

# The Role of Radiotracer Imaging in the Diagnosis and Management of Patients with Breast Cancer: Part 2—Response to Therapy, Other Indications, and Future Directions\*

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Breast cancer is the most common non-skin type of cancer and the second leading cause of cancer mortality in women. Advances in diagnosis and treatment have led to declines in mortality, despite an increase in breast cancer incidence. An advancing array of both local and systemic therapy options has led to increasingly individualized treatment. Imaging plays a key role in detecting breast cancer and directing its therapy. This continuing education article provides a comprehensive review of current and future radiotracer imaging methods applied to breast cancer, in the context of breast cancer management strategies and other nonnuclear imaging methods. Part 1 of the review provided an overview of clinical and biologic considerations in breast cancer and covered radionuclide imaging for detection and staging. Part 2 covers radionuclide imaging of breast cancer response to therapy, other clinical indications for radionuclide breast cancer imaging, and future directions, including molecular imaging.

**Key Words:** breast cancer; response to therapy; continuing education; molecular imaging; radionuclide imaging

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**B**reast cancer is the most common non-skin type of cancer and the second leading cause of cancer mortality in women (1). Although its incidence continues to rise, mortality has declined over the past several years (1,2). The decline has been attributed to both early diagnosis and more effective treatment (2). Advances in molecular cancer biology have led to an increased understanding of the biologic factors that

contribute to breast cancer pathogenesis and progression. This understanding has already led to new and effective treatments (3), and new targeted therapies continue to be developed and tested (4). Advances in breast cancer molecular biology have also yielded improvements in diagnosis through molecular pathology and molecular imaging (5,6).

Part 1 of this review provided an overview of breast cancer clinical features and biology and reviewed radionuclide imaging for breast cancer detection and staging. Part 2 reviews applications to therapeutic monitoring and other indications such as toxicity monitoring, followed by highlights of future directions, including that of molecular imaging.

## RESPONSE TO THERAPY

### Overview

A wide range of systemic therapy options for breast cancer exists, and breast cancer is one of the more responsive solid tumors (7). Therefore, the evaluation of response to therapy is an important diagnostic need for breast cancer. Systemic therapy is given in 1 of the 3 following clinical settings: adjuvant (after completion of definitive locoregional surgery to treat residual or metastatic microscopic disease), neoadjuvant (primary systemic therapy given before definitive local surgery), or metastatic (systemic therapy of stage IV disease).

For adjuvant therapy, there should be no residual macroscopic disease after surgery and, therefore, imaging is not helpful for response monitoring, except for restaging when recurrence is suspected. In the neoadjuvant and metastatic settings, imaging plays an important role in evaluating the success of therapy. Size-based criteria using anatomic imaging (mammography or ultrasound in the breast, CT or MRI elsewhere) have traditionally been used to measure response but have some significant limitations in both the neoadjuvant and the metastatic settings (8,9). Functional imaging overcomes many of these limitations and may, therefore, play an increasingly significant role in breast cancer response evaluation.

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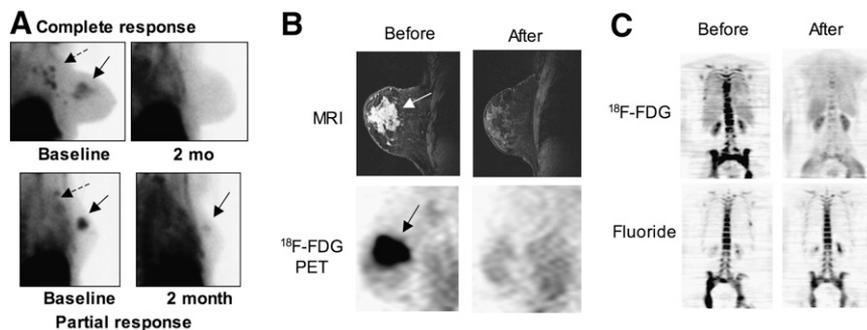
**Neoadjuvant Therapy.** Neoadjuvant (preoperative) systemic therapy has become the standard treatment for patients with locally advanced breast cancer, which is defined as primary breast cancer exceeding 5 cm, fixed axillary lymph nodes, or skin or chest wall invasion (10). Neoadjuvant therapy is increasingly used in patients with operable breast cancer (11). Most neoadjuvant therapy studies have tested cytotoxic chemotherapy regimens; however, neoadjuvant endocrine therapy has been studied (12), and the addition of agents such as trastuzumab to chemotherapy regimens for HER2-overexpressing tumors has increased response rates considerably in appropriately selected patients (13). Although neoadjuvant therapy, compared with similar adjuvant therapy, has not been shown to improve survival it does improve surgical options and provide prognostic information (11). Studies have demonstrated that the extent of residual breast and axillary disease after treatment is prognostic for both disease-free survival and overall survival (8,14,15). Patients demonstrating complete pathologic response (pCR) (defined as no residual invasive tumor on histopathology at posttherapy surgery), compared with patients without pCR, have improved long-term outcome (8,15). One of the primary aims of neoadjuvant therapy is, therefore, to assess the response of the primary tumor to the treatment regimen (16). Sized-based approaches for response evaluation, for example, physical examination and mammography, have not performed well in this setting and have trouble distinguishing pCR from other responses (8,9). This is, therefore, a role in which functional imaging may be particularly helpful.

Studies of  $^{18}\text{F}$ -FDG PET comprise the largest body of work on radiotracer imaging for response evaluation to neoadjuvant breast cancer treatment (17–19). Some of the earliest work using  $^{18}\text{F}$ -FDG to monitor cancer response was done in this clinical setting (20,21). Serial  $^{18}\text{F}$ -FDG PET has been widely studied as a method for assessing tumor response

to neoadjuvant chemotherapy, using a comparison to histopathology assessment of response from the postsurgery specimen as the gold standard (17,22). In these studies,  $^{18}\text{F}$ -FDG PET scans were obtained before therapy and then at 2 or more time points (early [after a single cycle of therapy], mid-therapy, and after therapy) during the course of neoadjuvant therapy (Fig. 1). The pretherapy scan serves as the baseline to assess future changes in the level of  $^{18}\text{F}$ -FDG uptake but is also important in defining the extent of disease that may affect postsurgical treatment, such as radiation therapy, by demonstrating occult nodal or distant metastatic disease (23,24).

Most studies evaluating  $^{18}\text{F}$ -FDG PET to assess response to neoadjuvant therapy have measured change in  $^{18}\text{F}$ -FDG uptake at mid-therapy, compared with at baseline, as a measure of response. The results of these studies are summarized in Table 1. One of the earliest studies of  $^{18}\text{F}$ -FDG PET to measure response was performed by Wahl et al. (21) and showed significant quantitative differences in the  $^{18}\text{F}$ -FDG uptake measured before and after 2 mo of therapy for responders versus nonresponders. Almost all of the subsequent studies reported similar findings and found that a primary tumor  $^{18}\text{F}$ -FDG uptake decline by approximately 50% or more was predictive of a good response (17,22). Perhaps more important, lesser declines in  $^{18}\text{F}$ -FDG uptake predicted poor response.

Studies evaluating change in  $^{18}\text{F}$ -FDG uptake early in the course of therapy suggest that early assessment of response is possible and predictive of subsequent pathologic response (Table 1). Three early studies measuring  $^{18}\text{F}$ -FDG uptake after the first cycle of therapy all demonstrated that early  $^{18}\text{F}$ -FDG PET predicted final response. In 2 of these, the data suggest that early  $^{18}\text{F}$ -FDG PET may demonstrate greater separation between responders and nonresponders than would mid-therapy imaging (21,25,26). More recently, Rousseau et al. (27) found that, using a 60% decrease in



**FIGURE 1.** Examples of radiotracer imaging to measure breast cancer response. (A) Prone lateral  $^{99\text{m}}\text{Tc}$ -sestamibi images of patients with locally advanced breast cancer obtained before and after neoadjuvant chemotherapy (left and right columns) for 2 patients (top and bottom row). Primary tumor (solid arrow) and nodal metastases (dashed arrow) are seen. Top patient demonstrated pCR, and bottom patient demonstrated clinical response but had residual tumor at postchemotherapy surgery. (B) Contrast-

enhanced breast MR (top) and sagittal  $^{18}\text{F}$ -FDG PET images (bottom) of patient with locally advanced breast cancer are shown taken before and after neoadjuvant chemotherapy (left and right columns), focusing on primary tumor (arrow). Patient demonstrated pCR to neoadjuvant chemotherapy as indicated by both MR and PET images after chemotherapy. (Adapted from (31).) (C) Coronal  $^{18}\text{F}$ -FDG PET (top) and  $^{18}\text{F}$ -fluoride images (bottom) taken before (left) and 4 mo after (right) endocrine therapy of breast cancer widely metastatic to bone with primarily lytic metastases. Patients had excellent clinical response with dramatic reduction in tumor markers and pain.  $^{18}\text{F}$ -FDG images show substantial resolution in abnormal uptake. Fluoride images show some change, but to much lesser extent than do  $^{18}\text{F}$ -FDG images.

