Validation of the Semiquantitative Static SUVR Method for ¹⁸F-AV45 PET by Pharmacokinetic Modeling with an Arterial Input Function

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Increased brain uptake of ¹⁸F-AV45 visualized by PET is a key biomarker for Alzheimer disease (AD). The SUV ratio (SUVR) is widely used for quantification, but is subject to variability based on choice of reference region and changes in cerebral blood flow. Here we validate the SUVR method against the gold standard volume of distribution (V_T) to assess cross-sectional differences in plaque load. Methods: Dynamic 60-min ¹⁸F-AV45 (291 ± 67 MBq) and 1-min ¹⁵O-H₂O (370 MBq) scans were obtained in 35 age-matched elderly subjects, including 10 probable AD, 15 amnestic mild cognitive impairment (aMCI), and 10 cognitively healthy controls (HCs). ¹⁸F-AV45 V_T was determined from 2-tissue-compartment modeling using a metabolite-corrected plasma input function. Static SUVR was calculated at 50-60 min after injection, using either cerebellar gray matter (SUVR_{CB}) or whole subcortical white matter (SUVR_{WM}) as the reference. Additionally, whole cerebellum, pons, centrum semiovale, and a composite region were examined as alternative references. Blood flow was quantified by ¹⁵O-H₂O SUV. Data are presented as mean ± SEM. Results: There was rapid metabolization of ¹⁸F-AV45, with only 35% of unchanged parent remaining at 10 min. Compared with V_T , differences in cortical $A\beta$ load between aMCI and AD were overestimated by SUVR_{WM} (+4% \pm 2%) and underestimated by SUVR_{CB} ($-10\% \pm 2\%$). V_T correlated better with SUVR_{WM} (Pearson r: from 0.63 for posterior cingulate to 0.89 for precuneus, P < 0.0001) than with SUVR_{CB} (Pearson r: from 0.51 for temporal lobe [P = 0.002] to 0.82 for precuneus [P < 0.0001]) in all tested regions. Correlation results for the alternative references were in between those for CB and WM. ¹⁵O-H₂O data showed that blood flow was decreased in AD compared with aMCI in cortical regions ($-5\% \pm 1\%$) and in the reference regions (CB, $-9\% \pm 8\%$; WM, $-8\% \pm 8\%$). **Conclusion:** Increased brain uptake of ¹⁸F-AV45 assessed by the simplified static SUVR protocol does not truly reflect $A\beta$ load. However, SUVR_{WM} is better correlated with V_T and more closely reflects V_T differences between aMCI and AD than SUVR_{CB}.

Key Words: Alzheimer's disease; ¹⁸F-AV45; kinetic modeling; cerebral perfusion; amyloid

J Nucl Med 2017; 58:1483–1489 DOI: 10.2967/jnumed.116.184481

 ${f A}$ ccording to the amyloid hypothesis, amyloid-eta (Aeta) aggregation plays a primary role in the etiology of Alzheimer disease (AD) (1). The formation of plaques consisting of Aβ in brain parenchyma probably starts decades before the onset of clinical symptoms (2). As such, AB deposition appears a suitable target for both early diagnosis and novel AD treatments (3,4). To date, the best validated method to diagnose AD is via measurement of $A\beta_{1-42}$, total tau and phospho-tau₁₈₁ levels in the cerebrospinal fluid (5). This, however, requires a lumbar puncture, and the concentration of AB isoforms in the cerebrospinal fluid is not a direct measure for AB levels in the brain. Therefore, noninvasive methods for detection and direct quantification of AB in the brain have been introduced more recently through amyloid PET imaging (6,7). Among all the small-molecule PET ligands that were developed in the last decade, ¹¹C-labeled Pittsburgh compound B is the most extensively studied (8,9). However, the short radioactive half-life of ¹¹C (20 min) limits its use as a diagnostic tool. In 2012, ¹⁸F-AV45 (¹⁸F-florbetapir, commercial name Amyvid [Eli Lilly]) was launched as the first Food and Drug Administration-approved commercial radiotracer for AB PET imaging in patients, because of its fast kinetics, high selectivity for AB plaques, and the longer half-life of ¹⁸F (110 min) (10).

Currently, the most widely used quantification index in amyloid PET imaging is the semiquantitative SUV ratio (SUVR), which is the ratio of radioactivity concentration of the target and reference regions. The whole cerebellum and cerebellar gray matter (CB) are commonly chosen as references because they are notably free from fibrillar A β in sporadic AD. However, converging evidence from the Alzheimer Disease Neuroimaging Initiative patients (11) as well as other recent studies (12–15) suggested that using SUVR normalized to the subcortical white matter (WM) could more

Received Sep. 21, 2016; revision accepted Mar. 6, 2017.

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Published online Mar. 23, 2017.

