Impact of Contamination from Scattered Photons in Singles-Mode Transmission Data on Quantitative Small-Animal PET Imaging

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In previous work, we described and validated a method of scatter correction for singles-mode transmission data using experimental preinjection data acquired with a dedicated rodent PET scanner. In the current work, we investigated the impact that our method has on the quantitative accuracy of small-animal PET. Methods: This investigation had 3 stages. We first confirmed the general validity of our method by applying it to preinjection transmission data from a different imaging system (a larger dedicated primate scanner). For these data, we evaluated the accuracy of the reconstructed distributions of linear attenuation coefficients (µ-values). In the second stage, we applied our attenuationmap reconstruction and scatter correction procedure for postinjection transmission data acquired with the dedicated rodent scanner. For these studies, we investigated the quantitative accuracy of reconstructed emission images that use attenuation correction derived from postinjection transmission data. In the third stage, we compared our scatter correction method with 2 more commonly used alternatives (automated rescaling and segmentation of the attenuation-map images). Results: For the primate scanner data, the average reconstructed µ-values with scatter correction were within 3% of the expected values for water and soft tissue, whereas uncorrected values were 19%-26% lower than their expected values. For the postinjection transmission studies, we found that the correct average µ-values and reconstructed activity concentrations consistent with wellcounter measurements were obtained only when scatter correction and emission contamination correction were applied to the transmission data. We also found that our transmission scatter correction provides more accurate µ-values and better image quantification than either rescaling or segmentation. Conclusion: Using different imaging systems (primate and rodent) and different scanning protocols (before and after injection), we found that our transmission scatter correction is more accurate (for both reconstructed µ-values and activity concentrations) than the existing alternatives.

Key Words: small-animal PET; attenuation correction; singlesmode transmission; postinjection transmission; quantitative PET

J Nucl Med 2008; 49:1852–1861 DOI: 10.2967/jnumed.108.051193 A ttenuation correction factors, which are necessary for quantitatively accurate PET, can be obtained using a singles-mode transmission scan. These scans are capable of a higher counting rate than are conventional coincidencemode transmission acquisitions. However, contamination from scattered photons and from emission activity (in postinjection transmissions) can be a significant problem. A correction for emission contamination can be obtained directly by acquiring a "mock" transmission scan (1), in which the transmission source is removed or shielded from the detectors. In previous work (2), we presented a scatter correction for singles-mode transmission data and validated our results using preinjection data from the microPET Focus 120 (Siemens Medical Solutions USA, Inc.) (3,4), a dedicated rodent scanner.

In this work, we applied our method of scatter correction using transmission data from the microPET Focus 220 (Siemens Medical Solutions USA, Inc.) (5), a scanner suitable for primate studies. The purpose of these studies was to test the general validity of our scatter correction for larger objects (subject to more photon attenuation and higher amounts of both single and multiple scatter) and for a larger-diameter PET scanner (with a decreased solid angle for the detection of both scattered and unscattered photons). Then, using data from the microPET Focus 120, we tested our analytic scatter correction for postinjection transmission data. For both scanners, we evaluated the accuracy of the reconstructed linear attenuation coefficients (µ-values). For the microPET Focus 120 data, we also investigated the quantitative accuracy of reconstructed emission images in terms of absolute quantitative units (MBq). For the attenuation and scatter correction applied to the emission data, we used attenuation maps reconstructed with and without correction for scatter and emission contamination obtained from mock scans.

Singles-mode transmission data are not typically corrected for scatter. More commonly, these data are reconstructed and the resulting attenuation-map images are segmented. In these segmentation algorithms, each voxel of the attenuation map is classified as a particular type of tissue and then reassigned a

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theoretic μ -value for that tissue type. To save time and reduce interuser variability, most segmentation routines are automated or require minimal user intervention (6,7). Alternatively, the reconstructed attenuation-map data could simply be rescaled using the knowledge that most biologic materials (e.g., soft tissue and muscle) have attenuation coefficients close to that of water. Using 2 simple phantom configurations (water cylinders filled with uniform concentrations of ¹⁸F activity), we examined how these 2 alternatives to transmission scatter correction can lead to both qualitative and quantitative errors in the reconstructed activity concentrations for PET images.

