# Comparison of the Amino Acid Tracers <sup>18</sup>F-FET and <sup>18</sup>F-DOPA in High-Grade Glioma Patients

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High-grade gliomas (HGGs) are the most common malignant primary tumors of the central nervous system. PET probes of amino acid transport such as O-(2-18F-fluoroethyl)-L-tyrosine (18F-FET), 3,4dihydroxy-6-18F-fluoro-L-phenylalanine (18F-DOPA), and 11C-methionine (11C-MET) detect primary and recurrent tumors with a high accuracy. <sup>18</sup>F-FET is predominantly used in Europe, whereas amino acid transport imaging is infrequently done in the United States. The aim of this study was to determine whether <sup>18</sup>F-FET and <sup>18</sup>F-DOPA PET/CT provide comparable information in HGG. Methods: Thirty <sup>18</sup>F-FET and <sup>18</sup>F-DOPA PET/CT scans were obtained before surgery or biopsy in 27 patients with high clinical suspicion for primary or recurrent HGG (5 primary, 22 recurrent tumors). <sup>18</sup>F-FET and <sup>18</sup>F-DOPA PET/CT images were compared visually and semiquantitatively (maximum standardized uptake value [SUV<sub>max</sub>], mean SUV [SUV<sub>mean</sub>]). Background (SUV<sub>max</sub> and SUV<sub>mean</sub>) and tumor-to-background ratios (TBRs) were calculated for both PET probes. The degree of  $^{18}\text{F-DOPA}$  uptake in the basal ganglia (SUV $_{\text{mean}}$ ) was also assessed. Results: Visual analysis revealed no difference in tumor uptake pattern between the 2 PET probes. The  $SUV_{mean}$  and  $SUV_{max}$ for  $^{18}\text{F-FET}$  were higher than those of  $^{18}\text{F-DOPA}$  (4.0  $\pm$  2.0 and 4.9  $\pm$ 2.3 vs. 3.5  $\pm$  1.6 and 4.3  $\pm$  2.0, respectively; all P < 0.001). TBRs for  $SUV_{mean}$  but not for  $SUV_{max}$  were significantly higher for  $^{18}F\text{-FET}$ than  $^{18}\text{F-DOPA}$  (TBR SUV<sub>mean</sub>: 3.8 ± 1.7 vs. 3.4 ± 1.2, P = 0.004; TBR SUV<sub>max</sub>:  $3.3 \pm 1.6$  and  $3.0 \pm 1.1$ , respectively; P = 0.086). <sup>18</sup>F-DOPA uptake by the basal ganglia was present (SUV<sub>mean</sub>, 2.6 ± 0.7) but did not affect tumor visualization. Conclusion: Whereas visual analysis revealed no significant differences in uptake pattern for <sup>18</sup>F-FET and <sup>18</sup>F-DOPA in patients with primary or recurrent HGG, both SUVs and TBRs for SUV<sub>mean</sub> were significantly higher for <sup>18</sup>F-FET. However, regarding tumor delineation, both tracers performed equally well and seem equally feasible for imaging of primary and recurrent HGG. These findings suggest that both PET probes can be used based on availability in multicenter trials.

**Key Words:** glioblastoma; positron emission tomography; PET; amino acids; FET; F-DOPA

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igh-grade gliomas (HGGs) are the most common malignant primary tumors of the central nervous system, accounting for 45%–50% of all gliomas with an incidence of about 3–4 per 100,000 inhabitants per year (1). Despite multimodality treatment approaches, 5-y survival is abysmal at 5% (2).

Patients are followed clinically for neurologic symptoms and through neuroimaging with MR imaging, the current clinical gold standard. One major diagnostic problem is the differentiation between contrast enhancement due to radiation necrosis (so-called pseudoprogression) and tumor recurrence (3). Nonspecific contrast enhancement also complicates therapy monitoring (4).