**TABLE 1.** Response Evaluation of Breast Cancer Therapy by <sup>18</sup>F-FDG PET

Reference	n	Therapy	Result
<b>Mid-therapy</b>			
Wahl et al. (21)	11	AC	R: -48% SUV, NR: -19% SUV
Bassa et al. (29)	15	FAC	All: -51% SUV
Schelling et al. (25)	24	EC or ET	mCR: -46% SUV, not mCR: -8% SUV
Smith et al. (26)	30	CVAP	mCR: -86% SUV, not mCR: -40% SUV
Mankoff et al. (94)	35	FAC or AC (weekly)	mCR: -65% MRFDG, PR: -49%, MRFDG, NR: -40% MRFDG
Rousseau et al. (27)	64	FEC, EC, or docetaxel	CR or near CR: -96% SUV <sub>max</sub> , R: -61% SUV <sub>max</sub> , NR: -36% SUV <sub>max</sub>
McDermott et al. (150)	96	Anthracycline-based	CR or near CR: -41% SUV, other: -27% SUV
Dunnwald et al. (41)	53	FAC or AC (weekly)	pCR: -82% MRFDG, not pCR: -62% MRFDG
<b>Early therapy</b>			
Wahl et al. (21)	11	AC	R: -22% SUV, NR: no change
Schelling et al. (25)	24	EC or ET	mCR: -54% SUV, not mCR: -19% SUV
Smith et al. (26)	30	CVAP	mCR: -77% SUV, not mCR: +1% SUV
Rousseau et al. (27)	64	FEC, EC, or docetaxel	CR or near CR: -60% SUV <sub>max</sub> , R: -36% SUV <sub>max</sub> , NR: -16% SUV <sub>max</sub>
McDermott et al. (150)	96	Anthracycline-based	CR or near CR: -20% SUV, other: -13% SUV
Berriolo-Riedinger (28)	47	FEC, CEX, ET, DCT in HER2+	pCR: -85% SUV <sub>max</sub> , not pCR: -26% SUV <sub>max</sub>

AC = doxorubicin/cyclophosphamide; R = responder; NR = nonresponder; FAC = 5-fluorouracil/doxorubicin/cyclophosphamide; EC = epirubicin/cyclophosphamide; ET = epirubicin/paclitaxel; mCR = pathologic macroscopic complete; CVAP = cyclophosphamide/vincristine/doxorubicin/prednisone; MRFDG = FDG metabolic rate; PR = partial responder; NR = nonresponder; pCR = pathologic complete responder; FEC = 5-fluorouracil/epirubicin/cyclophosphamide; CEX = cyclophosphamide/epirubicin/capecitabine; DCT = doxorubicin/carboplatin/trastuzumab.

baseline SUV as the threshold for response, <sup>18</sup>F-FDG PET was 61% sensitive and 96% specific after a single cycle and increased to 89% sensitive and 95% specific after 2 cycles of therapy. Another study by Berriolo-Riedinger reported similar findings (28). These findings suggest that <sup>18</sup>F-FDG PET may serve as an early predictor of chemotherapy response and, importantly, as an accurate predictor of lack of response, which is clinically relevant given the increasing number of new medical therapies available for breast cancer.

Studies performed after the completion of chemotherapy have shown that although residual <sup>18</sup>F-FDG uptake predicts residual disease, the absence of <sup>18</sup>F-FDG uptake is not a reliable indicator of pCR (17,29–31). This is especially true for axillary nodal disease, because the sensitivity for residual microscopic disease after therapy is low. In patients with gross residual disease, posttherapy <sup>18</sup>F-FDG PET has been shown to complement MRI to help define the extent of residual breast disease (32). Recent studies have shown that the presence of <sup>18</sup>F-FDG uptake after therapy is highly predictive of relapse (33). Therefore, even though <sup>18</sup>F-FDG PET may miss small-volume disease after therapy, the presence or absence of <sup>18</sup>F-FDG uptake may carry prognostic significance that may be important in directing the intensity of additional therapy and postsurgery surveillance.

Sestamibi imaging has also been used to assess response in the neoadjuvant setting (34,35). Studies have shown the ability of serial sestamibi imaging to discern pCR from other responses (34,35). The use of tumor uptake indices for evaluating changes in sestamibi uptake may be especially useful in this setting (36). Some comparisons of sestamibi and <sup>18</sup>F-FDG PET to measure response have

shown sestamibi to be similarly, and perhaps more, predictive of pathologic response (37). More recent studies have shown that, although sestamibi uptake after therapy may underestimate tumor extent (38), changes in sestamibi uptake with treatment and residual uptake after therapy predict relapse and survival (39). Comparative studies of sestamibi and PET blood flow measurements (40) suggest that the correspondence between sestamibi uptake and tumor blood flow may be the basis for response evaluation, and the similarity of sestamibi outcome data and PET blood flow data (39,41) supports this premise. Some studies have shown that serial sestamibi can measure response early after the start of treatment (42); however, some other studies have not supported these findings (43). Despite these successes, serial sestamibi imaging has not seen widespread use in the neoadjuvant setting; the infrequent use of sestamibi may be because, of the radiotracer methods, <sup>18</sup>F-FDG PET also provides staging information (which is important for many neoadjuvantly treated breast cancer patients).

Studies have also investigated sestamibi uptake and washout as a predictor of multidrug resistance mediated by P-glycoprotein (44). Some studies have shown that the rate of sestamibi washout before therapy, as an index of multidrug resistance, is predictive of response (45,46). Other studies have not supported the predictive value of sestamibi uptake indices (47,48). Some of the difference may be because both delivery (i.e., tumor blood flow) and retention contribute to sestamibi uptake and washout and may confound interpretation of sestamibi images for predicting drug transport (40). Ongoing work with PET probes of P-glycoprotein function in breast cancer may help in this regard (49,50).

Other functional imaging modalities have been used to evaluate breast cancer response in the neoadjuvant setting, including PET with other radiopharmaceuticals, Doppler ultrasound (51), breast MRI, breast magnetic resonance spectroscopy (MRS) (52,53), and optical imaging (54). Among these functional imaging modalities, contrast-enhanced MRI is the most widely used in clinical practice (55). Studies have shown that MRI,  $^{18}\text{F}$ -FDG PET, and sestamibi are similarly accurate in measuring response; however, MRI provides a detailed anatomic picture of the extent of disease, important for surgical planning after therapy (55). Some studies also suggest that changes in MRI provide prognostic information similar to that provided by  $^{18}\text{F}$ -FDG PET and sestamibi (56). Preliminary studies combining MRI and  $^{18}\text{F}$ -FDG PET have been promising in predicting response and delineating the extent of residual disease (32,57). Data suggest a correspondence between MRI and PET measures of tumor perfusion but not  $^{18}\text{F}$ -FDG PET measures of metabolism (58), and studies suggest that the 2 modalities provide complementary information on tumor biology (57). MRS has demonstrated an ability to measure early response to therapy (59) similar to that of  $^{18}\text{F}$ -FDG PET (60).

A particularly intriguing application of  $^{18}\text{F}$ -FDG to breast cancer treatment is as a pharmacodynamic measure of response for targeted therapy, to help predict the efficacy of targeted therapy.  $^{18}\text{F}$ -FDG PET may provide an early indication of biologic changes in response to targeted therapy that may indicate that the chosen drug has successfully hit the target and, perhaps more important, indicate when the targeted therapy has not impacted the cancer. Dehdashti et al. used  $^{18}\text{F}$ -FDG PET as an early indicator of the effect of endocrine therapy (61,62). In these studies, serial  $^{18}\text{F}$ -FDG PET showed that an increase in uptake in response to an estrogen receptor (ER) agonist, either the agonist flare of tamoxifen (62) or an estradiol challenge (61), predicted response and time to progression. Early studies also support the ability of serial  $^{18}\text{F}$ -FDG PET to measure response to lapatinib, a HER2-targeted agent (63). These studies indicate a potential of PET to identify early response to targeted therapy and merit further study.

**Metastatic Disease.** Radiotracer imaging methods have also been used to evaluate the response of stage IV, metastatic breast cancer (MBC), to therapy; however, fewer studies have been performed in this area than in others. The standard approach for response evaluation for MBC relies on changes in tumor size, typically using standard criteria such as Response Evaluation Criteria in Solid Tumors (RECIST) (64,65) and anatomic imaging, mostly CT. This approach works well for some sites of metastatic disease such as the liver and lungs. However, for other sites of disease, size-based criteria are problematic. This is true for regional nodal disease, in which the often small size of the lesion and tissue distortion from prior surgery or radiotherapy can make size-based response assessment challenging. Bone, a common site of breast cancer metastases, poses particular

challenges (66). These are areas in which functional radionuclide imaging can provide clinically relevant and important data for response assessment.

Although other radiotracer imaging approaches have been useful for evaluating primary tumor response, only  $^{18}\text{F}$ -FDG PET has seen widespread use for response evaluation of MBC. Several relatively small studies have been conducted evaluating serial  $^{18}\text{F}$ -FDG PET for measuring MBC response to treatment. Most studies showed, similar to neoadjuvant therapy, that response is accompanied by substantial drops in  $^{18}\text{F}$ -FDG uptake, typically 40%–50% or more from the pretherapy baselines (67–69). Some studies suggested that imaging as early as after 1 cycle of chemotherapy could predict response (69), akin to the neoadjuvant therapy setting. However, other studies have not shown similar accuracy for early repeated  $^{18}\text{F}$ -FDG PET (67). The level of  $^{18}\text{F}$ -FDG uptake after therapy has also been shown to be predictive. The study by Cachin et al. (70), similar to findings in the neoadjuvant setting, showed that the persistence of  $^{18}\text{F}$ -FDG uptake after therapy for patients undergoing high-dose chemotherapy predicted a significantly poorer outcome than did absence of abnormal uptake after therapy (i.e., for those patients who achieved a complete metabolic response).