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accurately track longitudinal A β changes in amyloid-positive subjects. Hence, the optimal reference region still remains debated. Moreover, because conditions of true equilibrium are not fulfilled in a bolus experiment at later time frames, SUVR may become sensitive to regional blood flow changes (11,16,17). This effect is of great concern because hypoperfusion is a well-known feature of AD pathology (18,19), and many elderly AD patients have concomitant cerebrovascular disease (20). Notwithstanding the success of SUVR already achieved for diagnostic and longitudinal follow-up purposes, absolute quantification by dynamic PET imaging combined with arterial blood sampling and subsequent pharmacokinetic modeling excels as the gold standard (9). This methodology provides the most accurate and robust quantification of tracer uptake, expressed as total volume of distribution (V_T).

The present study was designed as a validation of the static SUVR method versus V_T for 18 F-AV45 PET imaging in cognitively healthy controls (HCs) and amnestic mild cognitive impairment (aMCI) and AD subjects. Both subcortical WM and CB were examined as potential reference regions for SUVR. In addition, we quantified brain perfusion by the 18 F-AV45 delivery rate as well as via reference standard 15 O-water PET scans.

MATERIALS AND METHODS

Subjects

Thirty-eight subjects were enrolled in the study, including 10 HCs and 18 aMCI and 10 AD subjects. Each subject underwent dynamic 60-min ¹⁸F-AV45 PET scanning, ¹⁵O-water PET scanning, MRI, and an extensive neuropsychologic examination. All patients were recruited from the Memory Clinics of Hospital Network Antwerp (ZNA) Middelheim and Hoge Beuken and University Hospital Brussels on a prospective basis. Their demographics are summarized in Table 1. A consensus clinical diagnosis was made by a panel of 3 medical doctors experienced with neurodegenerative brain diseases and dementia and was based on diagnostic criteria of the National Institute on Aging-Alzheimer's Association (21) and information gathered during the baseline clinical diagnostic workup at enrollment in the study. The cohort of cognitively healthy elderly was recruited among relatives of colleagues and patients visiting the memory clinic. The inclusion criteria were no neurologic or psychiatric antecedents; no organic disease involving the central nervous system after extensive clinical examination; and no cognitive complaint or decline. Cognitive decline was defined as performance of more than 1.5 SDs below the appropriate mean on the

TABLE 1Demographics, Including All Subjects

Parameter	HC (n = 10)	aMCI (n = 18)	AD $(n = 10)$
Age (y)	69 ± 6	73 ± 8	73 ± 5
Sex			
Male	4	9	7
Female	6	9	3
Education (y)	14 ± 2	11 ± 4	12 ± 4
MMSE	29 ± 1	25 ± 3*	22 ± 4*

^{*}Significantly different from controls (1-way ANOVA, corrected for multiple comparisons via Dunnett, P < 0.05).

$$\label{eq:MMSE} \begin{split} \text{MMSE} &= \text{Mini-Mental State Examination.} \\ \text{Data are mean } \pm \text{SD, unless otherwise stated.} \end{split}$$

neuropsychologic subtests. Approval for the study was obtained from the Committee for Medical Ethics of the University of Antwerp/ University Hospital Antwerp (14/12/130) and of Hospital Network Antwerp (ZNA) (4310), and all participants or their representatives provided informed consent.

Radiosynthesis of ¹⁸F-AV45

¹⁸F-AV45 was synthesized from the precursor AV-105 by a modification of the method described by Yao et al. (22) on a Fluorsynthon I synthesis module (Comecer Netherlands) adapted for automated production of ¹⁸F-AV45.

Data Acquisition

A 3-dimensional T1-weighted magnetization-prepared rapid gradient echo scan (repetition time, 1,910 ms; echo time, 3.37 ms; inversion time, 1,100 ms) was obtained on a Siemens Trio Tim 3T MR scanner with 1-mm isotropic voxels. All PET scans were acquired using a Siemens Biograph mCT time-of-flight PET scanner. Dynamic ¹⁸F-AV45 PET data were recorded over 60 min after an intravenous bolus injection of 291 \pm 67 MBq (mean \pm SD). The images were reconstructed using a 3-dimensional iterative algorithm (TrueX ultraHD-PET; Siemens), with 2 iterations and 21 subsets, into 23 frames (8 \times 15, 3 \times 60, 5 \times 120, 5×300 , 2×600 s) followed by a gaussian filter of 2 mm in full width at half maximum on a $400 \times 400 \times 74$ mm matrix with a $1 \times 1 \times 3$ mm voxel size. All images were corrected for random and scattered coincidences and attenuation based on a delayed coincidence window and a low-dose CT, respectively. Dynamic 15O-water data were acquired over 2 min after a bolus injection of 370 MBq and reconstructed, with 3 iterations and 24 subsets, into 2 frames $(2 \times 60 \text{ s})$ followed by a gaussian filter of 4 mm in full width at half maximum on a $200 \times 200 \times 74$ mm matrix with a $2 \times 2 \times 3$ mm voxel size.