PET/CT with various probes of glucose metabolism and amino acid transport can contribute to improved posttreatment assessment (5). <sup>18</sup>F-FDG PET was first used for imaging brain tumors (6,7). However, later studies demonstrated limitations mainly due to the high normal gray matter <sup>18</sup>F-FDG activity (7,8). As an alternative, labeled amino acid analogs have been introduced (9,10). Among them, <sup>11</sup>C-methionine (<sup>11</sup>C-MET) has been studied most extensively (11). However, because of the short physical half-life of <sup>11</sup>C of only 20 min, the use of <sup>11</sup>C-MET PET has been limited to PET centers with access to an on-site cyclotron. To overcome this limitation, several <sup>18</sup>F-labeled amino acids have been introduced (10,12). 3,4-dihydroxy-6-<sup>18</sup>F-fluoro-L-phenylalanine (<sup>18</sup>F-DOPA) has been used for imaging brain tumors (13–16), neuroendocrine tumors (17), and movement disorders (18) for more than 20 y. However, the synthesis of <sup>18</sup>F-DOPA is laborious, and its availability is limited.

More recently, O-(2-<sup>18</sup>F-fluoroethyl)-L-tyrosine (<sup>18</sup>F-FET) has been introduced for PET brain tumor imaging (19). Similar to <sup>18</sup>F-DOPA, it can detect low-grade gliomas and HGGs (20–22). As an advantage, its radiosynthesis is straightforward, and it can thus be delivered via commercial radiopharmacies. Therefore, <sup>18</sup>F-FET has become the most commonly used radiotracer for brain tumor imaging with PET in Europe.

However, <sup>18</sup>F-FET is not approved by the Food and Drug administration for brain tumor imaging in the United States. Few sites in the United States are using <sup>18</sup>F-DOPA for brain imaging under an Investigational New Drug application.

Clinical studies have demonstrated a comparable accuracy for <sup>11</sup>C-MET PET and <sup>18</sup>F-FET or <sup>18</sup>F-DOPA PET in detecting primary and recurrent brain tumors (*13,20*). However, only 1 study has compared <sup>18</sup>F-FET with <sup>18</sup>F-DOPA (*23*). Such comparisons are important because comparable diagnostic information would greatly

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facilitate multicenter studies, which could include patients studied with one or the other PET probe of amino acid transport. The aim of this prospective study was therefore to determine whether <sup>18</sup>F-DOPA and <sup>18</sup>F-FET provide comparable diagnostic information visually and semiquantitatively.

## **MATERIALS AND METHODS**

#### Subjects

This prospective clinical study included patients with suspected primary or recurrent HGGs. The patients' Karnofsky score had to be at least 80% when they entered the study. All patients had to be able to understand the study procedures and to provide informed consent. The study adhered to the standards established in the Declaration of Helsinki. This study was reviewed by the Institutional Review Board of the Medical Faculty of the University of Würzburg (local ethics committee), and all patients signed a written informed consent form.

From August 2012 to December 2013, 27 patients (18 men and 9 women; mean age  $\pm$  SD, 54  $\pm$  14 y; range, 25–80 y) were enrolled. Five patients presented with suspected primary HGG, whereas 22 patients had suspected tumor recurrence (Table 1). Three patients were investigated twice, leading to a total of 30 pairs of  $^{18}\text{F-FET}$  and  $^{18}\text{F-DOPA}$  PET/CT scans.

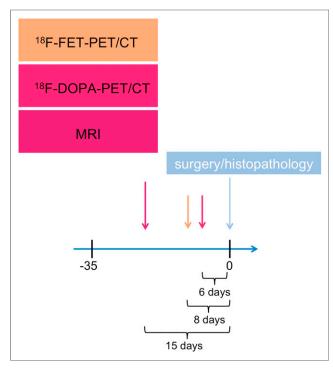
All patients with suspected recurrence (n=22) had undergone surgical resection. In addition to surgery, 20 of 22 patients had received chemotherapy and radiation (91%), whereas 2 of 22 patients had undergone radiation only (9%). The time from completion of radiation to the PET scan was more than 12 wk in all subjects. Recurrent disease was verified by tumor biopsy or surgery. All patients with primary tumors underwent tumor surgery.