A particularly vexing clinical problem for breast cancer clinicians is the evaluation of response of bone metastases (66). The size of many bony lesions is difficult to estimate and may not necessarily change with response. Bone metastases are, therefore, not considered evaluable for response by RECIST criteria (64). Bone scintigraphy continues to be an excellent tool for bone metastasis detection but is problematic for response evaluation. Changes in the bone scan significantly lag bone metastasis response and may even transiently worsen or “flare” in response to successful therapy (71,72). The flare phenomenon has been attributed to an increased uptake of diphosphonates that accompanies healing of the surrounding bone as the tumor is successfully treated, particularly for more lytic metastases (66). On the basis of molecular analyses, approximately 80% of bone metastases are primarily lytic (73). Even in the absence of an overt worsening or flare, the healing response likely contributes to the lag in bone scan changes after successful treatment. Fluoride PET, because it images bone metastases through a mechanism similar to bone scintigraphy, may yield comparable results. Preliminary results support this premise. A fluoride PET flare response has been reported (74), and early studies suggested that although fluoride PET provided quantitative information and interesting insights into physiologic changes in response to treatment, uptake did not change significantly with treatment (75).

The differences seen between bone scintigraphy and  $^{18}\text{F}$ -FDG PET for bone metastases (76) led some investigators to hypothesize that  $^{18}\text{F}$ -FDG PET might provide a useful and accurate means for assessing bone metastasis response. Specifically, although imaging such as CT, MRI, and bone scintigraphy rely on changes in the normal bone to identify

tumors,  $^{18}\text{F}$ -FDG PET might image hypermetabolism in the tumor itself and therefore provide useful information. Early studies supported this hypothesis. Stafford et al. (77) showed that changes in PET uptake correlated with the clinical assessment of response in bone-dominant MBC and, furthermore, that changes in  $^{18}\text{F}$ -FDG uptake correlated with changes in breast cancer tumor markers, which are used clinically to assess bone metastasis response. A follow-up report (78) showed that changes in  $^{18}\text{F}$ -FDG uptake predicted time to progression, a more robust clinical endpoint than response, and that the level of  $^{18}\text{F}$ -FDG predicted the likelihood of a skeletal event, perhaps as an index of the lytic nature of the bony lesions. Recent studies using  $^{18}\text{F}$ -FDG PET/CT showed similar findings and also showed that correlative changes in CT—in particular, increased sclerosis associated with response to treatment—could provide important complementary information to PET (79,80). The study by Tateishi et al. (80) also showed that both metabolic and structural changes on  $^{18}\text{F}$ -FDG PET/CT predicted bone metastasis time to progression. In interpreting  $^{18}\text{F}$ -PET studies to evaluate bone metastasis response, it is important to recognize that although an absence of uptake after therapy indicates a good response to treatment and favorable prediction for time to progression, it does not indicate an absence of disease. Examples showed that disease recurrence on PET occurred in sites of previously noted disease shortly after changing therapy, despite an absence of uptake before changing therapy (77). Nevertheless, the application of  $^{18}\text{F}$ -FDG PET/CT to bone metastasis response represents an important and clinically relevant application to breast cancer that is currently underutilized.

Some other newer imaging methods have also shown promise for evaluating breast cancer bone metastasis response. Early studies of diffusion MRI have shown promise as a method for response evaluation (81). Prospective, multicenter clinical trials are needed for  $^{18}\text{F}$ -FDG PET/CT or diffusion MRI to validate their efficacy as methods for evaluating bone metastasis response, enable their more widespread clinical trial participation, and improve clinical management for patients with bone-dominant breast cancer.

## OTHER INDICATIONS AND FUTURE DIRECTIONS

### Toxicity Monitoring

Methods commonly used in nuclear medicine play a role in toxicity monitoring for breast cancer patients. Some chemotherapy agents (e.g., doxorubicin) and, more recently, some targeted biologic agents such as trastuzumab can have significant cardiac toxicity (82). Periodic measurement of ejection fraction is needed to screen for cardiotoxicity in advance of overt symptoms of heart failure, especially for drugs such as doxorubicin, in which dysfunction is typically not reversible (82). Both radionuclide ventriculogram and echocardiography are used for the measurement of ejection fraction for monitoring cardiotox-

icity. Studies suggest that radionuclide ventriculography is less operator-dependent and more precise and therefore able to detect small decrements more reliably (83). Echocardiography involves neither radiation nor the need for blood labeling and injection. In practice, both methods are used, with the choice largely depending on institutional preference.

Osteoporosis is an increasingly common consequence of breast cancer systemic treatments, especially the aromatase inhibitors (84). Bisphosphonates are increasingly used to prevent osteoporosis in many patients, to limit progression in patients with bone metastases, and possibly as adjuvant therapy in patients with high-risk, nonmetastatic disease (85). Monitoring bone mineral density as an index of treatment effect on normal bone is important and is commonly performed using dual-energy x-ray absorptiometry (85).

### Prognostic Indicators

Predicting tumor behavior, and in particular the likelihood of cancer relapse or progression, is an increasingly important part of directing breast cancer therapy and is traditionally done by in vitro assay of tumor biopsy material. Early data suggest that parameters from radiotracer imaging may also carry prognostic information for breast cancer. Oshida (86) showed that the level of uptake in the primary tumor was predictive of disease-free survival. Other studies showed that the patterns of perfusion, measured by PET or MRI, and kinetics of  $^{18}\text{F}$ -FDG uptake were prognostically significant (41,57,87). Although it is unlikely that one would perform MRI or PET solely to gather prognostic information, such information may be readily available from scans obtained for staging or response evaluation and, therefore, merits further investigation.

### Future Directions: Molecular Imaging

As breast cancer treatment becomes increasingly targeted and individualized, demands on breast cancer diagnosis to help direct the therapeutic approach will increase. The imaging approaches in current clinical use for diagnosing and staging breast cancer—mammography, ultrasound, contrast-enhanced MRI, CT, bone scanning, and  $^{18}\text{F}$ -FDG PET/CT—provide important information on disease extent and response to treatment but are relatively nonspecific. More specific functional and molecular imaging methods, discussed below, are poised to provide the kind of diagnostic data that are needed to help take advantage of progress in our understanding of breast cancer biology and new therapeutic options. Selected examples of molecular and functional approaches include tumor perfusion and angiogenesis imaging, tumor receptor imaging, and novel approaches to early response to treatment imaging.

*Tumor Perfusion and Angiogenesis Imaging.* Tumor vascularity has been recognized as being important in cancer growth and metastasis and in the systemic delivery of therapeutic agents (88). Antiangiogenic agents such as

bevacizumab are now used as part of U.S. Food and Drug Administration–approved breast cancer treatments (89). For these reasons, the ability to image and quantify tumor perfusion and neovasculature is a clinically important need (90). Tumor perfusion is one of the earliest physiologic properties to be measured, and advances in methodology have led to increasingly quantitative approaches. The most physiologically robust and quantitative measures of tumor blood flow use freely diffusible imaging probes. With this approach, blood flow can be inferred from the time course of probe uptake and washout using methods developed and validated for measuring cerebral blood flow (91). One example is the use of  $^{15}\text{O}$ -water to measure tumor blood flow by PET, which yields measures of tumor blood flow in mL/min/g (92).  $^{15}\text{O}$ -water PET has been shown to yield reliable estimates of tumor blood flow for breast cancer (92,93). Recent studies have shown that serial measures of breast cancer perfusion by water PET in the neoadjuvant setting are highly predictive of response and survival (41,94).

Some studies have suggested that dynamic  $^{18}\text{F}$ -FDG PET and kinetic analysis may yield estimates of tumor perfusion, inferred from the  $^{18}\text{F}$ -FDG delivery ( $K_1$ ) kinetic parameter, comparable to  $^{15}\text{O}$ -water PET (95–97). Studies have shown that this parameter is predictive of response (96), and recent studies have suggested that changes in  $^{18}\text{F}$ -FDG  $K_1$  with therapy in the neoadjuvant setting are comparable to changes in perfusion from  $^{15}\text{O}$ -water in ability to predict response and survival (41). Thus, it may be possible to obtain information from dynamic  $^{18}\text{F}$ -FDG PET on both tumor perfusion and metabolism that is clinically predictive. This approach, therefore, merits further investigation.

Perhaps the mostly widely used approach to measure tumor perfusion in clinical practice is dynamic contrast-enhanced (DCE) MRI. Considerable effort has been devoted to the quantification of tumor perfusion and tumor capillary permeability using radiographic and MRI contrast agents (98,99). These agents have somewhat limited permeability across capillaries; therefore, their *in vivo* kinetics is dependent on both blood flow and capillary permeability. Increasingly sophisticated image acquisition and analysis methods—for breast DCE MRI, in particular—have led to the ability to measure regional breast cancer perfusion and capillary permeability in both animal models and patients (100). These methods have been applied in early trials of antiangiogenic breast cancer therapy and have yielded insights into the nature of response (101). Standardized methods for DCE MRI for clinical trials have been proposed (102).

Other approaches to measuring breast cancer perfusion include Doppler ultrasound (51) and optical imaging (54).

Although tumor perfusion imaging measures the physiologic consequences of angiogenesis, perfusion is not specific to tumor neovessels and is influenced by physiologic parameters not necessarily related to tumor angiogenesis (88). Targeted imaging probes can noninvasively

and specifically assess tumor neovasculature. PET probes based on labeled peptides that bind specifically to integrins expressed in neovessels have been studied in animals and tested in humans (103). MRI probes have been developed and are at an earlier stage of testing (98). Such agents may be especially helpful for therapies directed at tumor neovasculature, such as bevacizumab.