Simultaneously with the dynamic ¹⁸F-AV45 PET acquisition, continuous arterial blood sampling was performed by a coincidence detector system (Twilite; Swisstrace) to measure radioactivity in blood. To evaluate the in vivo metabolism of ¹⁸F-AV45, arterial blood samples (3 mL) were manually collected at 2, 5, 7, 15, 30, 45, and 60 min after injection and centrifuged for 5 min at 4°C (4,500g) to collect the plasma. Total radioactivity in the whole blood and plasma samples (300 µL) was measured in a cross-calibrated automated γ-counter (Wizard² 2480; Perkin Elmer). For plasma protein precipitation, acetonitrile (300 μL) was added to the plasma sample as well as 10 µL of ¹⁸F-AV45 reference standard (1 mg/mL) for ultraviolet cross-validation, after which the samples were vortexed for 30 s and centrifuged for 4 min at 4°C (4,500g). The radioactivity of supernatant and precipitate was measured in the γ -counter to derive the extraction yield. Separation of metabolites and intact tracer was achieved by injection of the supernatant (100 µL) into a high-performance liquid chromatography column (XBridge, C18, 5 μm, 150 × 4.6 mm [Waters]), operating at a flow rate of 1 mL/min and eluted with a mixture of NaOAC (0.05 M, pH 5.5)/acetonitrile (50/50, V/V). The eluate of the column was sampled by a fraction collector (0.5 min per fraction), and radioactivity was measured in the collected fractions using the automated γ-counter. The radioactivity due to ¹⁸F-AV45 and each radiometabolite was expressed as a percentage of the total peak areas.

Image Analysis

To define the regions of interest (ROIs), brain parcellation of the anatomic MR images was performed using automated knowledge-based segmentation in PMOD (version 3.6; PMOD Technologies Ltd.) for each subject. Interframe motion correction of the dynamic PET images was based on normalized mutual information registration with mid frames (5–15 min) as a reference. The motion-corrected ¹⁸F-AV45 and ¹⁵O-water data were separately coregistered onto the MR images to extract regional time-activity curves. Typical ¹⁸F-AV45 time-activity curves reflect those shown in a previous study (23).

ROI-based partial-volume correction using a gaussian kernel with a full width at half maximum of $5.1 \times 5.1 \times 5.1$ mm was performed to correct the time–activity curves (13). ROIs were averaged over the left and right hemispheres and included frontal, temporal, parietal, occipital, anterior cingulate, posterior cingulate, precuneus, and cerebellar cortical gray matter and whole subcortical WM reference regions. Additionally, alternative reference regions were also included: whole cerebellum, pons, part of the centrum semiovale, and a composite region (whole cerebellum + pons + semiovale).

Region-Based V_T . The time course of the whole blood activity measured with the Twilite was decay- and background-corrected and cross-calibrated with the PET scanner at the day of the experiment. Therefore, a 0.9% NaCl phantom and catheter were filled with 74 MBq of ¹⁸F-FDG and measured in the PET and Twilite systems, respectively, to obtain the calibration factor. The calibrated blood activity concentration was approximated by a 3-exponential function and multiplied by both plasma-to-blood ratio and parent fraction to derive the metabolite-corrected plasma input function. All blood data processing was performed in PMOD (version 3.6). For model selection an array of pharmacokinetic models was tested on a subset of subjects (i.e., 5 subjects from each diagnostic group). Models included a 1-tissuecompartment model (1TCM), a 2-tissue-compartment (2TC) 4-parameter model with fitted blood volume fraction (2TCM-4k_V_b) and without V_b correction (2TCM-4k with $V_b = 0$), a 2TC 3-parameter model (2TCM-3k, that is, irreversible binding $[k_4 = 0]$), and a simplified reference tissue model with fixed k_2 ' (SRTM2). The relative weights of the individual data points were set as the inverse of the variance of the PET measurement error (24). A delay was fitted between Twilite and PET measurements based on a coupled fit across the cerebral lobes. The 2TCM-4k_V_b using the metabolite-corrected plasma input function was selected on the basis of the lowest Akaike information criteria scores and applied to the dynamic PET data in PMOD. Total V_T was determined from the 2TCM fit for each region based (V_T = $K_1/k_2(1 + k_3/k_4)$, with K_1 (mL/min/mL) and k_2 (1/min) the tracer transport rates from plasma to tissue and back, and k_3 (1/min) and

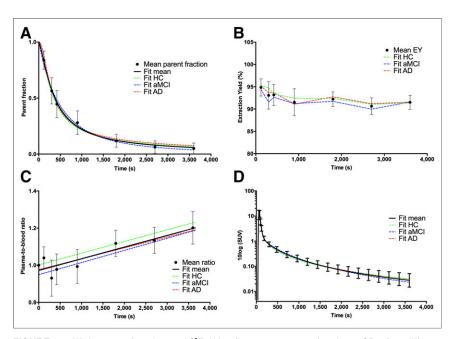


FIGURE 1. (A) Amount of unchanged ¹⁸F-AV45 (i.e., mean parent fraction ± SD, •) at different time points after injection for all subjects investigated; black solid line represents Watabe curve fit. Colored, dotted lines represent Watabe curve fits to group-averaged parent fractions. (B) Extraction yield (EY). (C) Plasma-to-whole-blood ratios fitted with linear function. (D) Metabolite-corrected plasma input functions.

 k_4 (1/min) the tracer transport rates from the nondisplaceable to the specific binding compartment and back, respectively.