MR imaging was done within 5 wk before surgery or tumor biopsy (mean, 15 d; median, 14 d; range, 1–35 d). <sup>18</sup>F-FET PET scans were obtained within a mean interval of 8 d (median, 6 d; range, 1–22 d), <sup>18</sup>F-DOPA PET scans within a mean interval of 6 d (median, 3 d; range, 1–18 d) before surgery. Histopathology served as a gold standard in all patients

**TABLE 1**Patients' Characteristics

| Patient no. | Age (y) | Sex | Primary/recurrent<br>HGG | Histology                  | Lesion site            | Previous therapy           |  |
|-------------|---------|-----|--------------------------|----------------------------|------------------------|----------------------------|--|
| 1           | 47      | М   | R                        | Primary GBM                | Left temporal          | Surgery, radiochemotherapy |  |
| 2           | 45      | F   | R                        | Secondary GBM              | Multifocal             | Surgery, radiochemotherapy |  |
| 3           | 49      | М   | R                        | Primary GBM                | Left frontal           | Surgery, radiochemotherapy |  |
| 4           | 50      | М   | R                        | Primary GBM                | Multifocal             | Surgery, radiochemotherapy |  |
| 5           | 73      | М   | R                        | Primary GBM                | Left frontal           | Surgery, radiochemotherapy |  |
| 6           | 42      | М   | R                        | Primary GBM                | Right parietooccipital | Surgery, radiochemotherapy |  |
| 7           | 43      | М   | R                        | Primary GBM                | Left parietal          | Surgery, radiochemotherapy |  |
| 8           | 41      | F   | R                        | Anaplastic astrocytoma III | Left temporal          | Surgery, radiochemotherapy |  |
| 9           | 44      | М   | R                        | Anaplastic astrocytoma III | Right temporal         | Surgery, radiochemotherapy |  |
| 10          | 70      | F   | R                        | Primary GBM                | Left temporal          | Surgery, radiochemotherapy |  |
| 11          | 51      | F   | R                        | Primary GBM                | Left parietal          | Surgery, radiochemotherapy |  |
| 12          | 60      | М   | R                        | Primary GBM                | Left temporal          | Surgery, radiochemotherapy |  |
| 13          | 67      | М   | R                        | Primary GBM                | Left frontal           | Surgery, radiochemotherapy |  |
| 14          | 55      | М   | R                        | Primary GBM                | Left temporal          | Surgery, radiochemotherapy |  |
| 15          | 54      | F   | R                        | Primary GBM                | Right cerebellar       | Surgery, radiochemotherapy |  |
| 16          | 33      | М   | R                        | Oligoastrocytoma III       | Left frontal           | Surgery, radiotherapy      |  |
| 17          | 57      | М   | R                        | Primary GBM                | Right parietal         | Surgery, radiochemotherapy |  |
| 18          | 58      | М   | R                        | Primary GBM                | Right temporooccipital | Surgery, radiochemotherapy |  |
| 19          | 61      | М   | R                        | Primary GBM                | Right parietal         | Surgery, radiochemotherapy |  |
| 14/2        | 55      | М   | R                        | Primary GBM                | Left parietal          | Surgery, radiochemotherapy |  |
| 17/2        | 55      | М   | R                        | Primary GBM                | Right parietal         | Surgery, radiochemotherapy |  |
| 20          | 25      | М   | R                        | Secondary GBM              | Right temporal         | Surgery, radiotherapy      |  |
| 21          | 75      | М   | R                        | Primary GBM                | Right temporal         | Surgery, radiochemotherapy |  |
| 10/2        | 71      | F   | R                        | Primary GBM                | Left temporal          | Surgery, radiochemotherapy |  |
| 22          | 33      | М   | R                        | Primary GBM                | Right frontal          | Surgery, radiochemotherapy |  |
| 23          | 77      | F   | Р                        | Primary GBM                | Right frontal          | None                       |  |
| 24          | 80      | F   | Р                        | Primary GBM                | Left parietooccipital  | None                       |  |
| 25          | 51      | М   | Р                        | Primary GBM                | Right frontal          | None                       |  |
| 26          | 65      | F   | Р                        | Primary GBM                | Left temporal          | None                       |  |
| 27          | 40      | F   | Р                        | Pilocytic astrocytoma i    | Right basal ganglia    | None                       |  |

10/2; 14/2; 17/2 = 3 patients were imaged twice.



