM, et al. Peptide receptor radionuclide therapy with <sup>177</sup>Lu-DOTATATE: the IEO phase I-II study. *Eur J Nucl Med Mol Imaging.* 2011;38:2125–2135.
38. Cwikla JB, Sankowski A, Seklecka N, et al. Efficacy of radionuclide treatment DOTATATE Y-90 in patients with progressive metastatic gastroenteropancreatic neuroendocrine carcinomas (GEP-NETs): a phase II study. *Ann Oncol.* 2010;21:787–794.
39. Vinjamuri S, Gilbert TM, Banks M, et al. Peptide receptor radionuclide therapy with <sup>90</sup>Y-DOTATATE/<sup>90</sup>Y-DOTATOC in patients with progressive metastatic neuroendocrine tumours: assessment of response, survival and toxicity. *Br J Cancer.* 2013;108:1440–1448.
40. Bushnell DL Jr, O'Doriso TM, O'Doriso MS, et al. <sup>90</sup>Y-edotreotide for metastatic carcinoid refractory to octreotide. *J Clin Oncol.* 2010;28:1652–1659.
41. Sansovini M, Severi S, Ambrosetti A, et al. Treatment with the radiolabelled somatostatin analog Lu-DOTATATE for advanced pancreatic neuroendocrine tumors. *Neuroendocrinology.* 2013;97:347–354.
42. Baum RP, Kluge AW, Kulkarni H, et al. [<sup>177</sup>Lu-DOTA]<sup>0</sup>-D-Phe<sup>1</sup>-Tyr<sup>3</sup>-octreotide (<sup>177</sup>Lu-DOTATOC) for peptide receptor radiotherapy in patients with advanced neuroendocrine tumours: a phase-II study. *Theranostics.* 2016;6:501–510.
43. Khan S, Krenning EP, van Essen M, Kam BL, Teunissen JJ, Kwekkeboom DJ. Quality of life in 265 patients with gastroenteropancreatic or bronchial neuroendocrine tumors treated with [<sup>177</sup>Lu-DOTA<sup>0</sup>,Tyr<sup>3</sup>]octreotate. *J Nucl Med.* 2011;52:1361–1368.
44. van Schaik E, van Vliet EI, Feelders RA, et al. Improved control of severe hypoglycemia in patients with malignant insulinomas by peptide receptor radionuclide therapy. *J Clin Endocrinol Metab.* 2011;96:3381–3389.
45. Strosberg J, El-Haddad G, Wolin E, et al. Phase 3 trial of <sup>177</sup>Lu-DOTATATE for midgut neuroendocrine tumors. *N Engl J Med.* 2017;376:125–135.
46. Pavel M, O'Toole D, Costa F, et al. ENETS consensus guidelines update for the management of distant metastatic disease of intestinal, pancreatic, bronchial neuroendocrine neoplasms (NEN) and NEN of unknown primary site. *Neuroendocrinology.* 2016;103:172–185.
47. Dalm SU, Nonnekens J, Doeswijk GN, et al. Comparison of the therapeutic response to treatment with a <sup>177</sup>Lu-labeled somatostatin receptor agonist and antagonist in preclinical models. *J Nucl Med.* 2016;57:260–265.
48. Wild D, Fani M, Fischer R, et al. Comparison of somatostatin receptor agonist and antagonist for peptide receptor radionuclide therapy: a pilot study. *J Nucl Med.* 2014;55:1248–1252.
49. Kratochwil C, Giesel FL, Bruchertseifer F, et al. <sup>213</sup>Bi-DOTATOC receptor-targeted alpha-radionuclide therapy induces remission in neuroendocrine tumours refractory to beta radiation: a first-in-human experience. *Eur J Nucl Med Mol Imaging.* 2014;41:2106–2119.
50. Pfeifer A, Knigge U, Binderup T, et al. <sup>64</sup>Cu-DOTATATE PET for neuroendocrine tumors: a prospective head-to-head comparison with <sup>111</sup>In-DTPA-octreotide in 112 patients. *J Nucl Med.* 2015;56:847–854.
