

---

# A Comparative Study of Renal Scintigraphy and Clearance with Technetium-99m-MAG<sub>3</sub> and Iodine-123-Hippurate in Patients with Renal Disorders

Roland Müller-Suur, Ingeborg Bois-Svensson, and Laszlo Mesko

Karolinska Institute, Departments of Clinical Physiology and Hospital Physics, Danderyds Hospital, Stockholm, Sweden

---

The aim of this study was to compare kit prepared technetium-99m-mercaptoacetyltriglycine (<sup>99m</sup>Tc-MAG<sub>3</sub>) with our routine radiopharmaceutical, iodine-123-hippurate our routine radiopharmaceutical, iodine-123-hippurate ([<sup>123</sup>I]OIH) for renal dynamic scintigraphy. Seventeen patients with different nephrologic disorders or hypertension were first studied with OIH and then reinvestigated with MAG<sub>3</sub> 2–8 days later. Renal MAG<sub>3</sub> gamma camera images were almost identical with those of OIH except for higher ( $p < 0.01$ ) liver-to-background ratios at 20 min postinjection, irrespective of kidney function. Urinary peristalsis was visible longer and more clearly in the MAG<sub>3</sub> studies. MAG<sub>3</sub> and OIH renograms showed identical relative kidney uptake ( $r = 0.99$ ), but elimination of MAG<sub>3</sub> from the kidneys was slower ( $p < 0.01$ ). The plasma clearance of MAG<sub>3</sub> was lower than that of OIH, but correlated ( $r = 0.92$ ) significantly. The plasma distribution volume and content in blood cells was lower ( $p < 0.01$ ), but the binding of MAG<sub>3</sub> to plasma proteins was higher, 90%, as compared with 74% for OIH,  $p < 0.01$ . Urinary excretion expressed as a percent of the given dose 60 min after injection was the same for the two substances. Thus, there are some significant differences in the renal handling, plasma distribution, and cell penetration between MAG<sub>3</sub> and [<sup>123</sup>I]OIH. MAG<sub>3</sub>, however, seems to have particular qualifications as a radionuclide for dynamic renal scintigraphy, especially in patients who require acute investigations or in those with low renal function.

J Nucl Med 1990; 31:1811–1817

---

Recently a new radiopharmaceutical for renal dynamic scintigraphy, technetium-99m-mercaptoacetyltriglycine (<sup>99m</sup>Tc-MAG<sub>3</sub>) has been introduced (1,2), which combines the advantage of high renal extraction

with a proper energy emission for gamma camera imaging. Animal experiments (1,3–6) outlined the excretion characteristics of MAG<sub>3</sub> in comparison with iodine-131-hippurate (1,4–6), chromium-51-EDTA, iodine-125-hippurate, and iodine-123-hippurate ([<sup>123</sup>I]OIH) (3), concluding that MAG<sub>3</sub> may be a suitable replacement for hippurate. Clinical trials of MAG<sub>3</sub> at different laboratories in comparison with <sup>131</sup>I-hippurate (2,7–11), <sup>123</sup>I-hippurate (12), and <sup>99m</sup>Tc-DTPA (13) have been reported. All authors concluded that <sup>99m</sup>Tc-MAG<sub>3</sub> is an efficacious radiopharmaceutical for renal radionuclide studies and a suitable replacement for hippurate as no side effects have hitherto been reported. The present clinical trial was performed to compare MAG<sub>3</sub> with OIH as a renal imaging agent with excellent imaging qualifications, a clearance corresponding to the effective renal plasma flow (ERPF), and a low irradiation dose to the patient. We also wanted to study whether the observation in rats of a significant extrarenal excretion to the bile (3) is present in patients. Furthermore, we studied the clearance of MAG<sub>3</sub> in comparison to hippurate because of the debate in literature about the excretion characteristics of MAG<sub>3</sub>.

