

- kocytes and indium-111-labeled human nonspecific immunoglobulin G: a prospective comparative study. *J Nucl Med* 1991;32:1854-1860.
5. Fischman AJ, Rubin RH, Khaw BA, et al. Detection of acute inflammation with ¹¹¹In-labeled nonspecific polyclonal IgG. *Semin Nucl Med* 1988; 18:335-344.
 6. Rubin RH, Young LS, Hansen P, et al. Specific and nonspecific imaging of localized Fisher immunotype 1 pseudomonas aeruginosa infection with radiolabeled monoclonal antibody. *J Nucl Med* 1988;29:651-656.
 7. Morrel EM, Tompkins RG, Fischman AJ, et al. Autoradiographic method for quantitation of radiolabeled proteins in tissues using indium-111. *J Nucl Med* 1989;30:1538-1545.
 8. Oyen WJG, Claessens RAMJ, Raemaekers JMM, de Pauw BE, van der Meer JWM, Corstens FHM. Diagnosing infection in febrile granulocytopenic patients with indium-111 labeled human IgG. *J Clin Oncol* 1992;in press.
 9. Hnatowich DJ, Childs RL, Lantaigne D, Najafi A. The preparation of DTPA-coupled antibodies radiolabeled with metallic radionuclides: an improved method. *J Immunol Meth* 1983;65:147-157.
 10. Fraker PJ, Speck JC. Protein and cell membrane iodinations with a sparingly soluble chloramide, 1,3,4,6-tetrachloro-3 α , 6-dephenylglycoluril. *Biochem Biophys Res Comm* 1978;80:849-857.
 11. Jain NC. Blood volume and water balance. In: Jain NC, ed. *Schalm's veterinary hematology*, fourth edition. Philadelphia: Lea & Febiger; 1986:91.
 12. Gorter A, Hiemstra PS, van der Voort EAM, van Es LA, Daha HR. Binding of human IgA1 to rat peritoneal macrophages. *Immunology* 1988; 69:207-212.
 13. Underdown B, Schiff JM. Immunoglobulin A: strategic defense initiative at the mucosal surface. *Ann Rev Immunol* 1986;4:389-417.
 14. Fischman AJ, Rubin RH, White JA et al. Localization of Fc and Fab fragments of nonspecific polyclonal IgG at focal sites of inflammation. *J Nucl Med* 1990;31:1199-1205.
 15. Buijs WCAM, Oyen WJG, Claessens RAMJ, Koenders EB, Meeuwis APW, Corstens FHM. Biodistribution and radiation dosimetry of indium-111 labeled immunoglobulin G. [Abstract] *Eur J Nucl Med* 1990;16:433.
 16. Rubin RH, Fischman AJ, Needleman M, et al. Radiolabeled, nonspecific, polyclonal human immunoglobulin in the detection of focal inflammation by scintigraphy: comparison with gallium-67-citrate and technetium-99m-labeled albumin. *J Nucl Med* 1989;30:385-389.
 17. Buscombe JR, Lui D, Ensing G, de Jong R, Ell PJ. Tc-99m-human immunoglobulin (HIG)—first results of a new agent for the localization of infection and inflammation. *Eur J Nucl Med* 1990;16:649-655.
 18. Abrams MJ, Juweid M, ten Kate CI et al. Technetium-99m-human polyclonal IgG radiolabeled via the hydrazino nicotinamide derivative for imaging focal sites of infection. *J Nucl Med* 1990;31:2022-2028.
 19. Calame W, Feitsma HJJ, Ensing GJ et al. Detection of a local Staphylococcal infection in mice with technetium-99m-labeled polyclonal human immunoglobulin. *J Nucl Med* 1991;32:468-474.
 20. Corstens FHM, van der Meer JWM. Chemotactic peptides: new locomotion for imaging of infection? *J Nucl Med* 1991;32:491-494.

EDITORIAL

Targeted Proteins for Diagnostic Imaging: Does Chemistry Make a Difference?

