EDITORIAL Myocardial Metabolism of Radioiodinated BMIPP

n this issue of The Journal of Nu-Lclear Medicine, Yamamichi et al. (1) describe the isolation and identification of catabolites from the outflow of a recirculating isolated rat heart system following administration of radioiodinated 15-(p-iodophenyl)-3-R,S-methylpentadecanoic acid (BMIPP) and the effects of perfusate substrates on metabolite formation. BMIPP is a modified fatty acid analogue of [15-(p-iodophenyl-) pentadecanoic acid (IPPA) in which a methyl group has been introduced into the beta (3)-position of the fatty acid chain to inhibit rapid myocardial catabolism (2,3). Increasing interest in the clinical use of ¹²³I-labeled fatty acids for myocardial imaging is illustrated by the broad use of 123 I-BMIPP in Japan, following its commercial introduction as Cardiodine by Nihon Medi-Physics Co., Inc., in 1993. The identification of the metabolites released from the myocardium described by Yamamichi et al. is important in further expanding our understanding of the BMIPP metabolism and the factors which may affect catabolism and myocardial washout of this important cardiac imaging agent.

Clinical use of ¹²³I-labeled modified fatty acid analogs which show delayed myocardial clearance for myocardial SPECT imaging is based on differences between regional myocardial fatty acid uptake patterns and flow tracer distribution which are often observed in various myocardial disorders (2,3). The use of ¹²³I-BMIPP in conjunction with flow tracers may represent a unique opportunity to correlate energy substrate metabolism with regional myocardial viability using SPECT. Although the physiological basis is not completely understood, differences between regional fatty acid and flow tracer distribution may indirectly reflect alterations in important parameters of metabolism which can be important for clinical decision making.

RATIONALE FOR DEVELOPING IODINE-123-LABELED METHYL-BRANCHED FATTY ACIDS

Naturally occurring, long-chain fatty acids (palmitic, oleic, etc.) are the principal energy source for the normoxic myocardium, and thus, radioiodinated fatty acid agents represent potential probes to evaluate differences in cardiac oxidative metabolism which are present in various myocardial disorders. For cardiac SPECT, especially with single- and double-headed cameras, significant tracer redistribution during the acquisition period must be minimized during the relatively long time periods required for camera rotation and data collection. Since straight chain fatty acids are rapidly metabolized by beta-oxidation, a variety of structural modifications have been introduced into fatty acid analogs to evaluate their inhibition of beta-oxidation, thus delaying myocardial tracer clearance and relative regional redistribution. In particular, methyl-branched fatty acids such as BMIPP have been widely investigated (2-6). The effects of methyl-substitution in delaying myocardial tracer clearance is based on the expected inhibition of the beta-oxidative process at the stage of obligatory NAD-dependent oxidation of the betahydroxy intermediate catalyzed by beta-hydroxyacyl-CoA dehydrogenase.

During the normal oxidative catabolism of aliphatic (straight chain) fatty acids, oxidation forms the beta-ketoacyl-CoA intermediate, which is the substrate for thiolase-catalyzed alpha, betacarbon-carbon bond cleavage with the release of the acetyl CoA two-carbon fragment. The chain length of the original fatty acid substrate is thus decreased by two carbon atoms and the resulting acyl-CoA product can then be recycled through the beta-oxidative chain.

The potential clinical use of radioiodinated fatty acids for cardiac imaging was first demonstrated in 1965 with 131 I-labeled iodo-oleic acid (7). For nearly 30 vr. use of radiolabeled fatty acids for myocardial imaging has fascinated many investigators (2-6). Development of the BMIPP analog at the Oak Ridge National Laboratory (ORNL) with radioiodine stabilized by attachment to the para-position of the terminal phenyl ring (8-11) was based on introduction of 3-methylbranching into IPPA. The inhibitory effect of the 3-methyl group of BMIPP on beta-oxidation results in longer myocardial retention, which has been demonstrated in extensive in vivo studies in laboratory animal species and, more recently, in humans (2,3).

Oxidative Products Formed from BMIPP

The results of Yamamichi et al. have for the first time identified the metabolites released from the myocardium following administration of BMIPP. As proposed earlier (12-15), the 3-methyl group is expected to interfere with betadation, but this impediment could be removed by initial alpha-oxidation, and the resulting alpha-methyl-substituted metabolite could then serve as a substrate for the usual beta-oxidative pathway. Consistent with this expectation, the principal metabolite identified by Yamamichi et al. is the initial alpha-oxidation product 14-(p-iodophenyl)-2(α)-R,S-methyltetradecanoic acid (AMIPT).