Region-Based SUVR. SUVR was calculated for each ROI from the 50- to 60-min postinjection data, using either CB, WM, or one of the alternatives as the reference region. The regional averaged ¹⁵O-water SUV was calculated as the tissue radioactivity concentration (first 1-min frame only) normalized by injected dose and body weight.

Voxel-Based Analysis. Parametric V_T images were obtained using Ichise's multilinear analysis 1 method (t* = 20 min) implemented in PMOD (version 3.3; PMOD Technologies Ltd.) (9,25). To validate the use of Ichise's multilinear analysis 1 method for pixelwise modeling, ROIs were applied to the parametric V_T images and regional means were correlated with V_T from 2TCM (uncorrected). A voxel-based representation of the ¹⁸F-AV45 SUVR was created from the 50- to 60-min time frame, using either CB or WM as the reference region. Images of global blood flow were generated from the first frame of ¹⁵O-water SUV data.

Statistical Analysis

Data were analyzed in Prism (version 6; GraphPad Software), and all parameters are described as mean \pm SEM. Group differences across brain regions were determined using 2-way ANOVA, and P values were corrected using Tukey's multiple-comparisons test. The Pearson correlation coefficient r was calculated to examine the relationship between $^{18}\text{F-AV45}$ V $_{\text{T}}$ and SUVR as well as between $^{18}\text{F-AV45}$ K $_{\text{T}}$ and $^{15}\text{O-water}$ SUV for the different regions.

RESULTS

Subjects

A total of 10 HCs and 18 aMCI and 10 AD subjects enrolled in the study, of which 3 aMCI subjects were excluded. The metabolite analysis of 1 aMCI patient failed, most likely due to contamination, and another patient was fitted with a large temporal delay between Twilite and PET measurements (60 s instead of 28 ± 3 s, mean \pm

SD) during kinetic modeling. The V_T values of a third aMCI subject were unrealistically high (more than 2 SDs from the average of all patients). There was no significant difference in age, sex, or years of education between all controls and patients (Table 1).

Clearance and Metabolism of ¹⁸F-AV45

There was rapid metabolization of ¹⁸F-AV45 (retention time, 7.5 min), with on average 35% ± 10% of intact parent remaining at 10 min (Fig. 1A). Three main metabolite peaks were detected in plasma eluting with retention times of 1.5 (M1), 4 (M2), and 9 (M3). Supplemental Figure 1 (supplemental materials are available at http://jnm.snmjournals.org) shows radiochromatograms and corresponding percentage peak areas of all compounds at the different time points. Supplemental Table 1 reports the percentage peak areas. The average (±SD) extraction yield significantly decreased (P < 0.0001, Student t test) from 94.8% \pm 1.9% to 91.5% \pm 1.6% (Fig. 1B). The average ratio of total radioactivity in plasma to blood significantly increased (P < 0.0001, Student t test) from

Brain region	V_b	K ₁	k ₂	<i>k</i> ₃	k_4	V_{T}
Precuneus						
HC	0.05 ± 0.01	0.77 ± 0.12	0.17 ± 0.02	0.02 ± 0.01	0.02 ± 0.01	8.5 ± 2.9
aMCI	0.05 ± 0.01	0.69 ± 0.15	0.14 ± 0.04	0.03 ± 0.01	0.02 ± 0.005	12.1* ± 3.0
AD	0.05 ± 0.01	0.63* ± 0.10	$0.13^* \pm 0.03$	0.03 ± 0.01	0.02 ± 0.007	12.6* ± 1.6
СВ						
HC	0.05 ± 0.01	0.66 ± 0.12	0.17 ± 0.03	0.01 ± 0.01	0.02 ± 0.01	7.5 ± 2.3
aMCI	0.06 ± 0.01	0.65 ± 0.11	0.17 ± 0.03	0.01 ± 0.01	0.01 ± 0.01	8.3 ± 2.3
AD	0.05 ± 0.01	0.59 ± 0.09	0.16 ± 0.03	0.01 ± 0.003	0.01 ± 0.01	8.3 ± 2.0
WM						
HC	0.02 ± 0.01	0.28 ± 0.06	0.14 ± 0.04	0.19 ± 0.10	0.04 ± 0.01	10.9 ± 3.0
aMCI	0.02 ± 0.01	0.26 ± 0.05	0.13 ± 0.04	0.16 ± 0.11	0.03 ± 0.02	11.5 ± 2.3
AD	0.02 ± 0.01	0.25 ± 0.04	0.11 ± 0.04	0.12* ± 0.04	0.03 ± 0.01	10.9 ± 1.7

^{*}Significantly different from controls (1-way ANOVA, corrected for multiple comparisons via Dunnett, P < 0.05).