**FIGURE 1.** Schematic outline of study. Within 35 d before tumor surgery or biopsy, MR imaging (mean, 15 d prior) and <sup>18</sup>F-FET PET/CT (mean, 8 d) and <sup>18</sup>F-DOPA PET/CT (mean, 6 d) were performed. Histopathology served as gold standard. Mean intervals to surgery are noted in time scale.

to verify diagnosis and the presence of viable tumor tissue. The study design is shown in Figure 1.

# Tracer Synthesis and PET/CT

 $^{18}\text{F-FET}$  was synthesized in-house on a TRACERlab FX-FN synthesis module (GE Healthcare) as previously described (23).  $^{18}\text{F-FET}$  was obtained at a 37%  $\pm$  5% radiochemical yield (non–decay-corrected), with a radiochemical purity of greater than 99%, as assessed by high-performance liquid chromatography and thin-layer chromatography analysis.  $^{18}\text{F-DOPA}$  was purchased from IASON GmbH.

PET was performed on a dedicated PET/CT scanner (Biograph mCT 64; Siemens).  $^{18}\text{F-DOPA}$  (175  $\pm$  39 MBq) and  $^{18}\text{F-FET}$  (217  $\pm$  13 MBq) were injected intravenously. CT scans for attenuation correction were acquired after 15 min ( $^{18}\text{F-DOPA}$ ) and 10 min ( $^{18}\text{F-FET}$ ), respectively, using a low-dose protocol (80 mAs, 120 kV, a 512  $\times$ 512 matrix, 5-mm slice thickness, increment of 30 mm/s, rotation time of 0.5 s, and pitch index of 0.8). PET emission data were acquired in 3-dimensional mode with a 200  $\times$  200 matrix for 20 min ( $^{18}\text{F-DOPA}$ ) and 10 min ( $^{18}\text{F-FET}$ ), respectively. After decay and scatter correction, PET data were reconstructed iteratively with attenuation correction using dedicated software (Esoft; Siemens).

#### **Image Analysis**

Images were analyzed as described by Fueger et al. (15). In brief, images were first inspected visually. Then the axial PET image slice displaying the maximum tumor uptake was selected. Tumor regions of interest (ROIs) were defined in 2 ways. First, a standardized 10-mm circular region was placed over the area with the peak activity. This first ROI was used to derive maximum standardized uptake values (SUV $_{max}$ ) and mean SUVs (SUV $_{mean}$ ). A normal reference brain region was defined by drawing an ROI (diameter, 25 mm) involving the entire contralateral hemisphere at the level of the centrum semiovale to derive tumor-to-background ratios. For  $^{18}$ F-DOPA PET, the basal ganglia were assessed by an additional ROI (diameter, 10 mm) in the contralateral striatum. The

radiotracer concentration in the ROIs was normalized to the injected dose per kilogram of patient's body weight to derive the SUVs.

#### Histopathology and Immunohistochemistry

The biopsy samples were formalin-fixed and paraffin-embedded. All tumor samples were histologically assessed and graded using standard hematoxylin and eosin sections (3–4  $\mu m$ ) according to the criteria of the World Health Organization (24). The astrocytic origin of the tumors was confirmed by positive immunoreaction for the glial fibrillary acidic protein (1:200; Dako). Oligodendroglial features were assured by the distinct pattern of microtubule-associated protein 2 immunoreactivity (1:250; Dako). In addition, to determine the proliferation activity of each tumor, a labeling index (%) was calculated after immunostaining for MIB-1 (Ki67 1:50; Dako) by determining the number of positive nuclei among 100 tumor cells per high power field (HPF) in 10 HPFs.