MATERIALS AND METHODS

Primate Scanner Studies

The microPET Focus 220, a dedicated PET scanner for primates (5), uses LSO scintillator blocks each consisting of a 12×12 crystal array (crystal size, $1.5 \times 1.5 \times 10$ mm). Each block is coupled to a position-sensitive photomultiplier tube by a fiberoptic bundle. There are a total of 504 detectors per ring and 48 crystal rings. The ring diameter of the scanner is 261 mm, and the axial length is 77 mm. All transmission data for this scanner were acquired using a pointlike ⁵⁷Co (122-keV photon emitter) transmission source that rotates and translates axially through the scanner field of view. For photons emitted by the ⁵⁷Co source, an extremely narrow energy window between 120 and 125 keV is used to reduce the acquisition of scattered photons during transmission scanning.

Singles-mode transmission data were acquired for a 50-mmradius water cylinder offset vertically from the center of the scanner field of view by 27 mm. For this study, no activity was present in the cylinder, the blank data were acquired for 2 h, and the duration of the transmission scan was 27 min. The primate transmission data were acquired before injection (no emission contamination was present in the scanner) using a 6.5-kg *Macaca nemestrina* monkey. Our scatter correction was originally validated for rats and mice (2), which typically weigh 300 and 30 g, respectively. For this study, the blank and transmission data were acquired for 2 h and 45 min, respectively. Because these scan durations were much longer than what is typically used for smallanimal PET transmission scans, we expected that these data would provide low-noise images in which the systematic effects of scatter would be more evident.

All transmission and blank data were processed using singleslice rebinning (8) with a maximum oblique angle of 8.9° (corresponding to a maximum ring difference of 23). All micro-PET Focus 220 transmission data were reconstructed using ordered-subsets transmission (9), a maximum a posteriori reconstruction algorithm for transmission tomography. For all our reconstructions, the size of the reconstructed attenuation-map images was 256 × 256, and we used 20 iterations with 4 subsets, a smoothing intensity parameter $\beta = 2^8$, and a Huber penalty function (10) with a cutoff parameter of $\delta = 0.5 \times \mu_{water}$, where μ_{water} is the linear attenuation coefficient for water at the appropriate photon energy. The Huber penalty function applies heavier quadratic smoothing for neighboring voxels with reconstructed μ -values that differ by less than δ . For differences greater than δ , linear smoothing is applied to discourage oversmoothing of abrupt changes, which should occur at the boundaries between different tissue types.

For reconstruction of the transmission data, we used our previously described scatter correction and reconstruction procedure (2). Our transmission scatter correction is similar to the single scatter simulation (11) that is now commonly used for coincidence-mode emission data. We use an estimate of the attenuation map (an image representing the distribution of linear attenuation coefficients within the subject being imaged) and the Klein–Nishina scattering crosssection to compute the contribution from single scattered (one Compton scatter interaction) photons to each crystal pair in the PET detector array during the transmission scan. Because our scatter correction requires an estimate of the attenuation map for its calculation, 5 distinct steps are needed:

- 1. We first reconstruct an initial attenuation map with no correction for scatter.
- This uncorrected attenuation-map image is processed into a histogram of linear attenuation coefficients (μ-values). The image is then automatically rescaled so that the maximum histogram bin corresponds to the correct μ-value for water.
- 3. We compute scatter sinogram data using the rescaled attenuation map.
- 4. Each scatter sinogram is rescaled to account for multiple scatter and scatter from outside the FOV and then subtracted from the experimental transmission data.
- 5. The scatter-corrected transmission data are then reconstructed.