Other metabolic products which were also identified by comparison with synthetic standards are 12-(p-iodophenyl)dodecanoic acid (PIPC₁₂) and 2-(p-iodophenyl)acetic acid (PIPA). BMIPP is thus apparently metabo-

Received Dec. 16, 1994; accepted Feb. 14, 1995. For correspondence or reprints contact: F.F. (Russ) Knapp, J., PhD, Group Leader, Nuclear Medicine Group, Oak Ridge National Laboratory, PO Box 2008, Oak Ridge, TN, 37831-6229. E-mail JKP @orml.gov.

lized, as had been expected, by initial alpha-oxidation in which the carboxyl carbon is lost. Since the impediment of the beta-methyl substituent has been removed, the alpha-methyl-substituted CoA fragment can then pass through the beta-oxidative chain, with p-iodophenylacetic acid being formed as the final oxidative product, which is a process similar to the oxidative metabolism of phytanic acid which is inhibited in Refsum's disease (9). An important aspect of this scenario which is vet unanswered, however, is the intracellular dynamics of the activated BMIPP species, since it has not vet been established if initial alphaoxidation occurs in the mitochondrial compartment. Initial alpha-oxidation of BMIPP CoA in an extramitochondrial compartment would be expected to require subsequent transfer via the carnitine shuttle through the mitochondrial membrane for beta-oxidative catabolism.

Earlier studies had used both nonworking (13, 14) and working (15) rat hearts to isolate an unidentified polar metabolite from the outflow of Langendorff preparations. In these studies, only one radioactive component was observed by thin-layer (TLC) and high-performance liquid chromatographic (HPLC) studies, and the same single metabolite was also observed in plasma samples from humans following intravenous administration of [¹²³I]BMIPP (2, 3, 16, 17). Although the identification of only one polar metabolite in earlier isolated heart studies, but several catabolites by Yamamichi et al. may result from different experimental conditions, this does not explain the single component observed in earlier patient plasma analysis (16,17). It is important to further extend the metabolite studies now that authentic samples are available.

The studies by Yamamichi et al. are important since they provide one missing link in our understanding of the intracellular metabolism of BMIPP and the identity of the catabolic species which immediately leaves the cell. Although the intracellular accumulation of BMIPP metabolites was not evaluated by Yamamichi et al. an important observation of earlier work (13-18) was that the oxidative catabolic products from BMIPP do not accumulate intracellularly and evidently immediately leave the cell after formation. Thus, the oxidative catabolites from BMIPP do not accumulate in myocytes and if present in myocardial lipid extracts must be present in very small quantities.

Intracellular Metabolism of BMIPP

Although the oxidative catabolite(s) of BMIPP had not been identified until the work by Yamamichi et al., the identity of the complex lipid products accumulating in the myocytes from metabolism of BMIPP are well understood and have been studied in detail (2,3). High-performance liquid chromatography has been used to examine the incorporation of radioiodinated BMIPP and IPPA into intracellular phospholipid fractions of myocardial lipids isolated from both rats and dogs. Structural modifications have unexpected effects on fatty acid metabolism. The IPPA straight-chain analog is incorporated into lecithin (phosphatidylcholine) in rat hearts, while BMIPP is primarily found in the cephalin (phosphatidylethanolamine) fraction (17). Physiological factors which affect myocardial uptake of BMIPP have been widely studied (2,3) and include the effects of ATP (19.20).

The correlation between myocardial ATP levels and BMIPP uptake could be especially important and this relationship has been investigated in normal and salt-sensitive Dahl-strain rats following administration of an inhibitor of mitochondrial carnitine acyltransferase I (tetradecylglycidic acid (TDGA) (20). Initial myocardial BMIPP uptake was not influenced by acute inhibition of beta-oxidation by the inhibitor. Intracellular ATP levels correlating with BMIPP retention are interpreted to reflect cystolic activation of BMIPP to BMIPP-CoA with slow shunting into triglyceride storage products. In the hypertrophied hearts of Dahl rats, a negative correlation was observed between ATP levels and

severely compromised myocardial **BMIPP.** Differences in mitochondrial and cytosolic ATP pools may explain these results, but additional studies are required to more fully evaluate the relationship between high-energy phosphate levels and BMIPP retention. Studies have pursued evaluation of the influence of methyl-substitution on the myocardial metabolism of BMIPP in rat hearts in vivo (21) and isolated Langendorff-perfused rat hearts (13-15), which demonstrated incorporation of BMIPP into triglyceride storage products correlating with the observed myocardial retention.