## Statistical Analysis

Quantitative data are presented as median, range, and mean  $\pm$  SD. The Wilcoxon signed-rank test and the Mann–Whitney test were used for paired and unpaired comparisons of quantitative parameters. Corresponding accuracies for recurrence detection were calculated using histopathology and clinical follow-up for validation. The  $\chi^2$  or Fisher exact test was conducted for comparison of frequency data between independent subgroups. Receiver-operating-characteristic curves were used to determine optimal cutoff values for defining disease recurrence and for the prediction of survival. The Fisher exact test was used to assess the association of 2 categoric variables.

Statistical analyses were performed using SPSS Statistics software for Windows (version 22.0; IBM, SPSS Inc.). All statistical tests were performed 2-sided, and a *P* value of less than 0.05 was considered to indicate statistical significance. No correction of *P* values was applied to adjust for multiple tests (25).

## **RESULTS**

#### **Tumor Characteristics**

In 26 of 27 patients (96%), the brain lesions were located supratentorially. Ten subjects (37%) presented with temporal (temporooccipital) lesions, 7 (26%) with frontal lesions, 6 (22%) had parietal (parietooccipital) tumors, and 2 (7%) had multifocal disease. In 1 patient, the glioma involved the basal ganglia. The only infratentorial HGG was located in the cerebellum.

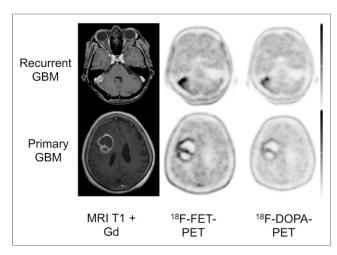
Final diagnosis was established by histopathology of surgical or biopsy samples. Seventeen of 22 patients with recurrent disease had glioblastoma multiforme (GBM). In 2 patients, a GBM had evolved from grade III oligodendroglioma. Two patients had an anaplastic astrocytoma, whereas 1 patient suffered from a grade III oligoastrocytoma.

In 4 of 5 patients with suspected primary GBM, the clinical diagnosis was confirmed by histopathology. The remaining patient had a pilocytic grade I astrocytoma.

# **Visual PET Image Analysis**

All primary and recurrent HGGs showed enhanced <sup>18</sup>F-DOPA and <sup>18</sup>F-FET uptake (sensitivity, 100%). All lesions were clearly delineated from normal brain tissue. Overall, the 2 tracers showed a matching image pattern on visual assessment (Fig. 2).

The tumor distribution of both tracers was homogeneous and comparable. In the patients with multifocal glioblastoma both <sup>18</sup>F-DOPA and <sup>18</sup>F-FET depicted all lesions (Fig. 3). Visually, individual differences in uptake intensity occurred, without a tracer preference; however, these were not of relevance for image interpretation. Despite significant basal ganglia <sup>18</sup>F-DOPA uptake



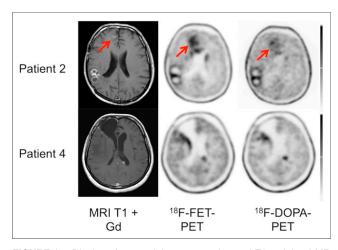
**FIGURE 2.** Display of transaxial contrast-enhanced T1-weighted MR imaging,  $^{18}\text{F-FET}$ , and  $^{18}\text{F-DOPA}$  PET/CT scans of patient with recurrent GBM and primary GBM. In both patients,  $^{18}\text{F-FET}$  uptake (SUV<sub>max</sub> and SUV<sub>mean</sub>, 6.1 and 5.6, respectively) was higher than that of  $^{18}\text{F-DOPA}$  (SUV<sub>max</sub> and SUV<sub>mean</sub>, 4.4 and 2.5, respectively).

(SUV  $_{\rm mean}$ , 2.6  $\pm$  0.7), tumor visualization was possible in all patients (Fig. 4).