Studies Using Postinjection Transmission Data

Experiments. Experimental postinjection transmission data were acquired using the microPET Focus 120. The microPET Focus 120 and microPET Focus 220 are similar scanners that share hardware and software. For example, they use the same block detector modules, have the same crystal size $(1.5 \times 1.5 \times 10)$ mm), have the same number of axial crystal rings (48), and share the same acquisition software. The primary difference between the 2 scanners is that the microPET Focus 120 has fewer crystals per axial ring (288 in total), arranged in a smaller-diameter (140 mm) ring. For each experiment using this scanner, 2 sets of singlesmode transmission data were acquired using pointlike ⁶⁸Ge (a positron emitter) and ⁵⁷Co (a 122-keV photon emitter) transmission sources. The average measured energy resolution of the system is 18% at 511 keV and 35% at 122 keV. For the annihilation photons emitted by the 68Ge source, an energy discrimination window between 350 and 750 keV is used. All transmission and blank data that were acquired using a ⁶⁸Ge source in this work were precorrected for contamination because of the naturally occurring ¹⁷⁶Lu radioactivity in the crystals (12). For the ⁵⁷Co source, the same energy discrimination settings were used as those described previously for the microPET Focus 220.

We acquired postinjection data for a 30-mm-radius water cylinder with a 68 Ge line source, containing approximately 1.3 MBq, attached to the side. The length of the line source was 115 mm, which is larger than the axial length of the scanner (77 mm); we therefore estimated that approximately 0.9 MBq of the line-source activity was present inside the scanner during the emission scan. Two sets of transmission and blank scans were acquired using a 14-MBq 68 Ge source and a 100-MBq 57 Co source. The

duration of all blank, transmission, and mock scans was 17 min. The duration of the emission scan was 30 min.

We also acquired transmission and blank data for a healthy rodent injected with ¹⁸F-FDG. We attached to the rat's shoulder a small glass vial (diameter of 10 mm and length of 34 mm) filled with a solution of ¹⁸F (2.0 mL). This vial was meant to approximate an external tumor similar to oncology studies in which human cancer cells are transplanted close to the surface of a rodent's skin (13, 14). At the beginning of the emission scan, we estimated that 6 MBq of ¹⁸F activity were present in the simulated tumor and about 24 MBq of ¹⁸F-FDG activity were distributed throughout the rat. At the time of the rodent studies, the ⁶⁸Ge and ⁵⁷Co transmission sources had activities of 10 and 72 MBq, respectively. For this study, all blank and transmission scans were acquired for 17 min, whereas the mock scans were acquired for only 8.5 min. This duration is more typical in animal studies, in which the mock acquisitions are short to minimize total scan times. The duration of the emission scan was 20 min.

Transmission Data Reconstruction. The postinjection transmission data acquired using the microPET Focus 120 were reconstructed using ordered-subsets transmission (9) with the same parameters as were described previously with the exception of image size, which was 128×128 for these data, and maximum oblique angle, which was 13.7° (corresponding to a maximum ring difference of 23) for single-slice rebinning with this scanner. For this analysis, we compared reconstructions performed with 3 different combinations of emission-contamination correction and scatter correction:

- 1. No correction. For these data, we reconstruct with no correction for scatter or for emission contamination, to determine the combined importance of these effects.
- 2. Correction for emission contamination only. For this reconstruction, we subtract the experimentally measured mock scan sinograms (corrected for radioactive decay of the emission tracer) from the transmission data. We then reconstruct the data with no correction for transmission photon scatter.
- Correction for both emission contamination and scatter. This correction also subtracts the decay-corrected mock scans from the transmission data but applies our scatter correction and reconstruction procedure.

Emission Data Reconstruction. The PET emission data, acquired using the microPET Focus 120, were sorted into 3-dimensional sinograms. The emission sinogram data were corrected for random events by subtracting data acquired in a delayed coincidence window. The sinogram data were then corrected for detector normalization using a component-based method (15). Using Fourier rebinning (16), the 3-dimensional sinograms were rebinned into a smaller set of 2-dimensional sinograms and then reconstructed using 2-dimensional filtered backprojection with a ramp filter cutoff at half the Nyquist frequency.