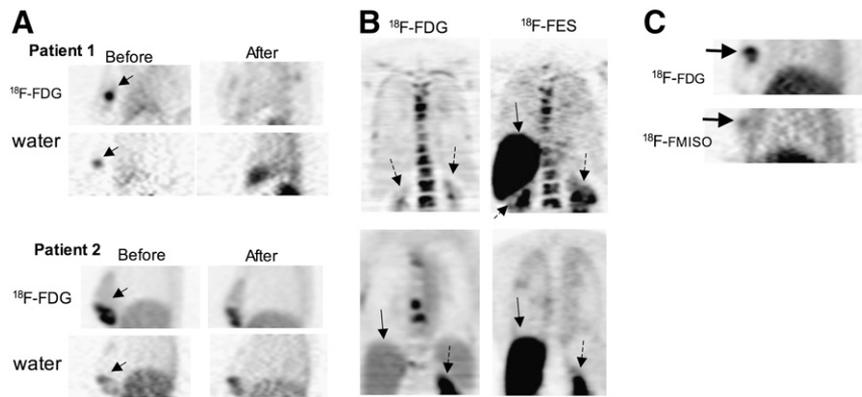
The combination of tumor perfusion and metabolism imaging may yield further insights. Studies have shown that unlike in normal tissues, the relationship between breast cancer metabolism and perfusion measured by  $^{18}\text{F}$ -FDG and water PET (Fig. 2) or by PET and MRI (57,96,97,104, 105) varies considerably. Studies have shown that the relationship between metabolism and perfusion is predictive of therapeutic response and patient outcome (41,57,105). An imbalance between metabolism and perfusion, indicated by high metabolism relative to perfusion, is associated with poor response and early relapse. More study of the biologic mechanism underlying these findings may yield insight into these striking results.

*Tumor Receptor Imaging.* The ability to measure the expression of specific proteins that are gene products associated with breast cancer has led to important advances in breast cancer treatment. The ability to image tumor receptors and proteins involved in extracellular signaling is particularly relevant to breast cancer treatment and includes the expression of ERs, a target for endocrine therapy (106), and HER2, also increasingly a target of tumor-specific treatment (3). Tumor receptors are measured by *in vitro* assay of biopsy material (107). Complementary advantages of imaging to measure receptor expression include its noninvasiveness, the ability to measure receptor expression in the entire disease burden (and thus the ability to avoid sampling error that can occur with heterogeneous receptor expression), and the potential for serial studies of *in vivo* drug effects on the target. A practical consideration is that imaging can assess receptor expression at sites that are challenging to sample and assay, for example, bone metastases, in which decalcification can make assay of tumor gene products challenging.

The imaging of tumor receptors poses some unique challenges (108). Even small molar quantities of the imaging agent may saturate the receptor and limit the ability to visualize receptor expression (109,110); thus, molecular imaging of tumor receptors has been most successful to date with radionuclide imaging, PET, and SPECT, with which it is possible to generate images with nanomolar or picomolar amounts of the imaging probe. For larger molecules, such as peptides and monoclonal antibodies, other labels suitable for optical imaging, MRI, and ultrasound imaging are possible (108); however, for small-molecule receptor imaging agents, such as labeled steroids for steroid receptors, radionuclide imaging appears to be the only feasible approach.

The most work to date for breast cancer tumor receptor imaging has been done for steroid receptors (108,111). Considerable efforts have gone into the development of

**FIGURE 2.** Examples of functional and molecular imaging of breast cancer. (A) Sagittal PET images are shown for 2 patients with breast cancer, imaged before and after neoadjuvant chemotherapy using  $^{18}\text{F}$ -FDG PET to measure tumor metabolism (top rows) and  $^{15}\text{O}$ -water PET to measure tumor perfusion (bottom rows). Patient 1 demonstrated pCR to chemotherapy, and patient 2 had significant residual disease at posttreatment surgery. Of interest, pretherapy metabolism and perfusion were well matched for patient 1, whereas metabolism and perfusion were both globally and regionally mismatched for patient 2, who had a poor response. (B) Coronal images are shown for 2 patients using  $^{18}\text{F}$ -FDG PET to identify active disease and  $^{18}\text{F}$ -FES to indicate ER expression. Both patients had extensive bony metastases arising from an ER-positive primary tumor. However, whereas  $^{18}\text{F}$ -FES PET showed that the patient had preserved ER expression in metastases, the absence of uptake for the patient in the bottom row suggests loss of ER expression. The patient in the top row responded to endocrine therapy; the patient in the bottom row did not. Normal liver (solid arrow) and kidney (dashed arrow) are indicated. (C) Sagittal images are shown from patient with locally advanced breast cancer (arrows) for  $^{18}\text{F}$ -FDG PET and  $^{18}\text{F}$ -fluoromisonidazole to indicate tumor hypoxia. Images suggested that inner core of the breast lesion was hypoxic. Patient demonstrated substantial clinical response to chemotherapy, but macroscopic viable tumor was found at postchemotherapy surgery.  $^{18}\text{F}$ -MISO =  $^{18}\text{F}$ -fluoromisonidazole.



radiopharmaceuticals for ER imaging (108,111). Although a variety of ER imaging agents have been tested, and continue to be developed and tested, the most successful ER imaging radiopharmaceutical to date is 16- $\alpha$ - $^{18}\text{F}$ -fluoro-17- $\beta$ -estradiol ( $^{18}\text{F}$ -FES) (110). This radiopharmaceutical can be synthesized with sufficient specific activity that high-quality patient images can be made with injections of less than 5  $\mu\text{g}$  of FES (Fig. 2). Regional estrogen binding is readily quantified by FES PET, and FES uptake has been validated as a measure of ER expression in breast tumors against ER expression assay of tissue samples by radioligand binding (112) and immunohistochemistry (113). FES uptake is readily visualized and quantified in primary and metastatic breast cancer (114) and can identify heterogeneous ER expression, for example, loss of ER expression in metastases arising from ER-expressing primary tumors (114,115). The level of FES uptake has been shown to be predictive of response to endocrine therapy (62,115), including heavily pretreated patients. Serial FES PET can also measure the pharmacodynamic effect of drugs on estradiol binding to the ER, yielding insights into determinants of drug efficacy (108).

Somatostatin receptor imaging of breast cancer using labeled peptides has also been studied (116,117). Although not a direct target or breast cancer therapy, somatostatin receptor expression measured by the uptake of labeled probes has been shown to be predictive of response to ER-directed therapy (117). A variety of other receptor agents has also been tested (108).

HER2 (ErbB2) expression in breast cancer has become an important indicator of prognosis and an increasingly important target for therapy (118). Recent efforts have focused on imaging HER2 expression in breast cancer. The most success and largest number of studies to date used

imaging probes based on immune recognition to image HER2 expression. Specific imaging probes based on radio-labeled antibodies or fragments, or novel constructs such as affibodies, have shown success in early studies (108). Studies using a  $^{68}\text{Ga}$ -labeled F(ab')<sub>2</sub> fragment of trastuzumab (119) demonstrated the feasibility of measuring regional HER2 expression in murine animal models. The imaging results nicely demonstrated alterations in HER2 expression accompanying experimental therapy using heat shock protein 90-directed agents (geldamycin analogs) to disrupt protein chaperoning and reduce HER2 expression (119). Studies using  $^{131}\text{I}$ - or  $^{111}\text{In}$ -labeled trastuzumab have demonstrated the ability to image tumor expression of HER2 and tumor and normal tissue accumulation of trastuzumab (120,121), although there has been some controversy about the significance of uptake in normal tissues prone to trastuzumab toxicity, such as the heart (120,121). Promising early patient studies have also been presented for  $^{89}\text{Zr}$ -labeled trastuzumab (122).

**Early Response Imaging.** Studies of  $^{18}\text{F}$ -FDG PET have shown that tumor glycolysis declines early in the course of treatment (25–27), providing a means for early response assessment. However, other pathways more closely tied to cellular growth and death may provide even earlier and more specific indications of therapeutic response. Imaging targets include protein and membrane lipid synthesis, cellular proliferation, and cell death or apoptosis.

Several methods have addressed tumor biosynthesis as an indicator of tumor growth, with approaches targeted to protein synthesis and membrane synthesis. The uptake of labeled amino acids, such as  $^{11}\text{C}$ -methionine, has been shown to correlate with tumor growth, and changes in uptake provide an early indication of breast cancer response to therapy

(123). This approach, however, is limited by the complex nature of amino acid metabolism pathways, making it difficult to measure protein synthetic rate versus amino acid transport and metabolism (124). Artificial amino acids have also been tested as indicators of amino acid transport (125).

Proliferating tumor cells also engage in enhanced lipid biosynthesis to provide the building blocks needed for cellular membranes (126). This process can also be assayed through molecular imaging using several different methods. Spurred by results in brain tumor imaging (127), MRS studies of breast cancer have shown increased choline pool sizes to be a feature of breast malignancy (52,128). Interestingly, changes in the choline concentration measured by MRS early in treatment appear to be a marker for early response to therapy, as early as 24 h after treatment with chemotherapy (59). These exciting early findings are now being tested in a large prospective cooperative group trial. Lipid metabolism can also be studied by PET using either  $^{11}\text{C}$ - or  $^{18}\text{F}$ -labeled choline or  $^{11}\text{C}$ -acetate (129), which enters lipid synthesis from the tricarboxylic acid cycle via fatty acid synthase. Fatty acid synthase has been shown to have increased activity and expression in cancer and may be a target for therapy (130). This approach has shown considerable promise in other tumors such as prostate cancer (131), including therapeutic response (132), but has not been applied to the same extent to breast cancer (133).

