- Mazziotta JC, Phelps ME, Plummer D, et al. Quantification in positron emission computed tomography. 5. Physical-anatomical effects. J Comput Assist Tomogr 1981;5:734-743.
- Langen KJ, Roosen N, Herzog H, et al. Investigation of brain tumors with <sup>90m</sup>Tc-HMPAO SPECT. Nucl Med Commun 1989;10:325–334.
- Lee FYF, Allalunis-Turner MJ, Siemann DW. Depletion of tumor versus normal tissue glutathione by buthionine sulfoximine. Br J Cancer 1987;56:33-38.
- Weibel ER. Practical methods for biological morphometry. In: Stereological Methods, volume 1. London, New York: Academic Press; 1979.
- Hung JC, Corlija M, Volkert A, et al. Kinetic analysis of technetium-99md,1-HM-PAO decomposition in aqueous media. J Nucl Med 1988;29:1568–1576.
- Hoffman TJ, Corlija M, Chaplin SB, et al. Retention of (<sup>99m</sup>Tc)-d,1-HM-PAO in rat brain: an autoradiographic study. J Cereb Blood Flow Metab 1988:8:38-43.
- Lassen NA, Andersen AR, Friberg L, et al. The retention of (99mTc)-d,1-HMPAO in the human brain after intracarotid bolus injection: a kinetic analysis. J Cereb Blood Flow Metab 1988;8(suppl 1):S12-S22.
- 20. Rowell NP, McCready VR, Cronin B, et al. 99mTc-labelled meso-HMPAO and glutathione content of human lung tumors. Nucl Med Commun

- 1989:10:503-508.
- Costa DC, Ell PJ, Cullum ID, et al. The in vivo distribution of 99mTc-HM-PAO in normal man. Nucl Med Commun 1986;7:647-658.
- Suess E, Huck S. (99mTc)-d,1-HMPAO accumulation in cultured neurons. *J Cereb Blood Flow Metab* 1989;9(suppl 1):S735.
- Costa DC, Lui D, Sinha AK, et al. Intracellular localisation of <sup>99m</sup>Tc-d,1-HMPAO and <sup>201</sup>T1-DDC in rat brain. *Nucl Med Commun* 1989;10:459– 466.
- Nakano S, Kinoshita K, Jinnouchi S, et al. Dynamc SPECT with iodine-123-IMP in meningiomas. J Nucl Med 1988;29:1627–1632.
- Nishimura T, Hayashida K, Uehara T, et al. Two patients with meningioma visualized as high uptake by SPECT with N-isopropyl-p-iodo-amphetamine (I-123). Neuroradiology 1988;30:351-354.
- Ali-Osman F, Caughlan J, Gray DS. Decreased DNA interstrand crosslinking and cytotoxicity induced in human brain tumors by 1,3-bis-(2chloroethyl)-1-nitrosurea after in vitro reaction with glutathione. Cancer Res 1989:49:5954-5958.
- Evans CG, Bodell WJ, Ross D, et al. Role of glutathione and related enzymes in brain tumor cell resistance to BCNU and nitrogen mustard [Abstract]. Proc Ann Meet Am Soc Cancer Res 1986;27:267.

## **EDITORIAL**

# Technetium-99m-HMPAO Retention and the Role of Glutathione: The Debate Continues

Many patients presenting with primary brain tumors face a grim prognosis due to the inability to control local disease (1). Secondary brain tumors present similar therapeutic difficulties, although survival probably depends more on the extent of extra-cerebral disease (2). However, as more aggressive antineoplastic regimens begin to demonstrate greater control of systemic disease, survival may depend on adequate treatment of central nervous system secondaries.

Tumor perfusion may play an important role in the success of antineoplastic therapies such as radiotherapy and chemotherapy due to their dependence on adequate blood flow for oxygenation and drug transport, respectively. A noninvasive means of studying brain tumor blood flow should help us to improve our understanding of the role perfusion plays in brain tumor therapy and prognosis.

#### rCBF TRACERS

Regional cerebral blood flow (rCBF) tracers for use with single-pho-

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ton emission tomography (SPECT) must possess, amongst other characteristics, a mechanism by which the activity extracted into the brain is fixed for a sufficient time to allow for image acquisition. In order to accomplish this, a "trapping" mechanism must exist. Ideally, a prerequisite for the trapping mechanism is that it is unaffected by pathology. It is then possible to begin to interpret tracer distribution images as true rCBF images in a range of disorders.