For each emission sinogram, 6 reconstructions were performed differing only in which attenuation map was used for the emission photon attenuation and scatter correction. These correspond to each of the 3 attenuation maps (no correction, correction for emission contamination only, and correction for emission contamination and scatter) and transmission sources (⁶⁸Ge and ⁵⁷Co). Before computing the attenuation correction factors from data acquired using the ⁵⁷Co transmission source (with a photon

energy of approximately 122 keV), it was first necessary to convert the μ -values for the annihilation photon energy (511 keV). This conversion was done using a piecewise linear rescaling similar to the bilinear interpolation methods (*17,18*) proposed for CT-based attenuation correction. All reconstructions were corrected for emission photon scatter using the single-scatter simulation method (*11*). Calibration to absolute quantitative reconstruction units was achieved by using a 0.3-MBq ⁶⁸Ge point source and rescaling the reconstructed counts (after all appropriate corrections) to a well-counter measurement.

Possible Alternatives to Transmission Scatter Correction

To illustrate the systematic errors that can be caused by rescaling or segmenting attenuation-map data without transmission scatter correction, we have reconstructed 2 sets of experimental emission data (using Fourier rebinning filtered backprojection with all the necessary corrections for quantitative PET described in the previous section) from the microPET Focus 120. For the first experiment, we used a 30-mm-radius water cylinder filled with a uniform concentration of ¹⁸F activity with singles-mode transmission data acquired using a ⁵⁷Co source. For the second experiment, we used a 45-mmradius water cylinder also filled with a uniform concentration of ¹⁸F activity but with transmission data acquired using a ⁶⁸Ge source. These experiments were selected based on our previous work (2), which suggested that rescaling or segmentation may be problematic if the transmission data are not corrected for contamination from scattered photons. The cylinders investigated here are closer in size to a larger rat and may therefore be subject to more transmission scatter than occurs in a typical mouse study. The primary focus of these studies was to investigate the influence of transmission scatter and not emission contamination. Therefore, for both experiments, the transmission scans were performed after the ¹⁸F had decayed to negligible amounts (we estimate that, for both studies, less than 0.3% of the detected counts in the transmission data were due to emission contamination).

For the first experiment, we reconstructed the attenuation-map data with and without our scatter correction. For the attenuation-map data that had not been corrected for scatter, we also performed a slice-by-slice rescaling for the attenuation-map data so that the average reconstructed μ -value inside the water cylinder was equal to the known μ -value of water. For the second experiment, we reconstructed transmission data with and without our scatter correction using MAPtr (7), a reconstruction-based automated segmentation routine that is distributed as part of the microPET Focus software package. The MAPtr algorithm effectively segments the attenuation-map image using the a priori knowledge that the reconstructed image should contain only a limited number of materials and tissue types (each with unique linear attenuation coefficients).

RESULTS

Primate Scanner Studies

The analyses of the attenuation maps for the water cylinder and for the *Macaca nemestrina* monkey are shown in Figures 1 and 2, respectively, and described in the legends. The amounts of central-processing-unit (CPU) time required for the calculation of the scatter correction were between 17 and 26 min using a 2.2-GHz processor.



Each ordered-subsets transmission reconstruction required about 21 min of CPU time using the same processor.

Studies Using Postinjection Transmission Data

Transmission Data Reconstruction. Figures 3 and 4 show examples of transverse slices through the reconstructed attenuation-map images for the cylinder and rodent studies, respectively. The images at top in each figure are reconstructions using the ⁶⁸Ge transmission data, and the images at bottom use the ⁵⁷Co data, which have been rescaled to the appropriate values for 511-keV photons. In Supplemental Figures 1A and 1B (supplemental materials are available online only at http://jnm.snmjournals.org), we also show histograms of reconstructed μ -values for the cylinder and rodent studies, respectively. For each study, histograms are shown for the ⁶⁸Ge and ⁵⁷Co attenuation maps and for the ⁵⁷Co attenuation maps that have been rescaled for 511-keV photons. With a 2.2-GHz processor, the scatter correction FIGURE 1. (A and B) Slices through reconstructed attenuation-map images with (A) and without (B) scatter correction. Results of ROI analysis in 3 perpendicular planes (transverse, coronal, and sagittal) for each attenuation map are shown. Average value and SD for all voxels within ROIs (boundaries of which are indicated by dashed lines in images) are below each image. (C) Profiles through reconstructed data (averaged over 10 axial planes and over all data between 2 dashed black lines in transverse images) and histograms of reconstructed µ-values for both attenuation maps. Data are for water cylinder (50-mm radius) and were acquired using micro-PET Focus 220 with 57Co transmission source. In each plot, we also indicate expected true linear attenuation coefficient for water ($\mu_{water} = 0.160 \text{ cm}^{-1}$ at 122 keV (18)). SC = scatter correction.