Aberrant cellular proliferation is a fundamental property of cancer, including breast cancer (134). Labeled compounds such as  $^{14}\text{C}$ - or  $^3\text{H}$ -thymidine have been an important method for measuring cellular proliferation through tissue sampling, dating back more than 40 years (135). Assay of breast tissue for cellular proliferation, typically by assay of MIB-1 (Ki-67) (136), is routinely performed in many centers. Early work used PET and  $^{11}\text{C}$ -thymidine to measure tumor proliferation by imaging, and quantitative imaging approaches were validated against *in vitro* assay gold standards (137). However, the short half-life of  $^{11}\text{C}$  (20 min) and the extensive *in vivo* metabolism of thymidine limit the feasibility of this approach for both animal and patient imaging. Thymidine analogs labeled with  $^{18}\text{F}$  (half-life, 109 min) have been developed and undergone considerable study in recent years (137,138). The most promising of these had been  $^{18}\text{F}$ -fluorothymidine ( $^{18}\text{F}$ -FLT), with notable recent results for both animal and patient breast cancer imaging (139–142).  $^{18}\text{F}$ -FLT PET appears especially promising for measuring the early effects of therapy on breast cancer growth, as suggested by recent studies (140,141) that showed the ability of serial  $^{18}\text{F}$ -FLT PET to identify early responders, and importantly, nonresponders. This is an exciting area of imaging research and likely to be of clinical importance in the future.

Cell death, or apoptosis, is a fundamental part of normal cellular physiology and an early indicator of therapeutic response (143). Methods for imaging cell death have been investigated. Many of these have been based on an exten-

sion of annexin V staining *in vitro*, which indicates apoptotic cells through binding to phosphatidylserines (144). The molecules are found only on the inner surface of plasma membranes and therefore normally not accessible to annexin V, a peptide, for binding. However, during apoptosis, these molecules are transiently exposed to the extracellular space, allowing binding of annexin (144). The earliest studies used  $^{99\text{m}}\text{Tc}$ -annexin and SPECT to measure apoptosis in animal models and patients (144,145). More recently, methods for annexin-based apoptosis imaging have been developed for PET, MRI, optical imaging, and ultrasound imaging (146). One limitation of this approach has been the transient nature of phosphatidylserine exposure during cell death, resulting in a fairly limited signal for imaging (147). Other approaches targeted to different aspects of the apoptotic cascade are being investigated (146). An alternative but less specific approach has been to use MRI measures of water diffusion through the extracellular space as an indirect measure of tumor cellularity (148,149). Increases in the diffusion coefficient, as an indicator of a decrease in tumor cellularity, have correlated with measures of apoptosis in animal models (148) and response to therapy in early patient studies (81,149). This method has the advantage of being available using existing MRI instrumentation for both animal and patient imaging, without the need for imaging probes, but provides a relatively indirect measure of cell death.

## CONCLUSION

Breast cancer is a common disorder in women and a leading cause of death. Imaging plays an important role in the detection, diagnosis, staging, and response evaluation of breast cancer. Radiotracer imaging methods play an important current role in breast cancer staging and response evaluation, including sentinel node lymphatic mapping, bone scintigraphy, and  $^{18}\text{F}$ -FDG PET/CT. Ongoing trials of dedicated devices for primary tumor imaging using breast-dedicated PET and SPECT may lead to the increased use for primary tumor diagnosis and determination of the extent of disease in the breast. However, more studies are needed. As breast cancer diagnosis and therapy become increasingly molecular and individualized, molecular breast cancer imaging will play a progressively more important role in breast cancer patient care. Radiotracer molecular imaging methods have already been shown to be accurate and helpful in directing the treatment of breast cancer. Future clinical trials will validate these methods and test their value in directing breast cancer treatment. Current practice and early experimental results suggest that radiotracer imaging will play an increasingly important role in the care of breast cancer patients in the future.

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## REFERENCES

1. Jemal A, Siegel R, Ward E, Murray T, Xu J, Thun MJ. Cancer statistics, 2007. *CA Cancer J Clin*. 2007;57:43–66.
2. Clarke M. Meta-analyses of adjuvant therapies for women with early breast cancer: the Early Breast Cancer Trialists' Collaborative Group overview. *Ann Oncol*. 2006;17(suppl 10):x59–x62.
3. Slamon DJ, Leyland-Jones B, Shak S, et al. Use of chemotherapy plus a monoclonal antibody against HER2 for metastatic breast cancer that over-expresses HER2. *N Engl J Med*. 2001;344:783–792.
4. Doyle DM, Miller KD. Development of new targeted therapies for breast cancer. *Breast Cancer*. 2008;15:49–56.
5. Lefferts JA, Bartels CL, Tsongalis GJ. Molecular oncology: current trends in diagnostics. *Future Oncol*. 2008;4:61–70.
6. Mankoff DA. Molecular imaging as a tool for translating breast cancer basic science. *Breast Cancer Res*. 2008;10(suppl 1):S3.
7. Gralow JR. Optimizing the treatment of metastatic breast cancer. *Breast Cancer Res Treat*. 2005;89(suppl 1):S9–S15.
8. Feldman LD, Hortobagyi GN, Buzdar AU, Ames FC, Blumenschein GR. Pathological assessment of response to induction chemotherapy in breast cancer. *Cancer Res*. 1986;46:2578–2581.
9. Moskovic EC, Mansi JL, King DM, Murch CR, Smith E. Mammography in the assessment of response to medical treatment of large primary breast tumor. *Clin Radiol*. 1993;47:339–344.
10. Chia S, Swain SM, Byrd DR, Mankoff DA. Locally advanced and inflammatory breast cancer. *J Clin Oncol*. 2008;26:786–790.
11. Gralow JR, Zujewski JA, Winer E. Preoperative therapy in invasive breast cancer: reviewing the state of the science and exploring new research directions. *J Clin Oncol*. 2008;26:696–697.
12. Ellis MJ, Ma C. Letrozole in the neoadjuvant setting: the P024 trial. *Breast Cancer Res Treat*. 2007;105(suppl 1):33–43.
13. Buzdar AU, Ibrahim NK, Francis D, et al. Significantly higher pathologic complete remission rate after neoadjuvant therapy with trastuzumab, paclitaxel, and epirubicin chemotherapy: results of a randomized trial in human epidermal growth factor receptor 2-positive operable breast cancer. *J Clin Oncol*. 2005;23:3676–3685.