 1.04 ± 0.06 at 2 min to 1.2 ± 0.09 at 60 min (Fig. 1C). The resulting metabolite-corrected plasma input function of the tracer is depicted in Figure 1D. There was no significant difference in extraction yield, plasma-to-blood ratio, and plasma input function between the 3 groups (colored, dotted lines in Fig. 1).

¹⁸F-AV45 Imaging Results

Regional-Based V_T . All brain regions required the 2TCM-4k_V_b to adequately describe the time–activity curves as reflected by the lowest Akaike information criteria scores (125 \pm 15, mean \pm SD). V_T based on 2TCM-4k_V_b showed strong regional correlation (r > 0.95, P < 0.0001) with V_T from 2TCM-4k with $V_b = 0$ (Akaike information criteria, 141 \pm 15). Significantly higher Akaike information criteria scores were reached for the 1TCM (227 \pm 12), 2TCM-3k (165 \pm 11), and SRTM2 (154 \pm 25) (P < 0.0001, 2-way ANOVA for each ROI and model). The estimated kinetic

parameters and V_T from the 2TCM-4k_ V_b in the precuneus, CB, and WM for all diagnostic groups are listed in Table 2. Mean V_T (Fig. 2) was significantly increased compared with controls in the frontal, parietal, precuneus, and anterior and posterior cingulate cortical gray matter for the aMCI subjects (on average $+32\% \pm 3\%$, P < 0.05). These regions and additionally the temporal lobe were significantly increased for AD subjects (on average $+37\% \pm 3\%$, P < 0.05). Occipital and cerebellar V_T were elevated for the aMCI and AD groups compared with the control group, although not significantly (P > 0.1).

Regional-Based SUVR. SUVR_{CB} (Fig. 3A) was significantly increased compared with controls in the frontal, parietal, precuneus, anterior cingulate, and posterior cingulate cortical gray matter for the aMCI subjects (on average $+45\% \pm 5\%$, P < 0.05). These regions, except anterior cingulate, were also significantly increased for AD subjects (on average $+41\% \pm 6\%$, P < 0.05). SUVR_{WM}

(Fig. 3B) had significantly increased tracer retention in the frontal lobe, parietal lobe, precuneus, and anterior and posterior cingulate for the aMCI subjects (on average $+38\% \pm 5\%$, P < 0.05). These regions and additionally the temporal lobe were significantly increased for AD subjects (on average $+48\% \pm 5\%$, P < 0.05).

Comparison of SUVR Versus V_T . Compared with V_T , differences in cortical $A\beta$ load between the aMCI and AD group were overestimated by $SUVR_{WM}$ ($+4\% \pm 2\%$) and underestimated by $SUVR_{CB}$ ($-10\% \pm 2\%$). Differences in cortical $A\beta$ load between the HC and aMCI groups were overestimated by both $SUVR_{WM}$ ($+5\% \pm 2\%$) and $SUVR_{CB}$ ($+12\% \pm 2\%$). Table 3 represents Pearson correlation coefficients between SUVR and V_T for the different ROIs. Across diagnostic groups, V_T correlated better with $SUVR_{WM}$ (r ranging from 0.63 for posterior cingulate to 0.89 for precuneus, P < 0.0001)

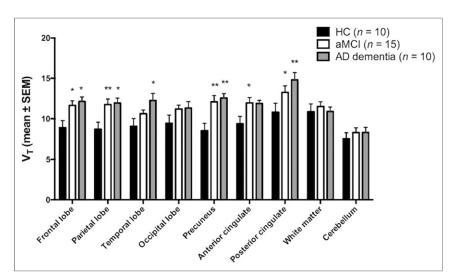


FIGURE 2. Regional ¹⁸F-AV45 V_T (mean \pm SEM) values for the 3 groups. Asterisks denote significant differences with HC group (*P < 0.05; **P < 0.01).

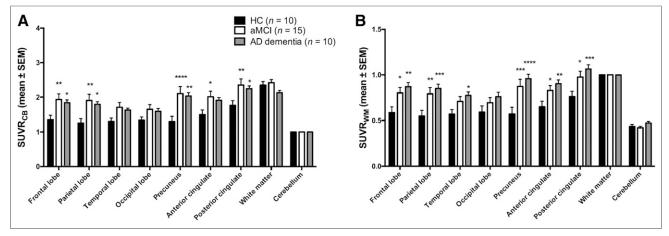


FIGURE 3. Regional ¹⁸F-AV45 SUVR_{CB} (A) and SUVR_{WM} (B) (mean \pm SEM) values for the 3 groups. Asterisks denote significant differences with HC group (*P < 0.05; **P < 0.01; ****P < 0.001; ****P < 0.0001).