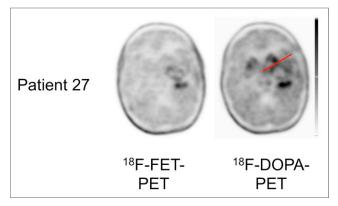
# Semiquantitative Image Analysis

In newly diagnosed and recurrent HGG, visual analysis of both tracers revealed no difference in tumor uptake pattern. However, differences in semiquantitative parameters existed. The SUV<sub>mean</sub> and SUV<sub>max</sub> for <sup>18</sup>F-FET was higher than that of <sup>18</sup>F-DOPA (4.0  $\pm$  2.0 and 4.9  $\pm$  2.3 vs. 3.5  $\pm$  1.6 and 4.3  $\pm$  2.0, respectively; both P < 0.001).

Background SUV<sub>mean</sub> and SUV<sub>max</sub> were similar for <sup>18</sup>F-FET and <sup>18</sup>F-DOPA ( $1.0 \pm 0.2$  and  $1.5 \pm 0.3$  vs.  $1.0 \pm 0.3$  and  $1.4 \pm 0.3$ ; P = not significant). <sup>18</sup>F-FET and <sup>18</sup>F-DOPA SUV<sub>mean</sub> and SUV<sub>max</sub> were closely correlated (r = 0.943; P < 0.0001). Consistently, TBR for SUV<sub>mean</sub> and SUV<sub>max</sub> were higher for <sup>18</sup>F-FET than for <sup>18</sup>F-DOPA (SUV<sub>mean</sub>,  $3.8 \pm 1.7$  vs.  $3.4 \pm 1.2$ ; SUV<sub>max</sub>,  $3.3 \pm 1.6$  and  $3.0 \pm 1.1$ , P = 0.004 and 0.086, respectively) (Table 2).



**FIGURE 3.** Display of transaxial contrast-enhanced T1-weighted MR imaging, <sup>18</sup>F-FET, and <sup>18</sup>F-DOPA PET/CT scans of 2 patients with multifocal GBM. In both patients, <sup>18</sup>F-FET and <sup>18</sup>F-DOPA depicted all lesions.



**FIGURE 4.** Display of transaxial <sup>18</sup>F-FET and <sup>18</sup>F-DOPA PET/CT scans of patient with primary glioma affecting basal ganglia. Striatal uptake does not significantly compromise tumor delineation.

Corresponding SUV<sub>max</sub>, SUV<sub>mean</sub>, and tumor-to-background values for <sup>18</sup>F-DOPA and <sup>18</sup>F-FET are shown for both subgroups (newly diagnosed and recurrent HGG) in Table 2.

#### DISCUSSION

On the basis of previous studies that have demonstrated a high accuracy of <sup>18</sup>F-DOPA imaging for detecting primary and recurrent brain tumors and a significant impact on patient management (26,27), we have recently initiated a randomized multicenter trial to determine the impact of <sup>18</sup>F-DOPA imaging on patient outcome by randomizing patients with suspected HGG recurrence into those who are managed using conventional diagnostic imaging versus those who will receive conventional imaging plus <sup>18</sup>F-DOPA PET (28). However, only a few sites in the United States offer <sup>18</sup>F-DOPA brain tumor imaging whereas European clinics more frequently provide <sup>18</sup>F-FET services. Thus, enrollment into the trial would be greatly facilitated if both <sup>18</sup>F-FET and <sup>18</sup>F-DOPA PET provide comparable diagnostic information and could thus be used for this trial. Before modifying the trial, a direct comparison of these 2 PET probes is, however, needed.

<sup>18</sup>F-labeled amino acids generally share the L-type amino acid transporters LAT1 and LAT2 (*29–31*). We therefore expected a similar distribution of these probes in HGG. By visual analysis, tracer distribution was indeed comparable, and both probes provided near-identical information. Differences in tumor-to-background ratios were negligible. Thus, both imaging probes provided concordant information.