for the cylinder and rodent studies required about 16 and 23 min of CPU time, respectively. Each ordered-subsets transmission reconstruction required about 3.5 min of CPU time using the same processor. This CPU time is about 10 times less than that required for reconstruction of the microPET Focus 220 data because of smaller sizes of the sinograms and images used for this scanner.

Emission Data Reconstruction. Supplemental Figure 2 shows transverse slices and profiles through the reconstructed activity-concentration image data for the cylinder and rodent studies. The activity image slices shown in Supplemental Figures 2A and 2B correspond to the same axial position as that shown for the attenuation-map images in Figures 3 and 4, respectively. For brevity, only the transverse images corresponding to the attenuation maps obtained using ⁵⁷Co transmission data with both types of correction are shown. The emission reconstructions and corrections required about 2 min of CPU time on the microPET Focus

FIGURE 2. (A and B) Slices through reconstructed attenuation-map images with (A) and without (B) scatter correction. Results of ROI analysis in 3 perpendicular planes (transverse, coronal, and sagittal) for each attenuation map are shown. Average value and SD for all voxels within ROIs (boundaries of which are indicated by dashed lines in images) are below each image. (C) Profiles through reconstructed data (averaged over 10 axial planes and over all data between 2 dashed black lines in transverse images) and histograms of reconstructed µ-values for both attenuation maps. Data are for primate transmission data. For this study, profile data were averaged over only 3 axial planes and pass through skull, brain, sinus, and snout. Note improved contrast between tissue and sinus for attenuation maps with scatter correction relative to that with no scatter correction. We expect that correct µ-value for soft tissue is approximately equal to $\mu_{Water} = 0.160$ cm^{-1} (18). SC = scatter correction.



acquisition and processing computer. We converted the concentration images to units of total activity per voxel (by simply multiplying by the voxel volume) and summed over all transverse planes in which either the line source or the simulated tumor was visible. We then placed circular regions of interest (ROIs) (with radii of 4.3 and 7.8 mm for the line-source and tumor ROIs, respectively) on each summed image and determined the total amounts of activity within each ROI. Table 1 compares the total reconstructed activity within the ROIs with the measured activity estimated using a well counter.

Possible Alternatives to Transmission Scatter Correction

Attenuation-Map Rescaling. Figure 5A shows profiles through the attenuation-map data for the first water cylinder experiment (30-mm radius, ⁵⁷Co transmission source): scatter-corrected, uncorrected, and uncorrected and rescaled. Figure 5B shows the corresponding data for the emission

reconstructions that were corrected using these attenuation maps. In the profiles, we also indicate the attenuation coefficient for water (μ_{water}) and the average reconstructed emission activity using the scatter-corrected attenuation map ($\lambda_{with SC}$). The location of the profiles is indicated in Supplemental Figure 3, which shows transverse attenuation and activity images for this experiment.

Attenuation-Map Segmentation. Figures 6A and 6B shows profiles through the segmented attenuation-map and reconstructed activity-concentration data, respectively, for the second water cylinder experiment (45-mm radius, ⁶⁸Ge transmission source). In the profiles, we also show the true attenuation coefficients for water and lung (μ_{water} and μ_{lung} , respectively) and the average activity ($\lambda_{with SC}$) corresponding to the scatter-corrected attenuation map. Transverse attenuation and activity images for this study are shown in Supplemental Figures 4A and 4B, respectively. The attenuation-map data were reconstructed and



segmented using transmission data with and without scatter correction.