The ability to image regional rCBF using nondynamic SPECT was brought about by the introduction of the <sup>123</sup>I-labeled-amines; IMP (p-iodo-N-isopropyl amphetamine) (3) and HIPDM (N,N,N,-trimethyl-N-[2-hydroxy-3-methyl-5-iodobenzyl]-1,3propanediamine) (4). Both compounds demonstrated high first-pass extraction in the brain and prolonged retention. While the exact "trapping" mechanism of the amines remains uncertain (5), they have proved useful in aiding the diagnosis of both cerebrovascular and neurologic disorders (6-8).

The application of these early SPECT tracers excluded the study of neurooncology due to the reported lack of a functional trapping mechanism in primary brain tumors (9-11), although there were reports of uptake in metastases to brain (12,13) and in isolated cases of primary brain tumors (14).

### Technetium-99m-HMPAO

Technetium-99m-d,1-HMPAO (exametazime) is the first clinically available <sup>99m</sup>Tc-labeled rCBF tracer. This tracer exhibits high first-pass extraction in the brain and prolonged retention, making it suitable for SPECT imaging using non-dynamic systems (15,16). Initial studies in patients with brain tumors indicated that, unlike IMP and HIPDM, this new tracer was capable of localizing in primary brain tumors (17).

The cerebral retention of <sup>99m</sup>Tc-HMPAO is believed to involve the intracellular conversion of the hydrophobic Tc-HMPAO to a species which is incapable of rapid back diffusion. The proposed mechanism of this conversion is thought to involve interaction of <sup>99m</sup>Tc-HMPAO with glutathione (GSH) (18). This theory is based on the similarity in measured conversion rates of <sup>99m</sup>Tc-d,1-HMPAO (to a less hydrophobic species) when ex-

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posed to rat brain homogenates or aqueous solutions of GSH. The meso diastereomer of 99mTc-HMPAO demonstrated a slower conversion rate but similar rates for brain homogenate and aqueous GSH (19). Pharmacokinetic modeling of the human distribution and retention of the meso and d,1 forms of 99mTc-HMPAO, using these experimentally derived conversion rates ("K3") for each, were shown to be in good agreement with measured human distributions (19). While these data support the hypothesis that GSH is responsible for intracellular trapping, such findings are not unequivocal proof.

#### **GSH**

GSH is a ubiquitous tripeptide, accounting for a third of the brain's total non-protein, acid-extractable nitrogen and at least 95% of the thiols in this fraction (20). GSH is involved in many cellular functions, it is a cofactor in the metabolism of xenobiotics, serves as a storage and intercellular transport form of cysteine, provides for the maintenance of protein sulfhydryl reduction and is involved in the protection of membrane lipids from peroxidation (21). In addition, GSH is reported to play a role in DNA synthesis, protein synthesis and amino acid transport as well as being a co-factor in several enzyme systems (22).

Since GSH is so intimately linked to a variety of metabolic functions and the detoxification of metabolic by-products, it seems likely that normal metabolically active tissue demanding high nutrient perfusion may also have high levels of GSH or GSH-related enzyme activity. Conversely, ischemic tissues which lack nutrient perfusion would be expected to exhibit low levels of GSH.

Data in the rat kidney indicates that induction of ischemia leads to rapid loss of GSH, secondary to a loss of ATP, thereby inhibiting GSH synthesis (23). In this case, a lack of synthesis coupled with rapid catabolism of renal GSH leads to GSH depletion. Similar studies indicate that reductions of

GSH occur in rat brain as a result of ischemia (24). However, brain GSH turnover is slow compared to the kidney and other organs (25) and it is possible that transient drops in perfusion would not produce rapid changes in brain GSH levels.

It therefore seems likely that GSH levels may parallel blood flow in certain tissues and that GSH levels may be reduced in hypoxic tissues. Recent evidence for the existence of a carrier-mediated transport system for the blood-brain barrier transport of GSH (26) suggests that a component of intracellular GSH content may be related to interorgan transport (25), which could be flow-dependent. Hence, it may prove difficult to separate outflow from GSH content in normal brain tissue.