Mismatch of Iodine-123-BMIPP and Flow Tracer Myocardial Distribution

The apparently unique, yet not well understood, property of BMIPP is the mismatch often observed between the regional distribution of this agent and flow tracers, although it has not vet been established if these differences are even related to the intracellular oxidative metabolism of BMIPP. Initial studies by Yonekura et al. and Yamamoto et al. clearly demonstrated differences in BMIPP and flow tracer distribution in the hearts of hypertensive animals using high-resolution autoradiographic techniques (22,23), and these important studies have relevance to the clinical applications of [¹²³I]BMIPP.

Animal studies with radioiodinated BMIPP have also evaluated the regional myocardial distribution of this tracer in various cardiac disease models and paved the way for subsequent human studies. Significant differences have been found between flow tracer (²⁰¹Tl) and methyl-branched fatty acid distribution in the free wall of the left ventricle and septal regions of hearts from hypertensive rats (22, 23) and rat and hamster models with hypertrophic and cardiomyopathic heart disease (24,25). BMIPP also has been used to evaluate cocaine-induced regional myocardial metabolic changes in hypertensive rats by comparison of regional perfusion of ²⁰¹Tl with differences in 2-deoxyglucose (2-DG) and radioiodinated BMIPP uptake (26, 27).

Early studies have also evaluated the properties of radioiodinated BMIPP in an ischemic canine model (28) and have shown differences in BMIPP versus ²⁰¹Tl uptake in border zone regions.

The uptake and clearance of ¹²³I-BMIPP has been recently studied by planar imaging in a canine occlusionreperfusion model (29) using ex vivo gamma camera imaging of the excised hearts.

The results from these extensive animal studies with radioiodinated BMIPP have provided an important foundation for humans studies now in progress. Only the global release of catabolites from rats in vivo or isolated heart preparations has been studied and the slow myocardial washout observed after administration of radioiodinated BMIPP in vivo or with isolated heart preparations represents only a small proportion of the total uptake. Regional mismatches in BMIPP uptake in comparison with flow tracer distribution may thus be expected to result from other factors unrelated to oxidative catabolism. Furthermore, the factors resulting in the mismatch phenomena between BMIPP and flow tracers are probably unrelated to the formation of oxidative products. They are probably related to differences in cellular uptake in comparison to normal myocytes and those cells which are viable but in which fatty acid metabolism is somehow impaired.

SUMMARY

After significant efforts over the last 30 yr, the use of ¹²³I-labeled fatty acids for routine assessment of cardiac disease may now be a reality with ¹²³I-BMIPP. The use of ¹²³I-labeled fatty acids may provide complementary information on myocardial viability, for example, identifying and assessing the presence of salvageable tissue. The results of these extensive clinical trials often report the mismatch between BMIPP and flow tracer distribution. But why? It will be important to continue the basic studies such as those reported by Yamamichi et al. to obtain a complete understanding of the metabolism and physiological factors affecting myocardial uptake of BMIPP.

Factors requiring additional study in animal models include biochemical and histological analysis of biopsy segments from myocardial regions which have decreased fatty acid relative to flow tracer uptake. An important issue for evaluation is the relative metabolism of the isomers of BMIPP. since BMIPP is a racemic mixture of the 3R-methyl and 3S-methyl isomers. It is certainly possible that only one isomer is catabolized to the metabolites identified by Yamamichi et al. and that the second isomer could be metabolically inactive. In addition to an evaluation of the relative intracellular metabolism of both isomers, the kinetics of tissue uptake and release of metabolites of the 3R- and 3S-methyl isomers of radioiodinated BMIPP should be evaluated. It is conceivable for instance, that one isomer may have less nontarget tissue uptake, in comparison with racemic 3-(R,S)-BMIPP.

Clinical protocols are being pursued at several institutions in western Europe (17, 30-32), and ¹²³I-labeled BMIPP (Cardiodine) is currently widely used in Japan as an approved radiopharmaceutical for the evaluation of impairments of myocardial fatty acid metabolism and for myocardial viability (6, 33-35). More than 2500 patient studies per month are currently conducted with Cardiodine in Japan in over 300 hospital-based nuclear medicine departments. This is estimated to represent about 14% of all nuclear cardiology studies in Japan. Through December 1994 more than 50,000 patient studies had been completed.

The results of these studies, in conjunction with clinical studies being pursued at several institutions in Europe, will hopefully stimulate the use of Cardiodine in the United States. We expect that the extensive data being documented on the use of ¹²³I-BMIPP in various myocardial disorders and the comparison of these results with commonly used flow tracers will answer these questions and provide the basis by which the broader possible role of BMIPP in nuclear cardiology will be further assessed. Lipid analyses, after either coronary sinus sampling or studies of serial blood samples, obtained from patients following intravenous administration of ¹²³I-BMIPP may confirm that these same metabolites identified in the rat heart studies reported by Yamamichi et al. are formed in humans.