14. McCready DR, Hortobagyi GN, Kau SW, Smith TL, Buzdar AU, Balch CM. The prognostic significance of lymph node metastases after preoperative chemotherapy for locally advanced breast cancer. *Arch Surg*. 1989;124:21–25.
15. Wolmark N, Wang J, Mamounas E, Bryant J, Fisher B. Preoperative chemotherapy in patients with operable breast cancer: nine-year results from National Surgical Adjuvant Breast and Bowel Project B-18. *J Natl Cancer Inst Monogr*. 2001;2001:96–102.
16. Gralow JR, Burstein HJ, Wood W, et al. Preoperative therapy in invasive breast cancer: pathologic assessment and systemic therapy issues in operable disease. *J Clin Oncol*. 2008;26:814–819.
17. Mankoff DA, Dunnwald LK. Changes in glucose metabolism and blood flow following chemotherapy for breast cancer. *PET Clin*. 2006;1:71–81.
18. Benard F, Turcotte E. Imaging in breast cancer: single-photon computed tomography and positron-emission tomography. *Breast Cancer Res*. 2005;7:153–162.
19. Quon A, Gambhir SS. FDG-PET and beyond: molecular breast cancer imaging. *J Clin Oncol*. 2005;23:1664–1673.
20. Minn H, Soini I. [<sup>18</sup>F]fluorodeoxyglucose scintigraphy in diagnosis and follow up of treatment in advanced breast cancer. *Eur J Nucl Med*. 1989;15:61–66.
21. Wahl RL, Zasadny K, Helvie M, et al. Metabolic monitoring of breast cancer chemohormonotherapy using positron emission tomography: initial evaluation. *J Clin Oncol*. 1993;11:2101–2111.
22. Rosen EL, Eubank WB, Mankoff DA. FDG PET, PET/CT, and breast cancer imaging. *Radiographics*. 2007;27(suppl 1):S215–S229.
23. Bellon JR, Livingston RB, Eubank WB, et al. Evaluation of the internal mammary lymph nodes by FDG-PET in locally advanced breast cancer (LABC). *Am J Clin Oncol*. 2004;27:407–410.
24. van der Hoeven JJ, Krak NC, Hoekstra OS, et al. <sup>18</sup>F-2-fluoro-2-deoxy-D-glucose positron emission tomography in staging of locally advanced breast cancer. *J Clin Oncol*. 2004;22:1253–1259.
25. Schelling M, Avril N, Nahrig J, et al. Positron emission tomography using [<sup>18</sup>F]fluorodeoxyglucose for monitoring primary chemotherapy in breast cancer. *J Clin Oncol*. 2000;18:1689–1695.
26. Smith I, Welch A, Hutcheon A, et al. Positron emission tomography using [<sup>18</sup>F]-fluorodeoxy-D-glucose to predict the pathologic response of breast cancer to primary chemotherapy. *J Clin Oncol*. 2000;18:1676–1688.
27. Rousseau C, Devillers A, Sagan C, et al. Monitoring of early response to neoadjuvant chemotherapy in stage II and III breast cancer by [<sup>18</sup>F]fluorodeoxyglucose positron emission tomography. *J Clin Oncol*. 2006;24:5366–5372.
28. Berriolo-Riedinger A, Touzery C, Riedinger JM, et al. [<sup>18</sup>F]FDG-PET predicts complete pathological response of breast cancer to neoadjuvant chemotherapy. *Eur J Nucl Med Mol Imaging*. 2007;34:1915–1924.
29. Bassa P, Kim EE, Inoue T, et al. Evaluation of preoperative chemotherapy using PET with fluorine-18-fluorodeoxyglucose in breast cancer. *J Nucl Med*. 1996;37:931–938.
30. Burcombe RJ, Makris A, Pittam M, Lowe J, Emmott J, Wong WL. Evaluation of good clinical response to neoadjuvant chemotherapy in primary breast cancer using [<sup>18</sup>F]-fluorodeoxyglucose positron emission tomography. *Eur J Cancer*. 2002;38:375–379.
31. Kim SJ, Kim SK, Lee ES, Ro J, Kang S. Predictive value of [<sup>18</sup>F]FDG PET for pathological response of breast cancer to neo-adjuvant chemotherapy. *Ann Oncol*. 2004;15:1352–1357.
32. Chen X, Moore MO, Lehman CD, et al. Combined use of MRI and PET to monitor response and assess residual disease for locally advanced breast cancer treated with neoadjuvant chemotherapy. *Acad Radiol*. 2004;11:1115–1124.
33. Emmering J, Krak NC, Van der Hoeven JJ, et al. Preoperative [<sup>18</sup>F] FDG-PET after chemotherapy in locally advanced breast cancer: prognostic value as compared with histopathology. *Ann Oncol*. 2008;19:1573–1577.
34. Maini CL, Tofani A, Sciuto R, et al. Technetium-99m-MIBI scintigraphy in the assessment of neoadjuvant chemotherapy in breast carcinoma. *J Nucl Med*. 1997;38:1546–1550.
35. Mankoff DA, Dunnwald LK, Gralow JR, Ellis GK, Drucker MJ, Livingston RB. Monitoring the response of patients with locally advanced breast carcinoma to neoadjuvant chemotherapy using [technetium-99m]-sestamibi scintimammography. *Cancer*. 1999;85:2410–2423.
36. Dunnwald LK, Hartnett SD, Mankoff DA. Utility and reproducibility of semiquantitative analysis of sestamibi breast images. *J Nucl Med Technol*. 1997;25:106–109.
37. Tiling R, Linke R, Untch M, et al. <sup>18</sup>F-FDG PET and <sup>99m</sup>Tc-sestamibi scintimammography for monitoring breast cancer response to neoadjuvant chemotherapy: a comparative study. *Eur J Nucl Med*. 2001;28:711–720.
38. Marshall C, Eremin J, El-Sheemy M, Eremin O, Griffiths PA. Monitoring the response of large (>3 cm) and locally advanced (T3-4, N0-2) breast cancer to neoadjuvant chemotherapy using <sup>99m</sup>Tc-sestamibi uptake. *Nucl Med Commun*. 2005;26:9–15.
39. Dunnwald LK, Gralow JR, Ellis GK, et al. Residual tumor uptake of [<sup>99m</sup>Tc]-sestamibi after neoadjuvant chemotherapy for locally advanced breast carcinoma predicts survival. *Cancer*. 2005;103:680–688.
40. Mankoff DA, Dunnwald LK, Gralow JR, et al. [Tc-99m]-sestamibi uptake and washout in locally advanced breast cancer are correlated with tumor blood flow. *Nucl Med Biol*. 2002;29:719–727.
41. Dunnwald LK, Gralow JR, Ellis GK, et al. Tumor metabolism and blood flow changes by positron emission tomography: relation to survival in patients treated with neoadjuvant chemotherapy for locally advanced breast cancer. *J Clin Oncol*. 2008;26:4449–4457.
42. Tiling R, Kessler M, Untch M, et al. Breast cancer: monitoring response to neoadjuvant chemotherapy using Tc-99m sestamibi scintimammography. *Onkologie*. 2003;26:27–31.
43. Wilczek B, von Schoultz E, Bergh J, Eriksson E, Larsson SA, Jacobsson H. Early assessment of neoadjuvant chemotherapy by FEC-courses of locally advanced breast cancer using <sup>99m</sup>Tc-MIBI. *Acta Radiol*. 2003;44:284–287.
44. Del Vecchio S, Zannetti A, Aloj L, Salvatore M. MIBI as prognostic factor in breast cancer. *Q J Nucl Med*. 2003;47:46–50.
45. Ciarmiello A, Vecchio SD, Silvestro P, et al. Tumor clearance of technetium 99m-sestamibi as a predictor of response to neoadjuvant chemotherapy for locally advanced breast cancer. *J Clin Oncol*. 1998;16:1677–1683.
46. Takamura Y, Miyoshi Y, Taguchi T, Noguchi S. Prediction of chemotherapeutic response by technetium 99m-MIBI scintigraphy in breast carcinoma patients. *Cancer*. 2001;92:232–239.
47. Travaini LL, Baio SM, Cremonesi M, et al. Neoadjuvant therapy in locally advanced breast cancer: <sup>99m</sup>Tc-MIBI mammoscintigraphy is not a reliable technique to predict therapy response. *Breast*. 2007;16:262–270.