## METHODS

### Patients

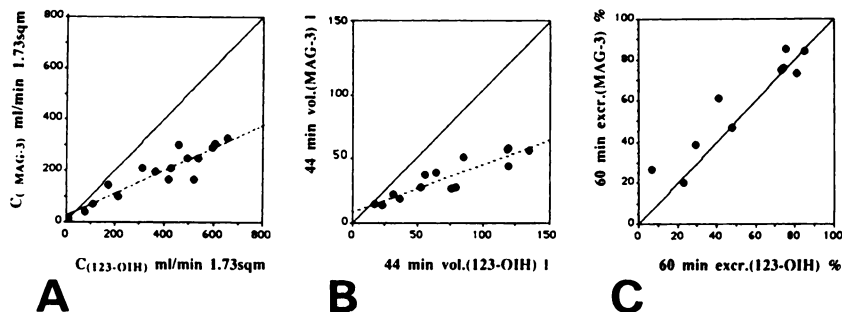
Patients in this study were referred for evaluation of different nephrologic disorders (two for renal cysts, two for pyelonephritis, two for renal dysplasia, one for examination after ureteral surgery, one for renal stone, one for acute renal failure, one for chronic renal insufficiency, and nine for hypertension). All were first investigated with routine <sup>123</sup>I-hippuran gamma camera renography. After information of the aim and the procedure of this study was presented, the patients gave their consent to participate in an additional renography with MAG<sub>3</sub> two to eight days later. The study was approved by the ethical committee of Karolinska Institute and the regional isotope-committee.

---

Received Apr. 24, 1989; revision accepted Sept. 26, 1989.  
For reprints contact: Roland Müller-Suur, Department of Clinical Physiology, Danderyds Hospital, S-18288 Danderyd, Stockholm, Sweden.

**FIGURE 1**

Correlation of clearance (A), volume of distribution (B), and of the percent excretion 60 min postinjection (C) between [<sup>123</sup>I]OIH (abscissa) and MAG<sub>3</sub> (ordinate). Solid lines are the lines of identity, broken lines the regression lines. See text for statistical calculations.



**Radionuclides**

For the first study, 1 mCi (37 MBq) of [<sup>123</sup>I]OIH (RadioEIR-isotopenservice, Würenlingen, Switzerland) was used, and for the second study 2 mCi (70 MBq) <sup>99m</sup>Tc-mercaptoacetyltryglycine (MAG<sub>3</sub>, Mallinckrodt, Petten, Netherlands), which was prepared 60 min before injection by a labeling kit (14). To the supplied MAG<sub>3</sub> powder in sealed glass vials, 4 ml 0.9% NaCl solution was added containing ~150 MBq <sup>99m</sup>TcO<sub>4</sub>, which was freshly eluted from a technetium generator (Tecegen S, Behringwerke, Frankfurt, FRG), eluate ≤ 0.2 ml. The vial was heated in a lead-shielded water bath to ~100°C for 10 min and then cooled under running water. The radiochemical purity (> 95%) was performed according to the manufacturer's recommendation (13). Two millicuries of MAG<sub>3</sub> (70 MBq), ~2 ml, were injected intravenously via a three-way connector to a butterfly line and flushed with saline. This MAG<sub>3</sub> preparation was bound at 90% to plasma proteins as measured by a centrifugation filtration technique (Amicon,

Centrifree, MA). The corresponding value for our [<sup>123</sup>I]OIH was 74%.