The imaging of occult infection is an important area of nuclear medicine. Vehicles for abscess localization have ranged from ⁶⁷Ga-citrate to radiolabeled leukocytes to radiolabeled immunoglobulin G (IgG) of current interest. Although ¹¹¹In labeled polyclonal IgG is probably the most widely cited protein being evaluated for focal infection scintigraphy (1-4), the mechanism of radiolabel accumulation remains unclear (5-8). In this issue of *The Journal of Nuclear Medicine*, Oyen et al. examined the roles of protein carrier and radiolabel in targeting of abscesses (9). They evaluated three different protein carriers and went on to assess the contribution of three different radiolabels and their associated chemistries in imaging experimental infectious foci in a rat model.

In the first part of the study, the authors compared radiolabeled IgG with immunoglobulin A (IgA) and human serum albumin (HSA) controls

since these proteins lack specificity for abscess. In each case ¹¹¹In labeling via a bifunctional chelate served as the standard radiolabel so that localization differences could be ascribed to individual protein distribution properties. Layered upon the targeting properties of the protein was the contribution of the radiolabels and their chemistries. In the second part of the study, the authors compared various radiolabels with IgG serving as the standard protein vehicle.

This study was well-conceived and designed to determine the role of protein carrier and radiolabel. However, an accurate interpretation of the role of the protein assumes the radiolabel serves as a radiotracer. Furthermore, an interpretation of the role of the radiolabel requires an analysis of its chemistry and an appreciation for the pharmacokinetics of its radioactive metabolites. Since these properties direct the biodistribution of radioactivity, it is instructive to briefly review relevant factors such as attachment stability, metabolic fate, and route of excretion characteristic of radioiodines, ¹¹¹In and ^{99m}Tc as used in this study.

dines, ¹¹¹In and ^{99m}Tc as used in this study.

IODINE AS RADIOTRACER

Radioiodine isotopes continue to be the most widely used protein radiolabels: ¹²³I for imaging, and ¹²⁵I and ¹³¹I for preclinical studies with their convenient longer half-lives and ready availability. The "easy" direct radioiodination approach in which the radioiodine is added to the activated ortho position of tyrosine is most often used, as was done in the Oyen et al. study (9). Label stability is usually sufficient to follow proteins in circulation or bound to cell surfaces. Once internalized by cells, however, catabolism releases peptide fragments or free amino acids with further metabolic processing ultimately releasing radioiodide (10). Deiodination may occur rapidly as in the example of the T-101 antibody in which imaging of cutaneous T-cell lymphoma is virtually precluded by rapid loss of radioactivity from tumor cells (11). Metabolically stabilized ligand chemistry has been developed which substan-

Received Dec. 10, 1991; accepted Dec. 12, 1991.
For reprints contact: Alan Fritzberg, NeoRx Corp., Seattle, WA 98119.

tially reduces the loss of radioiodide by the attachment of the radiolabel to non-activated para and meta positions on the aromatic ring, as the *p*-iodophenyl (12) or *m*-iodophenyl (13) derivative. While this avoids post-metabolic accumulation of radioiodide in the stomach and thyroid, retention of the iodobenzoate metabolites in target cells may be only slightly extended unless an approach such as the tyramine-cellobiose linker is used to enhance intracellular trapping of radiolabel following internalization and catabolism (14).

Oyen et al. (9) attribute the persistence of radioiodine relative to ^{111}In in the blood to deiodination and subsequent washin of radioiodide and radioiodinated fragments. This explanation seems unlikely since the iodine blood values were higher from two hours onward and the blood disappearance pharmacokinetics of small radioiodine species would be expected to be more rapid than that of IgG. Relative to ^{111}In , the ^{123}I radiolabel may in fact more accurately reflect the actual IgG blood disappearance rate. The abscess accumulation of ^{123}I -IgG was also correspondingly higher than that seen with ^{111}In -IgG at early times, but decreased more rapidly. Perhaps higher initial blood levels produced better early uptake of ^{123}I -IgG but deiodination resulted in diminished retention of the iodine radiolabel as described above.

INDIUM-111 AS RADIOTRACER

The ^{111}In labeling of IgG, IgA and HSA in the Oyen study (9) was accomplished using diethylenetriaminepentaacetic acid (DTPA) linked via the bicyclic anhydride derivative according to Hnatowich et al. (15). Analysis indicated 2–3 DTPA ligands per protein molecule.