By semiquantitative analysis,  $SUV_{mean}$  and  $SUV_{max}$  and TBRof SUV<sub>mean</sub> were significantly higher for <sup>18</sup>F-FET than <sup>18</sup>F-DOPA. This contrasts a recent report (23), in which quantitative <sup>18</sup>F-DOPA and <sup>18</sup>F-FET PET were compared in 8 patients with recurrent lowgrade astrocytoma and 8 patients with high-grade glioblastoma. <sup>18</sup>F-DOPA demonstrated superior contrast ratios for lesions outside the striatum (23). The authors argued that <sup>18</sup>F-DOPA uptake occurs via both the LAT1 and the LAT2 systems, whereas <sup>18</sup>F-FET is mainly transported by LAT2 (29,32). However, the in vivo relevance of this phenomenon for the pharmacokinetics of <sup>18</sup>F-DOPA versus those of <sup>18</sup>F-FET remains unknown (33). Additionally, the transport mechanism of <sup>18</sup>F-FET may be more complex. Although increased uptake of <sup>18</sup>F-FET was observed in gliomas and squamous cell carcinomas, no uptake of <sup>18</sup>F-FET was found in many extracranial tumors, especially in lymphomas and adenocarcinomas (34). This finding is in contrast to other tyrosine derivatives such as L-11C-tyrosine, 2-18F-fluoro-L-tyrosine, and

**TABLE 2**Semiquantitative Analysis of <sup>18</sup>F-DOPA and <sup>18</sup>F-FET

| Parameter           | <sup>18</sup> F-FET<br>SUV <sub>mean</sub> | <sup>18</sup> F-DOPA<br>SUV <sub>mean</sub> | <sup>18</sup> F-FET<br>SUV <sub>max</sub> | <sup>18</sup> F-DOPA<br>SUV <sub>max</sub> | <sup>18</sup> F-FET TBR<br>SUV <sub>mean</sub> | <sup>18</sup> F-DOPA TBR<br>SUV <sub>mean</sub> | <sup>18</sup> F-FET TBR<br>SUV <sub>max</sub> | <sup>18</sup> F-DOPA<br>TBR SUV <sub>max</sub> |  |  |
|---------------------|--|---|---|--|--|---|---|--|--|--|
| All                 |  |   |   |  |  |   |   |  |  |  |
| Mean                | 4.0  | 3.5   | 4.9                                       | 4.3  | 3.8  | 3.4   | 3.3   | 3.0  |  |  |
| SD                  | 2.0  | 1.6   | 2.3                                       | 2.0  | 1.7  | 1.2   | 1.6   | 1.1  |  |  |
| Range               | 2.0-11.9                                   | 2.0-9.2                                     | 2.2-13.3                                  | 2.3-10.3                                   | 2.0-10.8                                       | 2.1-8.4   | 1.8–9.5                                       | 2.0-6.9  |  |  |
| Р                   | <0   | .001  | < 0.001                                   |  | 0.004  |   | 0.086   |  |  |  |
| Newly diagnosed HGG |  |   |   |  |  |   |   |  |  |  |
| Mean                | 4.4  | 3.5   | 5.6                                       | 4.7  | 4.3  | 3.6   | 3.6   | 3.3  |  |  |
| SD                  | 1.4  | 1.7   | 2.0                                       | 2.7  | 1.4  | 1.2   | 1.3   | 1.3  |  |  |
| Range               | 3.2-6.8                                    | 2.5-6.5                                     | 4.0-9.0                                   | 2.9-9.4                                    | 2.9-6.2  | 2.6-5.4   | 2.7-6.0                                       | 2.2-5.5  |  |  |
| Р                   | 0.043                                      |   | 0.08                                      |  | 0.043  |   | 0.225   |  |  |  |
| Recurrent HGG       |  |   |   |  |  |   |   |  |  |  |
| Mean                | 3.9  | 3.5   | 4.7                                       | 4.2  | 3.7  | 3.4   | 3.2   | 3.0  |  |  |
| SD                  | 2–1  | 1.7   | 2.4                                       | 1.8  | 1.8  | 1.2   | 1.6   | 1.0  |  |  |
| Range               | 2.0-11.9                                   | 2.0-9.2                                     | 2.2-13.3                                  | 2.3-10.3                                   | 2.0-10.8                                       | 2.1-8.4   | 1.8–9.5                                       | 2.0-6.9  |  |  |
| Р                   | 0.008                                      |   | 0.003                                     |  | 0.034  |   | 0.201   |  |  |  |