DISCUSSION

Primate Scanner Studies

Figures 1 and 2 show that our reconstruction and scatter correction recovered linear attenuation coefficients (μ -values) within a few percentage points of their expected values for the singles-mode transmission data acquired using the microPET Focus 220. For example, with scatter correction the average reconstructed μ -values for the ROI analysis shown on the attenuation-map images were within





3% of their expected values for both the cylinder experiment and the primate study. The percentage errors in the reconstructed μ -values without scatter correction varied for different sized objects (errors were between 19% and 22% for the primate study and were about 25% for the cylinder study) and also depend on the location of the ROI (for the primate study the errors become larger closer to the monkey's body, where there is more scatter from outside the field of view).

The profiles shown in Figures 1C and 2C indicate that our scatter correction reduces the radial variation due to scatter for the uniform cylinder. The profile for the primate

FIGURE 4. Comparison of transverse attenuation-map images (averaged over 3 axial planes) reconstructed using ⁶⁸Ge (A) and ⁵⁷Co (B) postinjection transmission data acquired on microPET Focus 120. Data are for postinjection rodent study. Images correspond to transverse slice of rat's torso and include parts of its lungs and circular cross section of external tumor on left sides of images. EC = emission-contamination correction; SC = scatter correction.

TABLE 1 Comparison of Activity Within Circular ROIs for Emission Images Reconstructed Using Attenuation Maps with Different Combinations of Corrections

	Cylinder-study activity (MBq)		Rodent-study activity (MBq)	
Type of correction	⁶⁸ Ge	⁵⁷ Co	⁶⁸ Ge	⁵⁷ Co
Both emission contamination and attenuation	1.03	1.02	5.84	5.94
Only emission contamination	0.84	0.88	5.20	5.53
None	0.66	0.87	4.20	5.26

Based on well-counter measurements, we estimate that approximately 0.9 and 6.0 MBq of activity were present within ROIs for cylinder and rodent studies, respectively.

data passes through several types of tissue, including the skull, the brain, a nasal sinus, and parts of the monkey's snout. For the soft-tissue regions of this profile (brain and parts of the snout), we obtain reconstructed μ -values close to their expected values when scatter correction is applied. We also obtain better contrast (larger peak-to-valley ratios in the profiles) between reconstructed μ -values for soft tissue and sinus (air) when our scatter correction is applied.

Studies Using Postinjection Transmission Data

Transmission Data Reconstruction. For the histograms shown in Supplemental Figures 1A and 1B, we expect to

observe a peak corresponding to the reconstructed µ-value of water (or, equivalently, soft-tissue). For the cylinder study using the ⁶⁸Ge transmission data and reconstructed with no emission-contamination or scatter correction, no discernible peak can be found in the histograms (Supplemental Fig. 1A), and the attenuation-map images (Fig. 3A) are clearly distorted because of the presence of the line source (on the lower right side of the cylinder). This situation was even worse for the rodent study, in which the µ-value for almost every voxel of the ⁶⁸Ge attenuation map with no correction for emission contamination or scatter was equal to zero (Fig. 4A). This finding implies that for most lines of response the number of counts was greater in the ⁶⁸Ge transmission data than in the blank data (which would give physically incorrect negative µ-values if the nonnegativity constraint were not enforced during reconstruction). On the basis of our mockscan measurements, we estimate that the emission contamination contributed 19% and 78% of the counts in the ⁶⁸Ge transmission data for the cylinder and rodent studies, respectively. These results indicate how important emission contamination correction is for ⁶⁸Ge singles-mode transmission data.

For the ⁵⁷Co transmission data, we estimate that emission contamination contributed only 0.6% and 5.7% of the total counts for the cylinder and rodent transmission scans, respectively. It is clear that the contribution from emission photons depends on the amount of emission activity relative to the transmission source, the energy of the transmission source, and the energy discrimination settings used. Emission contamination can be reduced significantly, relative to



FIGURE 5. Profiles through transverse attenuation images (A) and activity-concentration images (B) for 30-mm-radius water cylinder using ⁵⁷Co transmission data. Profiles shown pass through center of cylinder (Supplemental Fig. 3) and were averaged over all axial planes to reduce noise and illustrate systematic effects of scatter in transmission data. Data correspond to scatter-corrected, uncorrected, and rescaled uncorrected attenuation maps. SC = scatter correction.



that in 68 Ge transmission data, simply by using energy discrimination with a lower-energy γ -emitting transmission source such as 57 Co.