Much effort has been expended and multiple strategies have been developed to improve ^{111}In protein labeling. The Hnatowich method utilizes one donor carboxylate group to link the chelating group to protein (15). Newer methods have been developed that allow chelation with all donor

groups and utilize a single linkage avoiding the potential for cross-linking leading to aggregation or altered protein tertiary structure. Gansow et al. (16) and Carney et al. (17) couple via carbon backbone derivatized with phenylisothiocyanate linkages, while Meares et al. (18) use a carbon backbone derivative of ethylenediamine-tetraacetic acid (EDTA). The molar substitution ratio of chelator to antibody has been shown to be an important parameter affecting both immunoreactivity and uptake in non-target organs (15,17,19) with caution suggested when ratios exceed 2 or 3:1.

Concerns surrounding ^{111}In as a protein radiotracer have focused on chelate stability and reticuloendothelial system clearance. Hepatic uptake and retention of ^{111}In radioactivity can result from either transchelation to transferrin which traffics to the liver, spleen and bone marrow or from protein degradation with retention of residual ^{111}In metabolites.

The ^{111}In biodistribution results presented by Oyen et al. (9) are generally consistent with previous studies characterizing the disposition of ^{111}In -labeled protein. In sum, significantly more uptake and retention in liver and spleen is seen with ^{111}In relative to $^{99\text{m}}\text{Tc}$ or radioiodine radiolabels. Somewhat surprisingly, renal uptake of ^{111}In labeled IgG was prominent. This observation and the more rapid blood disappearance of ^{111}In -IgG compared to ^{123}I -IgG suggest inherent serum instability or extraction of derivatized protein by liver and kidneys due to an excessive DTPA to IgG ratio.

TECHNETIUM-99m AS RADIOTRACER

The final protein radiolabel evaluated by Oyen et al. (9) was $^{99\text{m}}\text{Tc}$. The combination of superior imaging properties, widespread availability, and low cost indicate preference for $^{99\text{m}}\text{Tc}$ when possible. Since proteins are typically slow to target and slow to disappear from blood, their distribution properties may not be comple-

mentary with the 24-hr $^{99\text{m}}\text{Tc}$ window of imageability given its 6-hr half-life.

In this study, $^{99\text{m}}\text{Tc}$ labeling of IgG was accomplished by exchange to thiol derivatized protein prepared by reaction with 2-iminothiolane (20). This modification converts lysine side chain amine groups to amidines linked to a 3-carbon chain terminating in sulfhydryl suitable for binding technetium. While it is generally appreciated that sulfur is a major contributor to chelated technetium stability, the contribution of a single sulfhydryl donor is questionable without invoking assistance from collateral donor groups forming appropriately sized chelate rings. The metal donor contribution from the amidine group is not known. Even if strong donor capability was assumed, the bidentate unit forms a 7-membered chelate ring and would not be expected to be highly stable based on the findings of Davison et al. who investigated stability in a 5- and 6-membered chelate ring series of tetradentate N, S chelating agents (21). In that study, the tetradentate N_2S_2 system was significantly less stable with three 6-membered chelate rings.

Given the stability considerations raised above, it can be appreciated that $^{99\text{m}}\text{Tc}$ -IgG radiolabel disappears from the blood more rapidly than ^{123}I or ^{111}In -IgG radiolabels. In contrast to ^{111}In , a significant drop in $^{99\text{m}}\text{Tc}$ radioactivity retained at the abscess was seen. These results may be rationalized by assuming rapid initial accumulation at the site of inflammation (maximal uptake was reached by 6 hr postinjection with all of the variously labeled IgG) and subsequent loss of $^{99\text{m}}\text{Tc}$ due to inherent instability. Interestingly, useful imaging with $^{99\text{m}}\text{Tc}$ direct labeled proteins has been demonstrated. In some cases, only relative stability may be required for targeting with label loss actually contributing to blood disappearance and attainment of favorable target to background ratios for imaging. The 2-iminothiolane or similar direct labeling approach depends on achieving a balance of stability and instability, which seems ul-

timately more limited than a stable chelate and linkage approach with defined modifications to targeting proteins. Along this line, promising results have been reported with hydrazino nicotinamide linked ^{99m}Tc -IgG by Abrams et al. (22).