than with SUVR_{CB} (r ranging from 0.51 for temporal lobe P = 10.002] to 0.82 for precuneus [P < 0.0001]) in all tested regions. Within diagnostic groups, SUVR_{WM} correlated better with V_T than SUVR_{CB} for all tested regions, except for the anterior cingulate in AD and aMCI and the posterior cingulate in AD (Supplemental Table 2). Correlation coefficients between V_T and SUVR to alternative reference regions were in between those obtained for SUVRCB and $SUVR_{WM}$ (Supplemental Table 3). V_T and SUVR values that were not corrected for partial-volume effects are shown in Supplemental Figure 2. Correlation results of uncorrected data were in line with those of corrected data also revealing that there was a stronger correlation between V_T and $SUVR_{WM}$ than between V_T and $SUVR_{CR}$ (Supplemental Table 4). Group-averaged parametric maps of SUVR (50-60 min after injection) and V_T are presented in Figure 4. There was significant correlation between V_T of the 2TCM and Ichise's multilinear analysis 1 graphical analysis (r = 0.65 for cerebellar gray, r > 0.84 for other ROIs, P < 0.0001). Figure 4B depicts the apparent decrease in tracer uptake from the aMCI toward the AD stage for SUVR_{CB}, thereby confirming our regional-based results.

Regional Blood Flow Results

¹⁵O-water SUV (Figs. 5A and 5B) showed that blood flow was decreased in AD compared with HC and aMCI in cortical regions

Brain region	V _T – SUVR _{CB}	V _T – SUVR _{WM}
Frontal lobe	0.72*	0.82*
Parietal lobe	0.76*	0.85*
Temporal lobe	0.51 [†]	0.70*
Occipital lobe	0.69*	0.79*
Precuneus	0.81*	0.89*
Anterior cingulate	0.79*	0.74*
Posterior cingulate	0.54 [‡]	0.63*

Symbols denote significant correlations (*P < 0.0001, †P = 0.002, †P = 0.0009).

($-11\% \pm 3\%$ and $-5\% \pm 1\%$, respectively) and in the reference regions (CB, $-8\% \pm 12\%$ and $-9\% \pm 8\%$, respectively; WM, $-10\% \pm 12\%$ and $-8\% \pm 8\%$, respectively), albeit nonsignificantly (P > 0.1). Similar results were seen for 18 F-AV45 K_1 derived from the pharmacokinetic modeling procedure (cortex, $-13\% \pm 3\%$ and $-6\% \pm 1\%$, respectively; CB, $-11\% \pm 7\%$ and $-10\% \pm 6\%$, respectively; WM, $-11\% \pm 8\%$ and $-4\% \pm 7\%$, respectively) (Fig. 5C). Correlations between 15 O-water SUV and 18 F-AV45 K_1 were moderate (r ranging from 0.50 for WM and frontal and temporal cortex [P < 0.01] to 0.68 for posterior cingulate [P < 0.0001]), whereas stronger correlations were observed between the relative flow measures 15 O-water SUVR_{CB} and 18 F-AV45 $K_1/K_{1,CB}$ (r ranging from 0.76 for occipital cortex to 0.93 for temporal cortex, P < 0.0001).

DISCUSSION

Because of practical considerations, the SUVR method is preferred over absolute quantification for measuring plaque load in the brain, because it avoids the long dynamic scanning protocols with simultaneous arterial blood sampling and subsequent pharmacokinetic modeling. However, to determine whether SUVR truly reflects $A\beta$ load, it has to be validated against the quantitative gold standard $V_{\rm T}.$

Our metabolite analysis revealed that unchanged ¹⁸F-AV45 rapidly decreased over time with formation of 2 more polar and 1 less polar radiometabolite. The lowering of the extraction yield over time and the increasing plasma-to-blood ratio might be related to the formation of apolar metabolites that bind to plasma proteins. Prior studies described formation of 2 apolar metabolites identified as ¹⁸F-AV160 and ¹⁸F-AV267, but their contribution to the total PET signal was assumed to be minimal (*10*,23).

The regional 18 F-AV45 time—activity curves could be described by the 2TC model with a blood volume fraction component and were used to derive V_T . We found that tracer accumulation in cortical regions was highest for the AD group, intermediate for the aMCI group, and lowest for the HC group. HCs showed tracer accumulation predominantly in the WM.

Recent studies suggested using a WM reference region to quantify longitudinal SUVR changes (12-15). We confirmed for the first time by pharmacokinetic modeling on dynamic cross-sectional data that SUVR_{WM} more closely reflects V_T differences

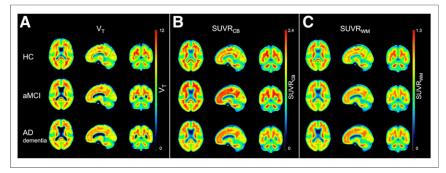


FIGURE 4. Average spatially normalized ^{18}F -AV45 V_T (A), SUVR_{CB} (B), and SUVR_{WM} (C) images for the 3 groups.