There are reports in the literature, however, which suggest that intracellular trapping may not wholely depend on the presence of intracellular GSH. El-Shirbiny et al. (27) reported a lack of correlation between organ GSH content and 99mTc-d,1-HMPAO uptake in the rat before and after glutathione depletion using diethyl maleate. Rowell et al. (28) studied the uptake of the 99mTc-meso-HMPAO in lung tumors, comparing uptake to lung tumor GSH levels. The driving hypothesis of the latter study was that the accumulation of 99mTc-meso-HMPAO was dependent more on GSH content than blood flow (19). No correlation could be demonstrated between 99mTc-meso-HMPAO uptake and tumor GSH concentration or tumor/normal lung GSH ratios in the small number of patients studied, although the intracellular GSH concentration in malignant lung tumors was generally higher than that of normal lung.

# RELATIONSHIP BETWEEN GSH LEVELS AND d,1-HMPAO

In this issue of *The Journal of Nuclear Medicine*, Suess et al. describe the relationship between tumor GSH levels and <sup>99m</sup>Tc-d,1-HMPAO uptake (as measured by tumor/cerebellum ratios) in patients with primary and

secondary brain tumors. They report a statistically significant correlation between tumor GSH levels and <sup>99m</sup>Tc-d,1-HMPAO uptake in both meningiomas and gliomas, giving further indirect support to the GSH "trapping" mechanism hypothesis. Conversely, no correlation could be demonstrated between GSH content in metastatic tumor deposits in the brain and <sup>99m</sup>Tc-d,1-HMPAO uptake. In addition, <sup>99m</sup>Tc-d,1-HMPAO uptake occurred in certain tumors even when tissue GSH levels were undetectable.

Two obvious findings are apparent in this report: 99mTc-d,1-HMPAO uptake into brain tumors often parallels GSH levels; and secondary tumors to the brain lack any such parallelism. Interpretation of these findings is not clear cut and may point to anatomic and physiologic differences between these two tumor groups based on their tissue of origin, the nature of their vascularization or blood supply and their biochemistry relative to GSH metabolism, including GSH synthesis and content and the activity of enzymes that utilize GSH as a substrate or co-factor.

The correlation of GSH with 99mTc-HMPAO uptake in primary brain tumors may reflect further subtleties of GSH metabolism that have yet to be considered. Recent data indicate that brain tumors possess elevated levels of an isoenzyme, of the glutathione-Stransferase (GST) family of enzymes, which catalyze the conjugation of reduced GSH to hydrophobic, electrophilic compounds (29,30). If GSH is indeed involved in the trapping of <sup>99m</sup>Tc-HMPAO, then it is not unlikely that the enzymes responsible for the conjugation of GSH to xenobiotics may play a role in this process. This may help explain the correlation found with primary brain tumors, but it does little to clarify the lack of relationship seen in metastases. Variable washout rates of the "converted" 99mTc-HMPAO species between secondary and primary brain tumors may also add to the discrepancy between GSH levels and observed uptake.

Further studies are required to gain a better understanding of the "trapping" mechanism of <sup>99m</sup>Tc-HMPAO in normal and neoplastic tissues. Determining the relationship between perfusion (using a freely diffusible tracer) and GSH tissue levels in normal and diseased tissues may be the first step in achieving this goal.

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#### REFERENCES

- Nelson DF, Urtasun RL, Saunders WM, et al. Recent and current investigations of radiation therapy of malignant gliomas. Semin Oncol 1986;13:46-55.
- Greig NH. Chemotherapy of brain metastases: current status. Cancer Treat Rev 1984;11:157– 186.
- Winchell HS, Baldwin RM, Lin TH. Development of I-123-labeled amines for brain studies. J Nucl Med 1980;21:940-946.
- Kung HF, Tramposch KM, Blau M. A new brain perfusion imaging agent: [I-131] HIPDM: N,N,N'-trimethyl-N-'[2-hydroxy-3-methyl-5iodobenzyl]-1,3-propanediamine. J Nucl Med 1983;24:66-72.
- Kung HF, Ohmomo Y, Kung M-P. Current and future radiopharmaceuticals for brain imaging with single-photon emission computed tomography. Semin Nucl Med 1990;22:290– 302.
- Holman BL, Hill TC, Polak JF. Cerbral perfusion imaging with iodine-123-labeled amines. Arch Neurol 1984;1060-1063.
- 7. Sharp P, Gemmel H, Cherryman G, et al. Application of iodine-123-labeled isopropyl-