51. Johnbeck CB, Knigge U, Loft A, et al. Head-to-head comparison of <sup>64</sup>Cu-DOTATATE and <sup>68</sup>Ga-DOTATOC PET/CT: a prospective study of 59 patients with neuroendocrine tumors. *J Nucl Med.* 2017;58:451–457.
52. Veenstra MJ, van Koetsveld PM, Dogan F, et al. Epidrug-induced upregulation of functional somatostatin type 2 receptors in human pancreatic neuroendocrine tumor cells. *Oncotarget.* May 19, 2016 [Epub ahead of print].
53. Waldherr C, Pless M, Maecke HR, Haldemann A, Mueller-Brand J. The clinical value of [<sup>90</sup>Y-DOTA]-D-Phe<sup>1</sup>-Tyr<sup>3</sup>-octreotide (<sup>90</sup>Y-DOTATOC) in the treatment of neuroendocrine tumours: a clinical phase II study. *Ann Oncol.* 2001;12:941–945.
54. Waldherr C, Pless M, Maecke HR, et al. Tumor response and clinical benefit in neuroendocrine tumors after 7.4 GBq <sup>90</sup>Y-DOTATOC. *J Nucl Med.* 2002;43:610–616.
55. Savelli G, Bertagna F, Franco F, et al. Final results of a phase 2A study for the treatment of metastatic neuroendocrine tumors with a fixed activity of <sup>90</sup>Y-DOTA-D-Phe<sup>1</sup>-Tyr<sup>3</sup> octreotide. *Cancer.* 2012;118:2915–2924.
56. Paganelli G, Sansovini M, Ambrosetti A, et al. <sup>177</sup>Lu-DOTA-octreotate radionuclide therapy of advanced gastrointestinal neuroendocrine tumors: results from a phase II study. *Eur J Nucl Med Mol Imaging.* 2014;41:1845–1851.
57. Delpassand ES, Samarghandi A, Zamanian S, et al. Peptide receptor radionuclide therapy with <sup>177</sup>Lu-DOTATATE for patients with somatostatin receptor-expressing neuroendocrine tumors: the first US phase 2 experience. *Pancreas.* 2014;43:518–525.
58. Rinke A, Muller HH, Schade-Brittinger C, et al. Placebo-controlled, double-blind, prospective, randomized study on the effect of octreotide LAR in the control of tumor growth in patients with metastatic neuroendocrine midgut tumors: a report from the PROMID Study Group. *J Clin Oncol.* 2009;27:4656–4663.
59. Caplin ME, Pavel M, Cwikla JB, et al. Lanreotide in metastatic enteropancreatic neuroendocrine tumors. *N Engl J Med.* 2014;371:224–233.
60. Pavel ME, Hainsworth JD, Baudin E, et al. Everolimus plus octreotide long-acting repeatable for the treatment of advanced neuroendocrine tumours associated with carcinoid syndrome (RADIANT-2): a randomised, placebo-controlled, phase 3 study. *Lancet.* 2011;378:2005–2012.
61. Yao JC, Shah MH, Ito T, et al. Everolimus for advanced pancreatic neuroendocrine tumors. *N Engl J Med.* 2011;364:514–523.
62. Yao JC, Fazio N, Singh S, et al. Everolimus for the treatment of advanced, non-functional neuroendocrine tumours of the lung or gastrointestinal tract (RADIANT-4): a randomised, placebo-controlled, phase 3 study. *Lancet.* 2016;387:968–977.
63. Raymond E, Dahan L, Raoul JL, et al. Sunitinib malate for the treatment of pancreatic neuroendocrine tumors. *N Engl J Med.* 2011;364:501–513.
64. Smit Duijzentkunst DA, Teunissen JJM, Kam BLR, Kwekkeboom DJ. Peptide receptor radionuclide therapy for neuroendocrine tumors [in Dutch]. *Tijdschr Nucl Geneesk.* 2016;38:1635–1644.