48. Ugur Y, Sari O, Ugur O, et al. Lack of correlation between Tc-99m-sestaMIBI uptake and cadherin expression in infiltrating ductal breast carcinoma as prognostic indicators. *Ann Nucl Med*. 2003;17:281–287.
49. Kurdziel KA, Kalen JD, Hirsch JI, et al. Imaging multidrug resistance with 4-[<sup>18</sup>F]fluoropaclitaxel. *Nucl Med Biol*. 2007;34:823–831.
50. Hsueh WA, Kesner AL, Gangloff A, et al. Predicting chemotherapy response to paclitaxel with <sup>18</sup>F-fluoropaclitaxel and PET. *J Nucl Med*. 2006;47:1995–1999.
51. Kedar RP, Cosgrove DO, Smith IE, Mansi JL, Bamber JC. Breast carcinoma: measurement of tumor response to primary medical therapy with color flow Doppler imaging. *Radiology*. 1994;190:825–830.
52. Bolan PJ, Nelson MT, Yee D, Garwood M. Imaging in breast cancer: magnetic resonance spectroscopy. *Breast Cancer Res*. 2005;7:149–152.
53. Lehman CD, Schnall MD. Imaging in breast cancer: magnetic resonance imaging. *Breast Cancer Res*. 2005;7:215–219.
54. Tromberg BJ, Cerussi A, Shah N, et al. Imaging in breast cancer: diffuse optics in breast cancer—detecting tumors in pre-menopausal women and monitoring neoadjuvant chemotherapy. *Breast Cancer Res*. 2005;7:279–285.
55. Hylton N. MR imaging for assessment of breast cancer response to neoadjuvant chemotherapy. *Magn Reson Imaging Clin N Am*. 2006;14:383–389.
56. Li KL, Partridge SC, Joe BN, et al. Invasive breast cancer: predicting disease recurrence by using high-spatial-resolution signal enhancement ratio imaging. *Radiology*. 2008;248:79–87.
57. Semple SI, Gilbert FJ, Redpath TW, et al. The relationship between vascular and metabolic characteristics of primary breast tumours. *Eur Radiol*. 2004;14:2038–2045.
58. Eby PR, Partridge S, White SW, et al. Metabolic and vascular features of dynamic contrast-enhanced breast magnetic resonance imaging and <sup>15</sup>O-water positron emission tomography blood flow in breast cancer. *Acad Radiol*. 2008;15:1246–1254.
59. Meisamy S, Bolan PJ, Baker EH, et al. Neoadjuvant chemotherapy of locally advanced breast cancer: predicting response with in vivo <sup>1</sup>H MR spectroscopy—a pilot study at 4 T. *Radiology*. 2004;233:424–431.
60. Tozaki M, Sakamoto M, Oyama Y, et al. Monitoring of early response to neoadjuvant chemotherapy in breast cancer with <sup>1</sup>H MR spectroscopy: comparison to sequential 2-[<sup>18</sup>F]-fluorodeoxyglucose positron emission tomography. *J Magn Reson Imaging*. 2008;28:420–427.
61. Dehdashti F, Mortimer JE, Trinkaus K, et al. PET-based estradiol challenge as a predictive biomarker of response to endocrine therapy in women with estrogen-receptor-positive breast cancer. *Breast Cancer Res Treat*. 2008;113:509–517.
62. Mortimer JE, Dehdashti F, Siegel BA, Trinkaus K, Katzenellenbogen JA, Welch MJ. Metabolic flare: indicator of hormone responsiveness in advanced breast cancer. *J Clin Oncol*. 2001;19:2797–2803.
63. Kawada K, Murakami K, Sato T, et al. Prospective study of positron emission tomography for evaluation of the activity of lapatinib, a dual inhibitor of the ErbB1 and ErbB2 tyrosine kinases, in patients with advanced tumors. *Jpn J Clin Oncol*. 2007;37:44–48.
64. Therasse P, Arbuck SG, Eisenhauer EA, et al. New guidelines to evaluate the response to treatment in solid tumors. European Organization for Research and Treatment of Cancer, National Cancer Institute of the United States, National Cancer Institute of Canada. *J Natl Cancer Inst*. 2000;92:205–216.
65. Therasse P, Eisenhauer EA, Byss M. Update in methodology and conduct of cancer clinical trials. *Eur J Cancer*. 2006;42:1322–1330.
66. Hamaoka T, Madewell JE, Podoloff DA, Hortobagyi GN, Ueno NT. Bone imaging in metastatic breast cancer. *J Clin Oncol*. 2004;22:2942–2953.
67. Couturier O, Jerusalem G, N’Guyen JM, Hustinx R. Sequential positron emission tomography using [<sup>18</sup>F]fluorodeoxyglucose for monitoring response to chemotherapy in metastatic breast cancer. *Clin Cancer Res*. 2006;12:6437–6443.
68. Dose Schwarz J, Bader M, Jenicke L, Hemminger G, Janicke F, Avril N. Early prediction of response to chemotherapy in metastatic breast cancer using sequential <sup>18</sup>F-FDG PET. *J Nucl Med*. 2005;46:1144–1150.
69. Gennari A, Donati S, Salvadori B, et al. Role of 2-[<sup>18</sup>F]-fluorodeoxyglucose (FDG) positron emission tomography (PET) in the early assessment of response to chemotherapy in metastatic breast cancer patients. *Clin Breast Cancer*. 2000;1:156–161.
70. Cachin F, Prince HM, Hogg A, Ware RE, Hicks RJ. Powerful prognostic stratification by [<sup>18</sup>F]fluorodeoxyglucose positron emission tomography in patients with metastatic breast cancer treated with high-dose chemotherapy. *J Clin Oncol*. 2006;24:3026–3031.
71. Coleman RE, Mashiter G, Whitaker KB, Moss DW, Rubens RD, Fogelman I. Bone scan flare predicts successful systemic therapy for bone metastases. *J Nucl Med*. 1988;29:1354–1359.
72. Schneider JA, Divgi CR, Scott AM, et al. Flare on bone scintigraphy following Taxol chemotherapy for metastatic breast cancer. *J Nucl Med*. 1994;35:1748–1752.
73. Lipton A. Pathophysiology of bone metastases: how this knowledge may lead to therapeutic intervention. *J Support Oncol*. 2004;2:205–220.
74. Wade AA, Scott JA, Kuter I, Fischman AJ. Flare response in <sup>18</sup>F-fluoride ion PET bone scanning. *AJR*. 2006;186:1783–1786.
75. Gralow J, Brenner W, Linden H, et al. Changes in tumor metabolism and local bone turnover in patients treated for bone-dominant metastatic breast cancer measured by fluorodeoxyglucose (FDG) and fluoride positron emission tomography (PET) [abstract]. *Breast Cancer Res Treat*. 2005;94(suppl 1):S261.
76. Cook GJ, Houston S, Rubens R, Maisey MN, Fogelman I. Detection of bone metastases in breast cancer by <sup>18</sup>FDG PET: differing metabolic activity in osteoblastic and osteolytic lesions. *J Clin Oncol*. 1998;16:3375–3379.
77. Stafford SE, Gralow JR, Schubert EK, et al. Use of serial FDG PET to measure the response of bone-dominant breast cancer to therapy. *Acad Radiol*. 2002;9:913–921.
78. Specht JM, Tam SL, Kurland BF, et al. Serial 2-[<sup>18</sup>F] fluoro-2-deoxy-D-glucose positron emission tomography (FDG-PET) to monitor treatment of bone-dominant metastatic breast cancer predicts time to progression (TTP). *Breast Cancer Res Treat*. 2007;105:87–94.
79. Du Y, Cullum I, Illidge TM, Ell PJ. Fusion of metabolic function and morphology: sequential [<sup>18</sup>F]fluorodeoxyglucose positron-emission tomography/computed tomography studies yield new insights into the natural history of bone metastases in breast cancer. *J Clin Oncol*. 2007;25:3440–3447.
80. Tateishi U, Gamez C, Dawood S, Yeung HW, Cristofanilli M, Macapinlac HA. Bone metastases in patients with metastatic breast cancer: morphologic and metabolic monitoring of response to systemic therapy with integrated PET/CT. *Radiology*. 2008;247:189–196.
81. Theilmann RJ, Borders R, Trouard TP, et al. Changes in water mobility measured by diffusion MRI predict response of metastatic breast cancer to chemotherapy. *Neoplasia*. 2004;6:831–837.
82. Ng R, Better N, Green MD. Anticancer agents and cardiotoxicity. *Semin Oncol*. 2006;33:2–14.
83. Godkar D, Bachu K, Dave B, Megna R, Niranjana S, Khanna A. Comparison and co-relation of invasive and noninvasive methods of ejection fraction measurement. *J Natl Med Assoc*. 2007;99:1227–1228, 1231–1224.
84. Chien AJ, Goss PE. Aromatase inhibitors and bone health in women with breast cancer. *J Clin Oncol*. 2006;24:5305–5312.
85. Aapro M, Abrahamson PA, Body JJ, et al. Guidance on the use of bisphosphonates in solid tumours: recommendations of an international expert panel. *Ann Oncol*. 2008;19:420–432.
86. Oshida M, Uno K, Suzuki M, et al. Predicting the prognoses of breast carcinoma patients with positron emission tomography using 2-deoxy-2-fluoro[<sup>18</sup>F]-D-glucose. *Cancer*. 1998;82:2227–2234.
87. Esserman L, Kaplan E, Partridge S, et al. MRI phenotype is associated with response to doxorubicin and cyclophosphamide neoadjuvant chemotherapy in stage III breast cancer. *Ann Surg Oncol*. 2001;8:549–559.
88. Jain RK. Antiangiogenic therapy for cancer: current and emerging concepts. *Oncology (Williston Park)*. 2005;19(4 suppl 3):7–16.
89. Hayes DF, Miller K, Sledge G. Angiogenesis as targeted breast cancer therapy. *Breast*. 2007;16(suppl 2):S17–S19.
90. Miller JC, Pien HH, Sahani D, Sorensen AG, Thrall JH. Imaging angiogenesis: applications and potential for drug development. *J Natl Cancer Inst*. 2005;97:172–187.
91. Kety SS. Basic principles for the quantitative estimation of regional cerebral blood flow. *Res Publ Assoc Res Nerv Ment Dis*. 1985;63:1–7.
92. Wilson CB, Lammertsma AA, McKenzie CG, Sikora K, Jones T. Measurements of blood flow and exchanging water space in breast tumors using positron emission tomography: a rapid and non-invasive dynamic method. *Cancer Res*. 1992;52:1592–1597.
93. Wells P, Jones T, Price P. Assessment of inter- and inpatient variability in C<sup>15</sup>O<sub>2</sub> positron emission tomography measurements of blood flow in patients with intra-abdominal cancers. *Clin Cancer Res*. 2003;9:6350–6356.
94. Mankoff DA, Dunnwald LK, Gralow JR, et al. Changes in blood flow and metabolism in locally advanced breast cancer treated with neoadjuvant chemotherapy. *J Nucl Med*. 2003;44:1806–1814.
95. Mullani NA, Herbst RS, O’Neil RG, Gould KL, Barron BJ, Abbruzzese JL. Tumor blood flow measured by PET dynamic imaging of first-pass <sup>18</sup>F-FDG uptake: a comparison with <sup>15</sup>O-labeled water-measured blood flow. *J Nucl Med*. 2008;49:517–523.
96. Tseng J, Dunnwald LK, Schubert EK, et al. <sup>18</sup>F-FDG kinetics in locally advanced breast cancer: correlation with tumor blood flow and changes in response to neoadjuvant chemotherapy. *J Nucl Med*. 2004;45:1829–1837.
97. Zasadny KR, Tatsumi M, Wahl RL. FDG metabolism and uptake versus blood flow in women with untreated primary breast cancers. *Eur J Nucl Med Mol Imaging*. 2003;30:274–280.