- amphetamine imaging to the study of dementia. J Nucl Med 1986;27:761-768.
- Moretti JL, Cinotti L, Cesaro P, et al. Amines for brain tomoscintigraphy. Nucl Med Commun 1987:8:581-595.
- LaFrance ND, Wagner HN, Whitehouse P, et al. Decreased accumulation of isopropyl-iodoamphetamine (I-123) in brain tumors. J Nucl Med 1981;22:1081-1083.
- Hill TC, Holman BL, Lovett R, et al. Initial experience with SPECT (single-photon emission tomography) of the brain using N-isopropyl I-123-p-iodoamphetamine. J Nucl Med 1982;23:191-195.
- Creutzig H, Schober O, Gielow P, et al. Cerbral dynamics of N-isopropyl-(I-123)p-iodoamphetamine. J Nucl Med 1986;27:178-183.
- Ell PJ, Cullum I, Donaghy M, et al. Cerebral blood flow studies with iodine-123-labelled amines. *Lancet* 1983;1:1348-1352.
- Szasz IJ, Lyster D, Morrison RT. Iodine-123-IMP uptake in brain metastases from lung cancer. J Nucl Med 1983;26:1342–1343.
- McVeigh MC, Merchut MP, Karesh SM, et al. Intracranial tumors: unexpected uptake of I-123-HIPDM. Radiology 1986;161:409-411.
- Ell PJ, Hocknell JML, Jarrit PH, et al. A Tc-99m-labelled radiotracer for the investigation of cerebral vascular disease. Nucl Med Commun 1985;6:437-441.
- Nowotnik DP, Canning LR, Cumming SA, et al. Development of a Tc-99m-labelled radiopharmaceutical for CBF imaging. Nucl Med Commun 1985:6:499-506.
- Babich JW, Keeling F, Flower MA, et al. Initial experience with Tc-99m-HMPAO in the study of brain tumors. Eur J Nucl Med 1988;14:39– 44
- Neirinckx RD, Harrison RC, Forster AM, et al. A model for the in vivo behavior of Tc-99m d,1-HMPAO in man [Abstract]. J Nucl Med 1987;28:559.
- Neirinckx RD, Burke JF, Harrison RC, et al. The retention mechanism of Tc-99m-HMPAO: intracellular reaction with glutathi-

- one. J Nucl Cereb Blood Flow Metab 1988;8(suppl 1):s4-s12.
- McIlwain H, Bachelard HS. Biochemistry and the central nervous system, fifth edition. New York: Churchill Livingstone; 1985:163-1170.
- Meister A. New aspects of glutathione biochemistry and transport: selective alteration of glutathione metabolism. FASEB 1984;43: 3031-3042.
- 22. Meister A, Anderson ME. Glutathione. Ann Rev Biochem 1983;52:711-760.
- Slusser SO, Grotyohann LW, Martin LF, Scaduto RC. Glutathione catabolism by the ischemic rat kidney. Am J Physiol 1990;258:F1547-F1553.
- Noguchi K, Higuchi S, Matsui H. Effects of glutathione isopropyl ester on GSH concentrations in the ischemic rat brain. Res Commun Pathol Pharmacol 1989;64:165-168.
- Griffith, Meister A. Glutathione: interorgan translocation, turnover and metabolism. Proc Natl Acad Sci 1979:11:5606-5610.
- Kannan R, Kuhlenkamp JF, Jeandidier E, et al. Evidence for carrier-mediated transport of glutathione across the blood-brain barrier in the rat. J Clin Invest 1990;85:2009-2013.
- el-Shirbiny AM, Sadek S, Owunwanne A, et al. Is Tc-99m-hexamethylene propylene amine oxime uptake in the tissues related to glutathione cellular content? Nucl Med Commun 1989;10:905-911.
- Rowell NP, McCready VR, Cronin B, et al. Tc-99m-labelled meso-HMPAO and glutathione content of human lung tumours. Nucl Med Commun 1989;10:503-508.
- Kantor RRS, Giardina SL, Bartolazzi A, et al. Monoclonal antibodies to glutathione S-transferase—immunohistochemical analysis of human tissues and cancers. *Int J Cancer* 1991:47:193-201.
- Hara A, Yamada H, Saki N, et al. Immunohistochemical demonstration of the placental form of gltuthione S-transferase, a detoxifying enzyme in human gliomas. Cancer 1990;66:2563-2568.