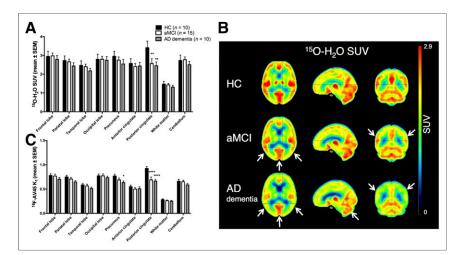


FIGURE 5. (A) Regional ¹⁵O-water SUV (mean \pm SEM) values for the 3 groups. (B) Average spatially normalized ¹⁵O-water SUV images. White arrows indicate regions with reduced flow compared with controls (parietotemporal cortex, posterior cingulate cortex, cerebellum). (C) ¹⁸F-AV45 delivery rate K_1 (mean \pm SEM). Asterisks denote significant differences with HC group (*P < 0.05; **P < 0.01; *****P < 0.0001).

between aMCI and AD and that, across and within all diagnostic groups, it is better correlated with V_T than $SUVR_{CB}$. However, within the HC and AD groups correlations were weaker, which might be primarily due to the wider range of V_T and SUVRs within the aMCI group. Other frequently used reference regions (whole cerebellum, pons, centrum semiovale, and a composite region) resulted in correlation coefficients in between those obtained for $SUVR_{CB}$ and $SUVR_{WM}$.

In line with the literature (16,26,27), flow was reduced in aMCIs compared with controls, notably in more posterior-situated brain regions, including the precuneus and posterior cingulate. The AD group showed an additional flow reduction in the frontal and parietotemporal cortex. As such, higher cortical A β load (i.e., increased SUVR) was related to lower cerebral blood flow. Van Berckel et al. (16) suggested from experimental 11 C-Pittsburgh compound B data and simulations that both global and regional flow changes affected the SUVRs. This effect was such that there was an increase in SUVR when cerebral blood flow was decreased in the target region while stable in the reference region (16,17). From our results, however, it appears that blood flow was also reduced in the reference regions of our AD subjects, whereas there was no significant tracer retention. Because WM regions have a slower tracer uptake and clearance rate than gray matter regions as

well as other binding characteristics, differential flow effects on the SUV of CB and WM might partly explain differences found when using either CB or WM as a reference region for $^{18}\text{F-AV45}$ SUVR. Quantification of flow-induced $^{18}\text{F-AV45}$ SUV changes is important to ensure reliable longitudinal follow-up of A β -modifying treatment effects and is topic of further study.

In this study, we have used V_T for validation of SUVR. Alternatively, the distribution volume ratio, defined as V_T in the target region normalized by V_T in the reference region (serving as an estimate of the nonspecific V_T), could have been considered instead of V_T . However, the distribution volume ratio also critically depends on the choice of a reference region. Another possible limitation is that WM SUV can be decreased by WM lesions (28), thereby increasing the false-positive rate for SUVR_{WM}. In our dataset, however, we did not find any difference between the average WM SUV across the 3 diagnostic groups.

CONCLUSION

This study validated the simplified static SUVR method against the gold standard $V_{\rm T}$ to assess cross-sectional differences in plaque load. $^{18}\text{F-AV45}$ assessed by SUVR does not truly reflect A β load. However, SUVR $_{WM}$ is better correlated with $V_{\rm T}$ and more closely reflects the $V_{\rm T}$ differences between the aMCI and AD group than SUVR $_{CB}$.

DISCLOSURE

This study was funded in part by IWT TGO BIOADAPTAD under grant agreement no. 120835 A14/0161, University Hospital Antwerp, the University of Antwerp Research Fund, the Institute Born-Bunge (IBB, www.bornbunge.be), the Belgian Science Policy Office Interuniversity Attraction Poles (IAP) program (BELSPO, www.belspo.be), the Flemish Government initiated Methusalem excellence grant (EWI, www.ewi-vlaanderen.be), and the Flanders Impulse Program on Networks for Dementia Research (VIND). No other potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

We are thankful to Caroline Berghmans and Philippe Joye (Molecular Imaging Center Antwerp) for support with processing of the blood samples.

REFERENCES

- Hardy JA, Higgins GA. Alzheimer's disease: the amyloid cascade hypothesis. Science. 1992;256:184–185.
- Pike KE, Savage G, Villemagne VL, et al. β-amyloid imaging and memory in non-demented individuals: evidence for preclinical Alzheimer's disease. *Brain*. 2007:130:2837–2844
- Sevigny J, Chiao P, Bussière T, et al. The antibody aducanumab reduces amyloidbeta plaques in Alzheimer's disease. Nature. 2016;537:50–56.