ROLE OF PROTEIN IN TARGETING INFECTIOUS FOCI

In the Oyen et al. study (9), IgA was used as a control immunoglobulin since it lacks Fc-gamma receptor affinity, in order to evaluate the role that receptor plays in IgG uptake and retention in infectious foci. Liver and spleen uptake of ^{111}In -IgA was higher than for ^{111}In -IgG indicating that the protein did alter the biodistribution properties. However, the abscess-to-background ratios were comparable for both IgG and IgA, which does not support a major role for specific receptor interactions. In addition, control ^{111}In -HSA showed comparable abscess uptake and retention. Thus, the localization of protein agents in abscesses may be governed largely by relative blood flow and the maintenance of local concentration dependent on circulation half-life. The target-to-background ratio attained in abscess may be governed more by factors relating to nonspecific recognition of labeled-altered proteins, internalization, catabolism, and either retention (^{111}In) or release (radioiodine, ^{99m}Tc), depending on radionuclide chemistry.

SUMMARY

The Oyen et al. study (9) is valuable in that it systematically evaluates several of the factors involved in radiolabeled protein uptake and retention in infectious foci. The role of particular proteins and their receptor specific interactions seems to be inconsequential in agreement with the findings of other (23). However, the role of the radiolabel was shown to be important and significant differences were delineated from comparisons of the radionuclides and their associated chemistries.

The conclusion implicating radionuclide chemistry and associated linkages underscores the need to optimize the attachment and labeling chemical modifications of protein carriers. Evaluation criteria should include serum stability, determination and assessment of the effect of molar substitution ratio, and potential for improving blood clearance without reducing the target-to-non-target ratio. Important areas for future study include characterization of radioactive metabolites and the design and synthesis of new ligands which direct the disposition of metabolites reducing retention in normal organs or accelerating renal excretion. Additionally, intracellular processing of radiolabel, compartmental distribution and strategies for augmenting internalization and retention within the target cell merit detailed exploration.

For each radionuclide of interest, ^{111}In , radioiodines, ^{99m}Tc and others, improved chemical moieties exist for controlling radiolabel fate. When carrying out mechanistic and evaluative studies, clear-cut conclusions will only be reached when defined and controlled chemistry is used. Having established a "gold standard," simplifications in radiolabeling and other chemical refinements can then be pursued with a quantitative understanding of the trade-offs in targeting agent performance versus other considerations such as cost reduction, simplicity, and convenience.