F.F. (Russ) Knapp, Jr. Nuclear Medicine Group Oak Ridge National Laboratory (ORNL) Oak Ridge, Tennessee

ACKNOWLEDGMENTS

Research at ORNL is supported by the Office of Health and Environmental Research, U.S. Department of Energy, under contract DE-AC05-85OR21400 with Martin Marietta Energy Systems, Inc. The author also thanks his many colleagues who have provided stimulating discussions on the clinical use of ¹²³I-BMIPP.

REFERENCE9

- Yamamichi Y, Hideo BS, Kusuoka H, et al. Metabolism of ¹²³I-labeled 15-(p-iodophenyl)-3-(R,S)-methylpentadecanoic acid (BMIPP) in perfused rat hearts: the evidence for initial α-oxidation and subsequent cycles of β-oxidation, and dependency on substrates. J Nucl Med 1995:36:1043-1050.
- Knapp Jr FF, Kropp J. Iodine-123-labeled fatty acids for myocardial SPECT: current status and future perspectives. *Eur J Nucl Med* 1995; 22: 361-381.
- Knapp Jr FF, Kropp J, Goodman MM, et al. The development of iodine-123-methylbranched fatty acids and their applications in nuclear cardiology. Ann Nucl Med 1993;7:1-14.
 European Heart Journal. 1985;6:1-106 [Entire
- issue]. 5. European-American Communications on Nu-
- clear Medicine 1990;21:201-266 [Entire issue]. 6. Annals of Nuclear Medicine 1993;7:S-II;1-116 [Entire issue].
- Evans JR, Gunton RW, Bakel RG, et al. Use of radioiodinated fatty acids for photoscans of the heart. *Circ Res* 1965;16:1-10.
- Knapp Jr FF, Goodman MM, Ambrose KR, et al. The development of radioiodinated 3-methyl-branched fatty acids for evaluation of myocardial disease by single-photon tomography. In: van der Wall EE, ed. Noninvasive measurement of cardiac metabolism, Amsterdam: Martinus Nijhoff Publishers; 1987:159-202.
- Knapp Jr FF, Goodman MM. The design and biological properties of iodine-123-labeled β-methyl-branched fatty acids. Eur Heart J 1985;6:71-84.
- 10. Goodman MM, Kirsch G, Knapp Jr FF. Synthesis and evaluation of radioiodinated terminal

p-iodophenyl-substituted α - and β -methylbranched fatty acids. *J Med Chem* 1984;27:390– 397.

- Knapp Jr FF, Ambrose KR, Goodman MM. New radioiodinated methyl-branched fatty acids for cardiac imaging. *Eur J Nucl Med* 1986; 12:S539–S544.
- Knapp Jr FF, Goodman MM, Callahan AP, Kirsch G. Radioiodinated 15-(p-iodophenyl)-3,3-dimethylpentadecanoic acid: a useful agent to evaluate myocardial fatty acid uptake. J Nucl Med 1986;27:521-531.
- Knapp Jr FF, Goodman MM, Reske SN, et al. Radioiodinated methyl-branched fatty acidsevaluation of catabolites formed in vivo. Nuc-Compact/Eur Am Commun Nucl Med 1990;21: 229-231.
- Knapp Jr FF, Reske SN, Ambrose KR, et al. Formation of polar catabolites from radioiodinated 15-(p-iodophenyl)-3-R,S-methylpentadecanoic acid (BMIPP) by isolated Langendorff rat hearts. NucCompact/Eur Am Commun Nucl Med 1990;21:133–139.
- Kropp J, Knapp Jr FF, Ambrose KR, et al. Release of an unexpected myocardial metabolite of radioiodinated 15-(p-iodophenyl)-3-R,Smethylpentadecanoic acid (BMIPP) from isolated rat hearts and canine hearts in vivo [Abstract]. J Nucl Med 1990;31:896.
- Dudczak R, Schmoliner R, Angelberger P, et al. Structurally-modified fatty acids: clinical potential as tracers of metabolism. *Eur J Nucl Med* 1986;12:45-48.
- Kropp J, Kohler U, Knapp Jr FF, Biersack JH. 15-(p-[I-123]iodophenyl)-3-R,S-methylpentadecanoic acid to evaluate ischemia in patients with coronary artery disease. *Eur J Nucl Med* 1991; 18:650.
- Kropp J, Ambrose KR, Knapp Jr FF, et al. Evaluation of the incorporation of IPPA and BMIPP into complex lipids from isolated rat hearts by high performance liquid chromatographic analysis (HPLC). Nucl Med Biol 1992; 19:283-288.
- 19. Fujibayashi Y, Yonekura Y, Takemura Y, et al.