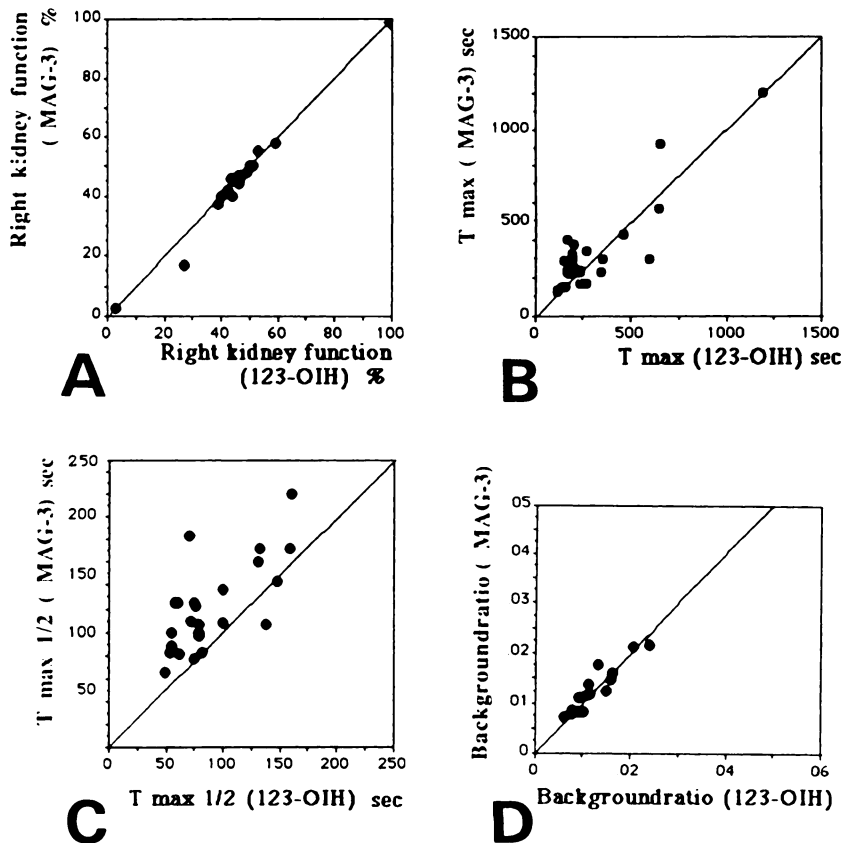
**Clearance Studies**

For [<sup>123</sup>I]OIH, plasma clearance was calculated according to Tauxe and Dubovsky (15), i.e., a 44-min one-sample "ERPF" method. For <sup>99m</sup>Tc-MAG<sub>3</sub> clearance, six venous plasma samples at 5, 10, 20, 44, 60, and 120 min postinjection were taken from the forearm opposite to the injection arm. MAG<sub>3</sub> clearance was calculated from the injected amount divided by the area under the curve of plasma activity versus time. Bi-exponential curve-fit analysis was used for the calculation of that area. In one patient with acute renal failure, we measured the renal clearance (i.e., U · V/P) by sampling urine over 15 hr. Plasma samples in that case were taken before and after the urine collection and averaged.

For calculation of urine excretion, patients were asked to void 60 min postinjection for urine analysis. The excretion

**FIGURE 2**

Correlation of right kidney function (A), time-to-peak activity ("Tmax") (B), time from peak to 50% activity ("Tmax 1/2") (C), and ratio between background and renal activity 2–4 min postinjection (D) between [<sup>123</sup>I]OIH (abscissa) and <sup>99m</sup>Tc-MAG<sub>3</sub> (ordinate). Lines of identity are shown. See text for statistics.



was then expressed as percent of the injected amount. For estimation of blood cell content in percent of whole-blood activity, blood was counted using the 44-min samples and corrected for plasma activity and hematocrit.

All counting was done in a well-type counter (Nuclear Data Instruments, Chicago, IL) and corrected for background and weight. Standards of OIH and  $MAG_3$  solutions were prepared at the time of dose preparation and the exact given dose was estimated from standard activity and weight difference of the syringe before and after injection.