Our results indicate that it is not sufficient to perform only the emission-contamination correction and that, to obtain accurate μ -values, one must also apply scatter correction. When correction was applied only for emission contamination in the cylinder study, the μ -value histogram peak positions in Supplemental Figure 1A were 36% and 30% lower than the expected values for water using the ⁶⁸Ge and ⁵⁷Co transmission data, respectively. For the attenuation maps with both corrections, however, the histogram peak positions were within 2% of their expected values for water for the cylinder study using each transmission source.

For the rodent study using the ⁵⁷Co transmission source, the peaks in the μ -value histograms in Supplemental Figure 1B were 25% (no emission contamination correction) and 16% (emission contamination correction only) lower than the expected value for soft tissue when transmission scatter correction was not applied. When both corrections were applied, the histogram peak value was within 1% of its expected value for the ⁵⁷Co transmission data. The attenuation maps with emission contamination corrections) for the rodent study using the ⁶⁸Ge transmission data were extremely noisy and no discernible peaks could be found in either of their μ -value histograms. We performed an ROI analysis (data not shown) in 3 perpendicular planes (transverse, coronal, and sagittal) for the attenuation maps recon-



structed from the ⁶⁸Ge transmission data using emission contamination correction only and both corrections. We found that, despite their poor image quality, the images reconstructed using both corrections had average μ -values within $\pm 13\%$ of their expected values for soft-tissue ROIs. The average values, for the same ROIs placed on the images corrected for emission contamination only, were between 30% and 34% lower than the expected value.

For the postinjection transmission data analyzed here, the 68 Ge and 57 Co blank data (which had the same duration as the transmission data) happened to have approximately the same average number of counts (about 350 per bin) in each bin of the single-slice rebinned sinogram data. This finding suggests that the improved quality of the 57 Co images is largely due to the lower amounts of emission contamination relative to that for the 68 Ge data and the better contrast between μ -values at lower energy.

The piecewise linear rescaling we have used here correctly converts the linear attenuation coefficients for water from the values expected for 122-keV photons to those for 511-keV photons. For the histograms shown in Supplemental Figures 1A and 1B, the water peaks are correctly shifted to the appropriate μ -values for the attenuation maps with both types of corrections. Based on values given by Berger et al. (*19*), we estimate that the effective linear attenuation coefficient for the line-source material is approximately 0.27 cm⁻¹ at 511 keV. We placed ROIs inside the vicinity of the line source for the ⁵⁷Co attenuation map (applying both corrections and rescaling for 511-keV

photons) and found that the average reconstructed $\mu\text{-values}$ were between 0.25 and 0.28 $\text{cm}^{-1}.$

Emission Data Reconstruction. The profiles shown in Supplemental Figures 2C and 2D indicate that the reconstructed activity concentrations are strongly influenced by the correction applied during the attenuation-map reconstructions. If we examine the results of the ROI analysis summarized in Table 1, we find that the total activities for the emission images reconstructed using only emissioncontamination correction were between 7% and 18% lower than the corresponding values for the images with both corrections. The total reconstructed activities for the images with no correction were between 35% and 15% lower than those for the images with both corrections. We also found that the total activity within the ROIs for each study were consistent (agreed to within 2%) for images reconstructed using either ⁶⁸Ge or ⁵⁷Co, provided that both emissioncontamination and scatter corrections were applied.