Myocardial accumulation of iodinated beta-methyl-branched fatty acid analogue, iodine-125-15-(p-iodophenyl)-3-(R,S)-methylpentadecanoic acid (BMIPP), in relation to ATP concentration. *J Nucl Med* 1990;31:1818–1822.

- Fujibayashi Y, Yonekura Y, Tamaki N, et al. Myocardial accumulation of BMIPP in relation to ATP concentration. *Ann Nucl Med* 1993;7: 15-18.
- Ambrose KR, Owen BA, Goodman MM, et al. Evaluation of the metabolism in rat hearts of two new radioiodinated 3-methyl-branched fatty acid myocardial imaging agents. *Eur J Nucl Med* 1987;12:486-491.
- 22. Yonekura Y, Brill AB, Som P, et al. Quantitative autoradiographic measurement of regional myocardial substrate utilization in hypertensive rats. *Science* 1985;227:1494–1496.
- 23. Yamamoto K, Som P, Brill AB, et al. Dual tracer autoradiographic study of β-methyl-(10¹⁴C) heptadecanoic acid and 15-p-(¹³¹I)-iodophenyl-β-methylpentadecanoic acid in normotensive and hypertensive rats. J Nucl Med 1986;27:1178-1183.
- Kurata C, Kobayashi A, Yamazaki. Dual tracer autoradiographic study with thallium-201 and radioiodinated fatty acid in cardiomyopathic hamsters. J Nucl Med 1989;30:80-87.
- Som P, Oster ZH, Goodman MM, et al. Microimaging studies with myocardial substrate utilization and perfusion in two models of non-coronary heart disease. NucCompact/Eur Am Commun Nucl Med 1990;21:259-262.
- Som P, Wang G-J, Oster ZH, et al. Myocardial uptake of cocaine and effects of cocaine on myocardial substrate utilization and perfusion in hypertensive rats. *Ann Nucl Med* 1993;7:19– 26.
- Wang G-J, Som P, Oster ZH, et al. Quantitative autoradiographic measurement of cocaine-induced regional myocardial metabolic changes in hypertensive rats. *Nucl Med Biol* 1994;21:245– 250.
- 28. Miller DD, Gill JB, Livni E, et al. Fatty acid

analogue accumulation: a marker of myocyte viability in ischemic-reperfused myocardium. *Circ Res* 1988;63:681-693.

- 29. Nishimura T, Sago M, Kihara K, et al. Fatty acid myocardial imaging using ¹²³I-β-methyl- iodophenyl pentadecanoic acid (BMIPP): comparison of myocardial perfusion and fatty acid utilization in canine myocardial infarction (occlusion and reperfusion model). *Eur J Nucl Med* 1989;15:341-345.
- 30. Kropp J, Juergans M, Glaenzer K, et al. Evaluation of ischemia and myocardial viability in patients with coronary artery disease (CAD) with iodine-123-labeled (15-(p-iodophenyl)-3-R,S-methylpentadecanoic acid (BMIPP). Ann Nucl Med 1993;7:93-100.
- DeGeeter FF, Franken P, Knapp Jr FF, Bossuyt A. Relationship between blood flow and fatty acid metabolism in subacute myocardial infarction: a study by means of Tc-99msestamibi and iodine-123-beta-methyl iodophemylpentadecanoic acid. Eur J Nucl Med 1994; 21:283-291.
- 32. Franken PR, De Geeter F, Dendale P, et al. Abnormal free fatty acid uptake in subacute myocardial infraction after coronary thrombolysis: correlation with wall motion and inotropic reserve. J Nucl Med 1994;35:1758-1765.
- 33. Tamaki N, Kawamoto M, Yonekura Y, et al. Regional metabolic abnormality in relation to perfusion and wall motion in patients with myocardial infarction: assessment with emission tomography using iodinated branched fatty acid analogue. J Nucl Med 1992;33:659-667.
- 34. Tamaki N, Kawamoto M, Yonekura Y, et al. Assessment of fatty acid metabolism using I-123 branched fatty acid: comparison with positron emission tomography. Ann Nucl Med 1993;7: 41-48.
- 35. Kawamoto M, Tamaki N, Yonekura Y, et al. Combined study with I-123 fatty acid and thallium-201 to assess ischemic myocardium: comparison with thallium redistribution and glucose metabolism. Ann Nucl Med 1994;8:847-854.