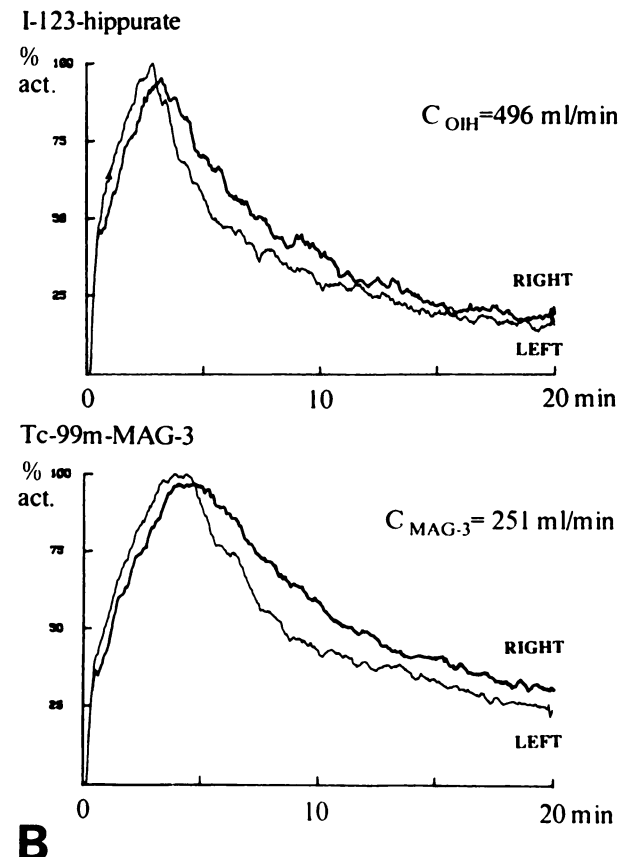
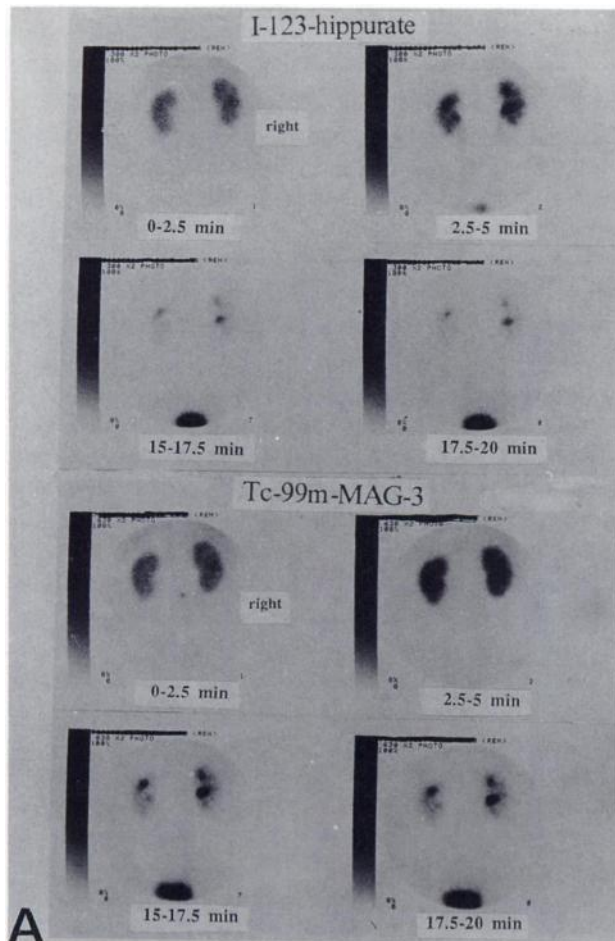
### Gamma Camera Studies

Patients were studied either supine or seated comfortably in a chair with the gamma camera head placed dorsally covering the region of the kidneys and ureters and part of the bladder. Identical body positions were used for both studies. The patients were normally hydrated with 0.1–0.2 liters of fluid orally just before the study. In case of poor tracer elimination from the kidneys, furosemide, 0.5 mg/kg body weight, was given intravenously 10–15 min after tracer injection. The data were collected in 2.5 sec frame mode by a computer system (Gamma 11) linked to a gamma camera (General Electric, 400 ACT, Milwaukee, IL) to which a high-sensitive, low-energy parallel-hole collimator was fitted. One patient with acute renal failure was, however, studied at bed-

side in the intensive care unit using a mobile camera (General Electric, 300A) with an internal computer system (Starcam) and the camera head positioned ventrally covering the kidney and ureters. Dynamic pictures were collected for 20 min and additional dorsal pictures, if possible and convenient for the patient, were collected at 60–62 min and 120–122 min postinjection. The data were displayed in a  $64 \times 64$  matrix on a high resolution video monitor.