Respective SUV<sub>max</sub> and SUV<sub>mean</sub> and tumor-to-background values for <sup>18</sup>F-DOPA and <sup>18</sup>F-FET for entire cohort and both subgroups (newly diagnosed and recurrent HGG).

L-3- $^{18}$ F-fluoro- $\alpha$ -methyltyrosine. As a possible explanation,  $^{18}$ F-FET may be taken up via a specific subtype of system L other than LAT1.

Transstimulation experiments in *Xenopus laevis* oocytes expressing LAT1 indicated that <sup>18</sup>F-FET influx via LAT1 was poor (30). It is tempting to speculate that <sup>18</sup>F-FET is selectively transported by LAT2. <sup>18</sup>F-FET transport in F98 glioma cells, however, is shared by serine, which is a substrate of LAT2 but not of LAT1 (20). At the same time, <sup>18</sup>F-FET shows no uptake in inflammatory tissue where LAT2 is not expressed (31,35). Therefore, it remains unclear whether the different results in both papers can be attributed to the particular tracer uptake or if they were caused by different study methodology.

A more likely explanation for the conflicting data may be the rather small patient sample size of the Heidelberg group, which only included 8 subjects with HGG. Thus, larger studies are needed to elucidate the subtle differences in the kinetics of the 2 PET probes.

The lower <sup>18</sup>F-DOPA SUV found in the current study may also be explained by peripheral aromatic amino acid decarboxylase activity that reduces the amount of tracer available for transport into tumor cells. Carbidopa administration might have led to higher tumor uptake. However, the effect of carbidopa administration for brain tumor imaging is still controversial (36–38).

The current study has several limitations. Because this study focused on HGG, patients with low-grade gliomas were not included. Because of logistic reasons, we could not enroll more than 5 patients with suspected primary HGG. However, the ongoing multicenter trial involves only patients with suspected brain tumor recurrence. Dynamic evaluation of <sup>18</sup>F-FET PET has shown high diagnostic power in tumor grading in untreated and recurrent tumors (*39,40*). We did not perform dynamic acquisitions to further investigate tracer kinetics. This approach was chosen to make the multicenter trial more practical. Moreover, the slightly different start of emission images (15 vs. 10 min for <sup>18</sup>F-DOPA and <sup>18</sup>F-FET) and the different

scan duration (20 vs. 10 min for <sup>18</sup>F-DOPA and <sup>18</sup>F-FET) have to be mentioned. Ideally, ROI definition would have been performed on MR images and then transferred to the PET images; however, because of heterogeneous MR acquisition protocols this was impossible. ROI definition for each tracer was done according to the corresponding hottest pixel to secure a high interobserver reproducibility, but also reanalysis defining the ROIs for both tracers in the same slice after aligning both PET images confirmed our findings.

# CONCLUSION

<sup>18</sup>F-FET showed higher SUVs and TBR for SUV<sub>mean</sub> in HGG than <sup>18</sup>F-DOPA if static imaging protocols were used, which are best suited for centers with a heavy daily workflow. <sup>18</sup>F-FET showed higher SUVs and TBR for SUV<sub>mean</sub> in HGG than <sup>18</sup>F-DOPA. However, both tracers depicted all primary and recurrent brain tumors. Given the similar tumor tracer distribution and excellent correlation between <sup>18</sup>F-FET and <sup>18</sup>F-DOPA SUVs, we conclude that both PET probes of amino acid transport can be used in trials and clinical routine.

# **DISCLOSURE**

The costs of publication of this article were defrayed in part by the payment of page charges. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734. No potential conflict of interest relevant to this article was reported.

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