98. Barrett T, Brechbiel M, Bernardo M, Choyke PL. MRI of tumor angiogenesis. *J Magn Reson Imaging*. 2007;26:235–249.
99. Padhani AR, Leach MO. Antivascular cancer treatments: functional assessments by dynamic contrast-enhanced magnetic resonance imaging. *Abdom Imaging*. 2005;30:324–341.
100. Yankeelov TE, Lepage M, Chakravarthy A, et al. Integration of quantitative DCE-MRI and ADC mapping to monitor treatment response in human breast cancer: initial results. *Magn Reson Imaging*. 2007;25:1–13.
101. Wedam SB, Low JA, Yang SX, et al. Antiangiogenic and antitumor effects of bevacizumab in patients with inflammatory and locally advanced breast cancer. *J Clin Oncol*. 2006;24:769–777.
102. Leach MO, Brindle KM, Evelhoch JL, et al. Assessment of antiangiogenic and antivascular therapeutics using MRI: recommendations for appropriate methodology for clinical trials. *Br J Radiol*. 2003;76(special issue 2003):S87–S91.
103. Beer AJ, Haubner R, Sarbia M, et al. Positron emission tomography using [<sup>18</sup>F]galacto-RGD identifies the level of integrin  $\alpha_v\beta_3$  expression in man. *Clin Cancer Res*. 2006;12:3942–3949.
104. Hentschel M, Paulus T, Mix M, Moser E, Nitzsche EU, Brink I. Analysis of blood flow and glucose metabolism in mammary carcinomas and normal breast: a H<sub>2</sub><sup>18</sup>O PET and <sup>18</sup>F-FDG PET study. *Nucl Med Commun*. 2007;28:789–797.
105. Mankoff DA, Dunnwald LK, Gralow JR, et al. Blood flow and metabolism in locally advanced breast cancer: relationship to response to therapy. *J Nucl Med*. 2002;43:500–509.
106. Jordan VC, Brodie AM. Development and evolution of therapies targeted to the estrogen receptor for the treatment and prevention of breast cancer. *Steroids*. 2007;72:7–25.
107. Osborne CK, Yochmowitz MG, Knight WA III, McGuire WL. The value of estrogen and progesterone receptors in the treatment of breast cancer. *Cancer*. 1980;46(12, suppl):2884–2888.
108. Mankoff DA, Link JM, Linden HM, Sundararajan L, Krohn KA. Tumor receptor imaging. *J Nucl Med*. 2008;49(suppl 2):149S–163S.
109. Katzenellenbogen J. The pharmacology of steroid radiopharmaceuticals: specific and non-specific binding and uptake selectivity. In: Nunn A, ed. *Radiopharmaceuticals: Chemistry and Pharmacology*. New York, NY: Marcel Dekker; 1992:297–331.
110. Katzenellenbogen JA, Welch MJ, Dehdashti F. The development of estrogen and progesterone radiopharmaceuticals for imaging breast cancer. *Anticancer Res*. 1997;17:1573–1576.
111. Katzenellenbogen JA. Designing steroid receptor-based radiotracers to image breast and prostate tumors. *J Nucl Med*. 1995;36(6, suppl):8S–13S.
112. Mintun MA, Welch MJ, Siegel BA, et al. Breast cancer: PET imaging of estrogen receptors. *Radiology*. 1988;169:45–48.
113. Peterson LM, Mankoff DA, Lawton TJ, et al. Quantitative imaging of estrogen receptor expression in breast cancer with PET and <sup>18</sup>F-fluoroestradiol. *J Nucl Med*. 2008;49:367–374.
114. Dehdashti F, Mortimer JE, Siegel BA, et al. Positron tomographic assessment of estrogen receptors in breast cancer: comparison with FDG-PET and in vitro receptor assays. *J Nucl Med*. 1995;36:1766–1774.
115. Linden HM, Stekhova SA, Link JM, et al. Quantitative fluoroestradiol positron emission tomography imaging predicts response to endocrine treatment in breast cancer. *J Clin Oncol*. 2006;24:2793–2799.
116. Mezi S, Primi F, Orsi E, Capocetti F, Scopinaro F, Schillaci O. Somatostatin receptor scintigraphy in metastatic breast cancer patients. *Oncol Rep*. 2005;13:31–35.
117. Van Den Bossche B, Van Belle S, De Winter F, Signore A, Van de Wiele C. Early prediction of endocrine therapy effect in advanced breast cancer patients using <sup>99m</sup>Tc-depreotide scintigraphy. *J Nucl Med*. 2006;47:6–13.
118. Harris L, Fritsche H, Mennel R, et al. American Society of Clinical Oncology 2007 update of recommendations for the use of tumor markers in breast cancer. *J Clin Oncol*. 2007;25:5287–5312.
119. Smith-Jones PM, Solit D, Afroze F, Rosen N, Larson SM. Early tumor response to Hsp90 therapy using HER2 PET: comparison with <sup>18</sup>F-FDG PET. *J Nucl Med*. 2006;47:793–796.
120. de Korte MA, de Vries EG, Lub-de Hooge MN, et al. <sup>111</sup>Indium-trastuzumab visualises myocardial human epidermal growth factor receptor 2 expression shortly after anthracycline treatment but not during heart failure: a clue to uncover the mechanisms of trastuzumab-related cardiotoxicity. *Eur J Cancer*. 2007;43:2046–2051.
121. Perik PJ, Lub-De Hooge MN, Gietema JA, et al. Indium-111-labeled trastuzumab scintigraphy in patients with human epidermal growth factor receptor 2-positive metastatic breast cancer. *J Clin Oncol*. 2006;24:2276–2282.
122. Dijkers E, Lub-de Hooge MN, Kosterink JG, et al. Characterization of <sup>89</sup>Zr-trastuzumab for clinical HER2 immunoPET imaging [abstract]. *J Clin Oncol*. 2007;25:140s.
123. Jansson T, Westlin JE, Ahlstrom H, Lilja A, Langstrom B, Bergh J. Positron emission tomography studies in patients with locally advanced and/or metastatic breast cancer: a method for early therapy evaluation? *J Clin Oncol*. 1995;13:1470–1477.
124. Ishiwata K, Enomoto K, Sasaki T, et al. A feasibility study on L-[1-carbon-11]tyrosine and L-[methyl-carbon-11]methionine to assess liver protein synthesis by PET. *J Nucl Med*. 1996;37:279–285.
125. Hubner KF, Thie JA, Smith GT, et al. Positron emission tomography (PET) with 1-aminocyclobutane-1-[<sup>11</sup>C]carboxylic acid (1-[<sup>11</sup>C]-ACBC) for detecting recurrent brain tumors. *Clin Positron Imaging*. 1998;1:165–173.
126. Glunde K, Jacobs MA, Bhujwalla ZM. Choline metabolism in cancer: implications for diagnosis and therapy. *Expert Rev Mol Diagn*. 2006;6:821–829.
127. Nelson SJ. Multivoxel magnetic resonance spectroscopy of brain tumors. *Mol Cancer Ther*. 2003;2:497–507.
128. Stanwell P, Mountford C. In vivo proton MR spectroscopy of the breast. *Radiographics*. 2007;27(suppl 1):S253–S266.
129. Groves AM, Win T, Haim SB, Ell PJ. Non-[<sup>18</sup>F]FDG PET in clinical oncology. *Lancet Oncol*. 2007;8:822–830.
130. Lupu R, Menendez JA. Targeting fatty acid synthase in breast and endometrial cancer: an alternative to selective estrogen receptor modulators? *Endocrinology*. 2006;147:4056–4066.
131. Powles T, Murray I, Brock C, Oliver T, Avril N. Molecular positron emission tomography and PET/CT imaging in urological malignancies. *Eur Urol*. 2007;51:1511–1520.
132. Yu EY, Mankoff DA. Positron emission tomography imaging as a cancer biomarker. *Expert Rev Mol Diagn*. 2007;7:659–672.
133. Zheng QH, Stone KL, Mock BH, et al. [<sup>11</sup>C]Choline as a potential PET marker for imaging of breast cancer athymic mice. *Nucl Med Biol*. 2002;29:803–807.
134. Tannock IF. Cell proliferation. In: Tannock IF, Hill RP, eds. *The Basic Science of Oncology*. New York, NY: McGraw-Hill; 1992:154–177.
135. Cleaver JE. *Thymidine Metabolism and Cell Kinetics*. Amsterdam, The Netherlands: North-Holland Pub. Co.; 1967.
136. Pinder SE, Wencyk P, Sibbering DM, et al. Assessment of the new proliferation marker MIB1 in breast carcinoma using image analysis: associations with other prognostic factors and survival. *Br J Cancer*. 1995;71:146–149.
137. Mankoff DA, Shields AF, Krohn KA. PET imaging of cellular proliferation. *Radiol Clin North Am*. 2005;43:153–167.
138. Shields AF, Grierson JR, Dohmen BM, et al. Imaging proliferation in vivo with [<sup>18</sup>F]FLT and positron emission tomography. *Nat Med*. 1998;4:1334–1336.
139. Been LB, Elsinga PH, de Vries J, et al. Positron emission tomography in patients with breast cancer using <sup>18</sup>F-3'-deoxy-3'-[<sup>18</sup>F]-thymidine (<sup>18</sup>F-FLT): a pilot study. *Eur J Surg Oncol*. 2006;32:39–43.
140. Kenny L, Coombes RC, Vigushin DM, Al-Nahhas A, Shousha S, Aboagye EO. Imaging early changes in proliferation at 1 week post chemotherapy: a pilot study in breast cancer patients with 3'-deoxy-3'-[<sup>18</sup>F]fluorothymidine positron emission tomography. *Eur J Nucl Med Mol Imaging*. 2007;34:1339–1347.
141. Pio BS, Park CK, Pietras R, et al. Usefulness of 3'-[<sup>18</sup>F]-thymidine with positron emission tomography in predicting breast cancer response to therapy. *Mol Imaging Biol*. 2006;8:36–42.
142. Smyczek-Gargya B, Fersis N, Dittmann H, et al. PET with [<sup>18</sup>F]fluorothymidine for imaging of primary breast cancer: a pilot study. *Eur J Nucl Med Mol Imaging*. 2004;31:720–724.
143. Hockenbery D. Defining apoptosis. *Am J Pathol*. 1995;146:16–19.
144. Blankenberg F, Ohtsuki K, Strauss HW. Dying a thousand deaths: radionuclide imaging of apoptosis. *Q J Nucl Med*. 1999;43:170–176.
145. van de Wiele C, Lahorte C, Vermeersch H, et al. Quantitative tumor apoptosis imaging using technetium-99m-HYNIC annexin V single photon emission computed tomography. *J Clin Oncol*. 2003;21:3483–3487.
146. Schoenberger J, Bauer J, Moosbauer J, Eilles C, Grimm D. Innovative strategies in vivo apoptosis imaging. *Curr Med Chem*. 2008;15:187–194.
147. Tait JF, Smith C, Blankenberg FG. Structural requirements for in vivo detection of cell death with <sup>99m</sup>Tc-annexin V. *J Nucl Med*. 2005;46:807–815.
148. Morse DL, Galons JP, Payne CM, et al. MRI-measured water mobility increases in response to chemotherapy via multiple cell-death mechanisms. *NMR Biomed*. 2007;20:602–614.
149. Lee KC, Moffat BA, Schott AF, et al. Prospective early response imaging biomarker for neoadjuvant breast cancer chemotherapy. *Clin Cancer Res*. 2007;13:443–450.
150. McDermott GM, Welch A, Staff RT, et al. Monitoring primary breast cancer throughout chemotherapy using FDG-PET. *Breast Cancer Res Treat*. 2007;102:75–84.