

METHODOLOGIC ASPECTS OF MYOCARDIAL BLOOD FLOW QUANTIFICATION WITH 1-¹¹C-ACETATE PET

TO THE EDITOR: I have read with interest the article of Sciacca et al. (1) reporting on a comparison of H₂¹⁵O and 1-¹¹C-acetate for quantification of myocardial blood flow with PET. The aim of this study, to contribute to the validation of 1-¹¹C-acetate as a quantitative flow tracer in myocardial PET, is certainly important. However, the article exhibits methodologic problems concerning tracer kinetic modeling as well as correction of partial-volume effects.

The authors assume the rate of tissue clearance to be equal to perfusion, F (Fig. 1; Eqs. 1 and 2 (1)). This assumption is not justified: It is known from animal data (2) that within approximately 15 s after passage of a very short intraarterially injected bolus, tracer concentration drops to an approximately constant plateau of about 2- to 3-min duration (the initial drop corresponding to the finite first-pass extraction). Tissue clearance occurs only after this plateau phase and is numerically an order of magnitude smaller than F . Therefore, postulating a clearance rate equal to F seems inconsistent with the actual behavior of acetate, and parameter estimation bias can be expected. It would actually be less problematic to neglect tissue clearance altogether during the first 3 min. It should be noted, however, that restricting data evaluation to this time range leads to substantial loss of statistical accuracy of the perfusion estimates in comparison with using longer fitting intervals. I believe, therefore, that a 1-compartment model without parameter constraints (i.e., setting K_1 to zero and allowing for arbitrary clearance rates in Eqs. 1 and 2 (1)) would be a more adequate model. Such a model has the particular advantage of being able to fit the data over much longer time intervals (3,4).

Furthermore, I consider the described method for recovery correction to be questionable. Essentially, the F_{MM} (which can be identified with the recovery coefficients if fractional blood volume is neglected) are adjusted in such a way that the flow values derived with H₂¹⁵O and 1-¹¹C-acetate coincide if the individually adapted F_{MM} are used. The good agreement of the perfusion values is, therefore, no proof of adequacy of the recovery correction method. Rather, the derived F_{MM} empirically correct for all other sources of errors, such as model configuration and assumption of a constant extraction fraction. This conclusion is substantiated as follows. The general solution of Equations 1–3 (1) can be written as:

$$Q_{\text{tissue}}(t) = (F_{MM} \cdot E \cdot F) \times g(t) + F_{BM} \cdot C_a(t),$$

where E is the unidirectional extraction fraction, $g(t)$ describes the shape of the tissue signal, F_{BM} includes both spillover of counts from blood in the adjacent left ventricular cavity and counts from

the fractional blood volume within the tissue region of interest, and $C_a(t)$ is the tracer concentration in arterial blood. The amplitude of $g(t)$ is given by:

$$A = F_{MM} \cdot E \cdot F. \quad \text{Eq. 1}$$

This amplitude is an easily identifiable parameter, but as long as arbitrary tissue clearance is allowed for (no influence of F on $g(t)$), it is impossible to identify the individual factors contributing to A . Even under the assumptions used by Sciacca et al. (1) (i.e., clearance equal to flow) it is still impossible to differentiate between F_{MM} and E .

Because Sciacca et al. (1) found relatively large discrepancies in the flow estimates obtained with H₂¹⁵O and 1-¹¹C-acetate when using fixed values for E and F_{MM} , they describe a procedure for improving the recovery correction, which essentially is equivalent to solving Equation 1 for F_{MM} using the flow value determined in the H₂¹⁵O investigation for F . Correlating these individual adapted F_{MM} to echocardiographic findings is then performed to enable individual recovery correction. Thus, agreement of flow values derived with 1-¹¹C-acetate and H₂¹⁵O is enforced by the method.

Two other questions come immediately to mind in this context. Why did the authors not use the individual F_{MM} that have been apparently derived from the corresponding H₂¹⁵O scans? How do the F_{MM} derived by the authors' method compare with those obtained from the H₂¹⁵O scans?

In conclusion, I think that the data of Sciacca et al. (1) make a valuable contribution to the ongoing efforts to validate 1-¹¹C-acetate as a quantitative flow tracer, but the methods chosen to evaluate the acetate investigations seem to be inadequate. A modification of the model configuration, in combination with an implicit recovery correction as proposed, for instance, by Hutchins et al. (5), seems to be desirable.

REFERENCES

1. Sciacca RR, Akinboboye O, Chou RL, Epstein S, Bergmann SR. Measurement of myocardial blood flow with PET using 1-¹¹C-acetate. *J Nucl Med.* 2001;42:63–70.
2. Armbricht JJ, Buxton DB, Schelbert HR. Validation of [1-¹¹C]acetate as a tracer for noninvasive assessment of oxidative metabolism with positron emission tomography in normal, ischemic, posts ischemic, and hyperemic canine myocardium. *Circulation.* 1990;81:1594–1605.
3. Buck A, Wolpers HG, Hutchins GD, et al. Effect of carbon-11-acetate recirculation on estimates of myocardial oxygen consumption by PET. *J Nucl Med.* 1991;32:1950–1957.
4. van den Hoff J, Burchert W, Wolpers HG, Meyer GJ, Hundeshagen H. A kinetic model for cardiac PET with [1-carbon-11]-acetate. *J Nucl Med.* 1996;37:521–529.
5. Hutchins GD, Caraher JM, Raylman RR. A region of interest strategy for minimizing resolution distortions in quantitative myocardial PET studies. *J Nucl Med.* 1992;33:1243–1250.

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