- Timmers M, Van Broeck B, Ramael S, et al. Profiling the dynamics of CSF and plasma Abeta reduction after treatment with JNJ-54861911, a potent oral BACE inhibitor. Alzheimers Dement Transl Res Clin Interv. 2016;2:202–212.
- Forlenza OV, Radanovic M, Talib LL, et al. Cerebrospinal fluid biomarkers in Alzheimer's disease: diagnostic accuracy and prediction of dementia. Alzheimers Dement (Amst). 2015;1:455–463.
- Herholz K, Ebmeier K. Clinical amyloid imaging in Alzheimer's disease. Lancet Neurol. 2011;10:667–670.
- Vandenberghe R, Adamczuk K, Van Laere K. The interest of amyloid PET imaging in the diagnosis of Alzheimer's disease. Curr Opin Neurol. 2013; 26:646-655.
- Klunk WE, Engler H, Nordberg A, et al. Imaging brain amyloid in Alzheimer's disease with Pittsburgh compound-B. Ann Neurol. 2004;55:306–319.
- Price JC, Klunk WE, Lopresti BJ, et al. Kinetic modeling of amyloid binding in humans using PET imaging and Pittsburgh compound-B. J Cereb Blood Flow Metab. 2005;25:1528–1547.
- Choi SR, Golding G, Zhuang Z, et al. Preclinical properties of ¹⁸F-AV-45: a PET agent for Aβ plaques in the brain. J Nucl Med. 2009;50:1887–1894.
- Jagust WJ, Landau SM, Koeppe RA, et al. The Alzheimer's Disease Neuroimaging Initiative 2 PET Core: 2015. Alzheimers Dement. 2015;11:757–771.
- Landau SM, Fero A, Baker SL, et al. Measurement of longitudinal β-amyloid change with ¹⁸F-florbetapir PET and standardized uptake value ratios. *J Nucl Med*. 2015;56:567–574.
- Brendel M, Högenauer M, Delker A, et al. Improved longitudinal [18F]-AV45 amyloid PET by white matter reference and VOI-based partial volume effect correction. *Neuroimage*. 2015;108:450–459.
- Chen K, Roontiva A, Thiyyagura P, et al. Improved power for characterizing longitudinal amyloid-β PET changes and evaluating amyloid-modifying treatments with a cerebral white matter reference region. J Nucl Med. 2015;56:560– 566.
- Shokouhi S, Mckay JW, Baker SL, et al. Reference tissue normalization in longitudinal ¹⁸F-florbetapir positron emission tomography of late mild cognitive impairment. Alzheimers Res Ther. 2016;8:2–12.
- van Berckel BNM, Ossenkoppele R, Tolboom N, et al. Longitudinal amyloid imaging using ¹¹C-PiB: methodologic considerations. J Nucl Med. 2013;54:1570–1576.

- Cselényi Z, Farde L. Quantification of blood flow-dependent component in estimates of beta-amyloid load obtained using quasi-steady-state standardized uptake value ratio. J Cereb Blood Flow Metab. 2015;35:1485–1493.
- Mazza M, Marano G, Traversi G, Bria P, Mazza S. Primary cerebral blood flow deficiency and Alzheimer's disease: shadows and lights. *J Alzheimers Dis.* 2011; 23:375–389.
- Austin BP, Nair VA, Meier TB, et al. Effects of hypoperfusion in Alzheimer's disease. J Alzheimers Dis. 2011;26(suppl 3):123–133.
- Niemantsverdriet E, Feyen B, Le Bastard N, et al. Overdiagnosing vascular dementia using structural brain imaging for dementia work-up. J Alzheimers Dis. 2015;45:1039–1043.
- McKhann GM, Knopman DS, Chertkow H, et al. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. Alzheimers Dement. 2011;7:263–269.
- Yao C-H, Lin K-J, Weng C-C, et al. GMP-compliant automated synthesis of ¹⁸F-AV45 (Florbetapir F 18) for imaging β-amyloid plaques in human brain. *Appl Radiat Isot.* 2010;68:2293–2297.
- Wong DF, Rosenberg PB, Zhou Y, et al. In vivo imaging of amyloid deposition in Alzheimer disease using the radioligand ¹⁸F-AV-45 (florbetapir F-18). J Nucl Med. 2010;51:913–920.
- Bertoldo A, Vicini P, Sambuceti G, Lammertsma AA, Parodi O, Cobelli C. Evaluation of compartmental and spectral analysis models of [¹⁸F]FDG kinetics for heart and brain studies with PET. *IEEE Trans Biomed Eng.* 1998;45:1429–1449.
- Ichise M, Toyama H, Innis RB, Carson RE. Strategies to improve neuroreceptor parameter estimation by linear regression analysis. J Cereb Blood Flow Metab. 2002:22:1271–1281.
- Becker GA, Ichise M, Barthel H, et al. PET quantification of ¹⁸F-florbetaben binding to β-amyloid deposits in human brains. J Nucl Med. 2013;54:723–731.
- Chen YJ, Rosario BL, Mowrey W, et al. Relative ¹¹C-PiB delivery as a proxy of relative CBF: quantitative evaluation using single-session ¹⁵O-water and ¹¹C-PiB PET. J Nucl Med. 2015;56:1199–1205.
- Matías-Guiu JA, Cabrera-Martín MN, Matías-Guiu J, et al. Amyloid PET imaging in multiple sclerosis: an ¹⁸F-florbetaben study. BMC Neurol. 2015;15:243.