Alan R. Fritzberg
Paul L. Beaumier
NeoRx Corporation
Seattle, Washington

REFERENCES

1. Rubin RH, Fischman AJ, Callahan RJ, et al. In-111-labeled nonspecific immunoglobulin scanning in the detection of focal infection. *N Engl J Med* 1989;321:935-940.
2. LaMuraglia GM, Fischman AJ, Strauss HW, et al. Utility of indium-111-labeled human immunoglobulin G scan for the detection of focal vascular graft infection. *J Vasc Surg* 1989;10:20-28.
3. Oyen WJG, Claessens RAMJ, van Horn JR, van der Meer JWM, Corstens FHM. Scintigraphic detection of bone and joint infections with indium-111 labeled nonspecific polyclonal human immunoglobulin G. *J Nucl Med* 1990;31:403-412.
4. Wegener WA, Velchik MG, Weiss D, et al. Infectious imaging with indium-111-labeled nonspecific polyclonal human immunoglobulin. *J Nucl Med* 1991;32:2079-2085.
5. Fischman AJ, Rubin RH, Khaw BA et al. Detection of acute inflammation with indium-111 labeled non-specific polyclonal IgG. *Semin Nucl Med* 1988;18:335-344.
6. Rubin RH, Young LS, Hansen P, et al. Specific and nonspecific imaging of localized Fisher immunotype 1 *Pseudomonas aeruginosa* infection with radiolabeled monoclonal antibody. *J Nucl Med* 1988;29:651-656.
7. Morrel EM, Tompkins RG, Fischman AJ, et al. Autoradiographic method for quantitation of radiolabeled proteins in tissues using indium-111. *J Nucl Med* 1989;30:1538-1545.
8. Oyen WJG, Claessens RAMJ, Raemaekers JMM, de Pauw BE, van der Meer JWM, Corstens FHM. Diagnosing infection in febrile granulocytopenic patients with indium-111-labeled human IgG. *J Clin Oncol* 1991;in press.
9. Oyen WJG, Claessens RAMJ, van der Meer JWM, Corstens FHM. Biodistribution and kinetics of radiolabeled proteins in rats with focal infection. *J Nucl Med* 1992;33:398-402.
10. Hidalgo JU, Nadler SB. Stability studies on ^{131}I -labeled albumin. *J Nucl Med* 1962;3:268-272.
11. Carrasquillo JA, Mulshine JL, Bunn PA, et al. Indium-111 T-101 monoclonal antibody is superior to iodine-131 T-101 in imaging of cutaneous T-cell lymphoma. *J Nucl Med* 1987;28:281-287.
12. Wilbur DS, Hadley SW, Hyland MD, et al. Development of a stable radioiodinating reagent to label monoclonal antibodies for radiotherapy of cancer. *J Nucl Med* 1989;30:216-226.
13. Zalutsky MR, Narula AS. A method for the radiohalogenation of proteins resulting in decreased thyroid uptake of radioiodine. *Appl Radiat Isot* 1987;38:1051-1055.
14. Ali SA, Eary JF, Warren SD, Badger CC, Krohn KA. Synthesis and radiodination of tyramine cellobiose for labeling monoclonal antibodies. *Nucl Med Biol* 1988;15:557-561.
15. Hnatowich DJ, McGann J. DTPA-Coupled proteins procedures and precautions. *Nucl Med Biol* 1987;14:563-568.
16. Gansow OA. Newer approaches to the radiolabeling of monoclonal antibodies by use of metal chelates. *Nucl Med Biol* 1991;18:369-381.
17. Carney PL, Rogers PE, Johnson DK. Dual isotope study of iodine-125 and indium-111 labeled antibody in athymic mice. *J Nucl Med* 1989;30:374-384.
18. Cole WC, DeNardo SJ, Meares CF, et al. Comparative serum stability of radiochelates for antibody radiopharmaceuticals. *J Nucl Med* 1987;28:83-90.
19. Kuhlmann L, Steinstrasser A. Effect of DTPA to antibody ratio on chemical, immunological and biological properties of the ^{111}In -labeled F(ab')₂ fragment of the monoclonal antibody 431/31. *Nucl Med Biol* 1988;15:617-627.
20. Goedemans WTh, Panek KJ, Ensing GJ, de Jong MthM. A new, simple method for labeling of proteins with ^{99m}Tc by derivatization with 1-imino-4-mercaptoputyl groups. In: Nicolini M, Bandoli G, Mazzi U, eds. *Technetium and rhenium in chemistry and nuclear medicine*. New York: Raven Press; 1990:595-603.

21. Davison A, Jones AG, Orvig C, et al. A new class of oxotechnetium (5+) chelate complexes containing a TcON_2S_2 core. *Inorg Chem* 1981;20:1629-1632.
22. Abrams MJ, Juweid M, tenKate CI, et al. Tech-

- netium-99m-human polyclonal IgG radiolabeled via the hydrazino nicotinamide derivative for imaging focal sites of infection in rats. *J Nucl Med* 1990;31:2022-2028.
23. Rubin RH, Fischman AJ, Needleman M, et al.

Radiolabeled nonspecific, polyclonal human immunoglobulin in the detection of focal inflammation by scintigraphy: comparison with gallium-67 citrate and technetium-99m-labeled albumin. *J Nucl Med* 1989;30:385-389.

(continued from p. 344)

SELF-STUDY TEST

Radiobiology and Radiation Protection

ANSWERS

exposure and conception (it has been observed that avoiding conception for a time interval after irradiation greatly reduces the production of mutations).

ITEMS 25-29: Multistage Development of Radiation-Induced Cancer

ANSWERS: 25, T; 26, F; 27, T; 28, T; 29, T

For many types of radiation-induced cancer, the epidemiologic evidence suggests that events subsequent to irradiation are required to produce a cell that is capable of uncontrolled proliferation. For example, in irradiated populations no excess risk of breast or lung cancer has been seen until the exposed individuals have reached ages at which these cancers usually are observed in nonirradiated populations. This suggests that induction of these cancers requires one or more time-dependent factors in addition to whatever role ionizing radiation plays in their causation.