If we compare the total reconstructed activities in Table 1 with the total activities estimated from the well-counter measurements, we find that values for the images with both corrections were within about 3% of the measured values for the rodent study but were overestimated by about 14% for the line-source and cylinder data. However, the well counter measurement for the pointlike tumor used for the rodent study is subject to less uncertainty than that of the line source. There are systematic errors associated with measuring the activity of an extended line source using a well counter, because it is calibrated for point sources centered in the bottom of the well. For example, we have observed that the measured activity for a pointlike source can be as much as 6% lower when the source is located 30-40 mm above the bottom of the well counter, compared with when it is centered in the bottom of the well. Because the line source is 115 mm long, the well-counter measurement for its activity may also be underestimated. Another source of uncertainty occurs because the well-counter measurement for the line source is subject to photon attenuation, whereas the reconstructed image has been corrected for attenuation. In the activity values given in the legend of Table 1, we have compensated for this effect by multiplying the well-counter measured activity by an approximate attenuation correction factor. We estimate that 9% of photons are attenuated in the line-source material, assuming that all photons are emitted from the center of the line source in a plane perpendicular to the length of the line source. It is possible that we have underestimated the attenuation effect, because photons emitted from the center of the line source in any nonperpendicular plane will be subject to more photon attenuation.

Possible Alternatives to Transmission Scatter Correction

Figures 5 and 6 and Supplemental Figures 3 and 4 show that the erroneous radial variation, which is visible in attenuation-map data that have not been corrected for scatter, causes systematic errors in the reconstructed emission images. Figure 5 indicates that rescaling the noncorrected attenuation maps does not eliminate the radial variation and that these errors propagate into the reconstructed emission images. The most uniform activityconcentration profiles were obtained using our scatter correction method. Figure 6 indicates that this radial variation due to scatter in the attenuation-map images also can lead to errors in automated segmentation routines. In the uncorrected attenuation-map profiles and images shown in Figure 6A and Supplemental Figure 4A, many voxels are incorrectly assigned the linear attenuation coefficient for lung tissue. As we can see from the activity profiles and images shown in Figure 6B and Supplemental Figure 4B, this type of segmentation error causes an underestimation of the reconstructed activity for regions lying deeper inside the object (especially for regions incorrectly assigned the μ -value for lung) relative to those closer to the surface. This is not meant to be a criticism of the segmentation method of Nuyts et al. (7) but is instead an indication of the importance of accurate scatter correction for singles-mode transmission data. When our scatter correction is applied, these segmentation errors do not occur (all voxels are assigned the correct μ -value for water).

We have found in the current and our previous work (2)that errors in the reconstructed µ-values without scatter correction can vary widely depending on the size of the subject, on the transmission source, and on the radial and axial positions within each attenuation-map image. Transmission scatter is most significant for subjects that are large relative to the PET scanner ring diameter (primates or heavier rats) and for regions of highly nonuniform attenuation (torso or whole-body imaging). Even for small animals such as mice, however, we have found that transmission scatter correction can be important (e.g., without scatter correction the reconstructed attenuation coefficients for a mouse were 19% lower than the expected value for soft tissue (2)). These results suggest that, for animal studies, the errors that could be caused by using rescaled or segmented attenuation maps without scatter correction may be difficult to predict and could result in misleading reconstructed emission images and errors in the biologic parameters derived from them. For example, the measurement of neuroreceptor binding potentials in small-animal PET can be particularly sensitive to attenuation errors, because this measurement depends critically on the relative amounts of striatal and cerebellar activity (20). In rats, the cerebellum is in a region subject to higher attenuation (and, as a result, higher transmission photon scattering) than is the striatum. The rescaling and segmentation errors for attenuation maps without scatter correction, as illustrated in Figures 5 and 6 and Supplemental Figures 3 and 4, would lead to errors in this type of activity ratio.

CONCLUSION

Our results indicate that our reconstruction and scatter correction procedure can easily be adapted to different PET scanners (primate and rodent models of the microPET Focus) and different imaging protocols (pre- and postinjection data acquisition). For all cases, the average reconstructed µ-values with scatter correction were within 3% of the expected values for water and soft tissue for both phantom and animal studies. Without scatter correction (and emission contamination correction for postinjection transmission), the reconstructed µ-values were biased and varied widely depending on the size of the object, the local position within the object, the amount of emission activity within the scanner, and the type of transmission source. For the postinjection transmission data acquired using the microPET Focus 120, we found that the accuracy of reconstructed emission activity concentrations depends strongly on the scatter and emission-contamination correction applied to the singles-mode transmission data. We also found that our scatter correction algorithm is more accurate than the existing alternatives (segmentation or rescaling of uncorrected attenuation maps).

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