Regions of interest (ROIs) were set over both kidneys using the 1–3-min postinjection data and over the semilunar background surrounding the individual kidneys. Additional ROIs ( $4 \times 4$  pixel) were set over the liver, spleen, and abdominal aorta. From the kidney regions, renogram curves were corrected for kidney depth, and individual semilunar backgrounds of relative renal function (i.e., uptake 60–120 sec postinjection) were calculated. Kidney depth was measured using ultrasound after the first study. The time-to-peak activity ( $T_{max}$ ) was measured and tracer elimination from the kidneys was calculated as the ratio of activity 10 min after peak and peak activity. Also, the time from  $T_{max}$  to 50% of the activity at  $T_{max}$  was calculated ( $T_{max} \frac{1}{2}$ ) as an index of elimination.

Liver and spleen uptake was determined as the ratio of activity per pixel of liver or spleen region (see above) at 2–4 min, 18–20 min, 60–62 min, or 120–122 min postinjection to the activity per pixel of the background region (aorta).



**FIGURE 3**

(A) Images and (B) renograms of a patient referred for investigation of hypertension. Both studies with  $[^{123}I]OIH$  (upper panels) and with  $MAG_3$  (lower panels) show normal pattern, substantially identical.  $C_{OIH}$  and  $C_{MAG_3}$  give plasma clearance values.

Background activity was measured as the ratio of 2–4 min activity of aorta background divided by the sum of the right and left renal activity at the same time.

Ureteral peristalsis was studied by two different approaches. The first consisted in displaying the images on the screen in a dynamic “cine-mode” to assess the peristaltic movements visually. The second method consisted in displaying them in a condensed form as a time-space matrix described by Müller-Schauenburg (16) using a special software for the Gamma 11 computer. The steps for matrix generation are:

1. ROI series covering the ureter, the first being the renal pelvis.
2. Corresponding ROI curve series.
3. Background subtraction for each segment in each curve (subtracting either the average or the minimum).
4. The inscription of the subtracted data into a  $64 \times 64$  matrix [y–line = ROI number (i.e., “space”, x–row = frame number; i.e., “time”).]
5. Displaying the matrix with the upper threshold set to ~20% of the maximum. Ortho-grade peristaltic contractions of the ureter appear as oblique lines in this time-space matrix during tracer elimination from the kidney, i.e., after T<sub>max</sub> on the renogram, normally 1–3/min.

### Statistical Analysis

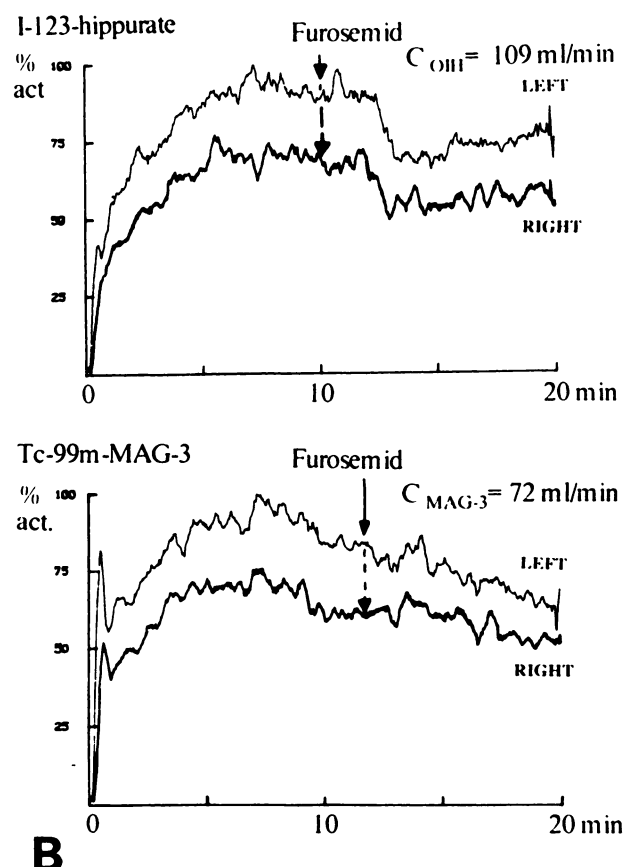
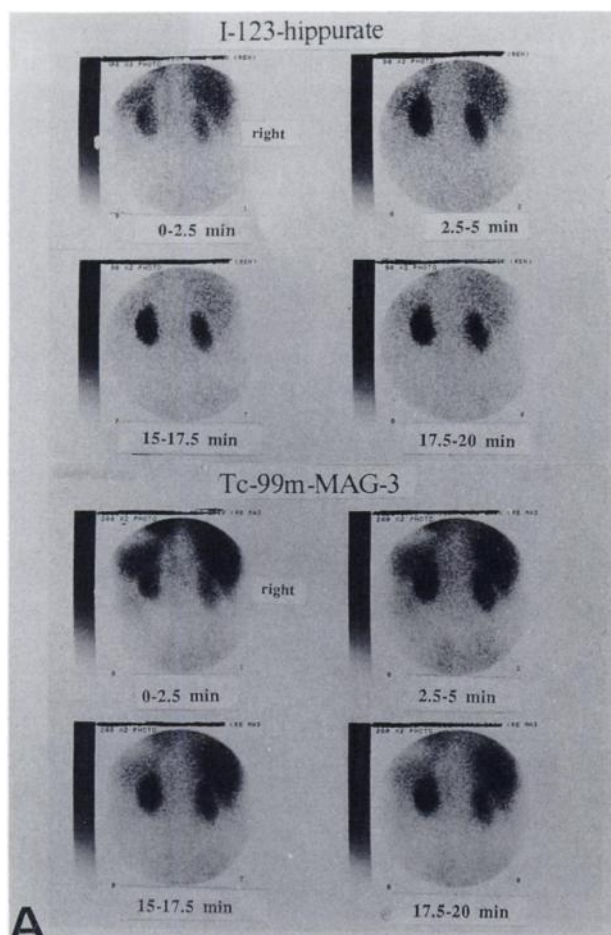
Analysis was performed using the Student’s paired t-test or conventional regression analysis to give the correlation coefficient,  $r$ . Two-tailed analysis,  $p < 0.01$ , was viewed as significant. Mean values and s.e.m. are given.