Bone cancer and leukemia, on the other hand, have appeared in excess within a few years after exposure, suggesting that the multiple stages must occur rapidly or that they may not be required to complete the carcinogenic process. These observations do not support the concept of multistage tumor induction by radiation.

Another marked contrast that distinguishes radiation-induced leukemia and bone cancer is the return of risk to near normal levels within a period of 30 yr or less after irradiation, whereas, with other cancers the risk period may extend to the end of life. These long latent periods again imply a multistage process. The literature of experimental carcinogenesis abounds with examples in which cocarcinogens or promoting agents modify the dose-response curve and the latent period for radiation carcinogenesis. This reduction in latent period by "promoters" indicates that the process (promotion) is at least a second step after initiation.

Recent studies of malignant transformation by viral oncogenes and activated cellular oncogenes suggest that cellular malignant transformation may require activation by more than one cellular oncogene. It is possible, thus, that the long latent periods that characteristically elapse between irradiation and clinical appearance of the cancer may result from the need for activation of recessive oncogenes or other sequential steps.

References

1. Bishop JM. The molecular genetics of cancer. *Science* 1987;235:305-311.
2. Land H, Parada LF, Weinberg RA. Cellular oncogenes and multistep carcinogenesis. *Science* 1983;222:771-778.

ITEMS 30-33: Radiation-Induced Cancer in Humans

ANSWERS: 30, F; 31, T; 32, T; 33, F

The presence of radiation-induced cancers in a human population is difficult to detect and to quantitate because the cancers induced by radiation are indistinguishable from those occurring naturally. Their existence can be detected only on the basis of a statistically significant excess in irradiated individuals above the natural incidence. Detection of radiation-induced cancers is also difficult because of the long latent periods (typically 10 yr or more for solid tumors) between irradiation and detection, as shown in the following table.

Approximate Latent Periods (yr) for Radiation-Induced Cancers in Humans

Type	Minimum	Mean	Total Period of Expression
Leukemia	2-4	10	25-30
Bone	2-4	15	25-30
Thyroid	5-10	20	> 40
Breast	5-15*	23	> 40
Other solid tumors	10	20-30	> 40

*Varies with age at exposure

Adapted from Ref. 2, below.

Radiation-induced cancers are considered to be the most important late somatic effect of radiation. Leukemia induced by radiation stands out because of the natural rarity of the disease, the relative ease of its induction by radiation, and its short latent period (2-4 yr). When the total risk of radiation-induced cancer is considered, however, it is clear that the risk of induced solid tumors exceeds that of leukemia. For the A-bomb survivors, the ratio of radiation-induced solid tumors to leukemias is now approximately 4:1. The major sites of solid cancers induced by whole-body radiations are breast (in women), thyroid, lung, and some digestive organs.

References

1. Kato H, Schull WJ. Studies on the mortality of A-bomb survivors. 7. Mortality, 1950-1978: Part 1. Cancer mortality. *Radiat Res* 1982;90:395-432.
2. Pizzarello DJ, Witcofski RL. *Medical Radiation Biology*, 2nd Ed. Philadelphia: Lea and Febiger, 1982:42-64.

ITEMS 34-38: Sex Dependence of Radiation Carcinogenesis

ANSWERS: 34, T; 35, F; 36, F; 37, T; 38, F

The incidence of radiation-induced human breast and thyroid cancer is such that the total cancer risk is greater for women than for men. Breast cancer occurs almost exclusively in women, and absolute-risk estimates for thyroid cancer induction by radiation are higher for women than for men (as is the case for the natural incidence). With respect to other cancers, the radiation risks in the two sexes are approximately equal.

Reference

1. National Academy of Sciences. *The Effects on Populations of Exposure to Low Levels of Ionizing Radiation*. Report of the Advisory Committee on the Biological Effects of Ionizing Radiation (BEIR). Washington, D.C.: Division of Medical Science, National Academy of Sciences, National Research Council, 1980: 167-176.