## RESULTS

### Clearance Studies

None of the patients complained of any side effects from MAG<sub>3</sub>. Figure 1 shows the clearance studies data. MAG<sub>3</sub> clearance was significantly lower ( $p < 0.001$ ) than that of OIH (Fig. 1A) but correlated well ( $y = 26.134 + 0.4409x$ ,  $r = 0.92$ ,  $p < 0.001$ ). The distribution volume of the 44-min postinjection of MAG<sub>3</sub> was significantly lower than that for OIH (range 14–59 liters and 17–136 liters, respectively) but correlated well ( $y = 9.001 + 0.371x$ ,  $r = 0.82$ ,  $p < 0.001$ ), see Figure 1B. The excretion rate at 60 min postinjection showed no significant difference between MAG<sub>3</sub> and OIH as seen in Figure 1C where the excretion values are near the line of identity.

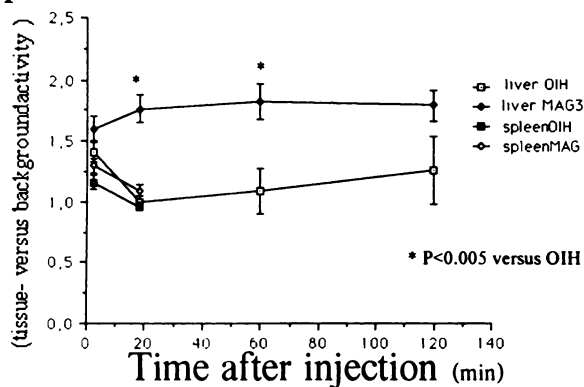
The activity in blood cells was measured in eight



**FIGURE 4**

(A) Images and (B) renograms of a patient referred for evaluation of suspected urinary tract obstruction. The patient suffered from end-stage renal failure due to recurrent urinary tract infection due to an old spinal cord injury. Significant liver uptake can be seen, particularly in the MAG<sub>3</sub> study (lower panel). Obstruction is excluded by the fact that injection of furosemide shows improved the elimination of the tracers.  $C_{OIH}$  and  $C_{MAG_3}$  give plasma clearance values.

## Uptake



**FIGURE 5**

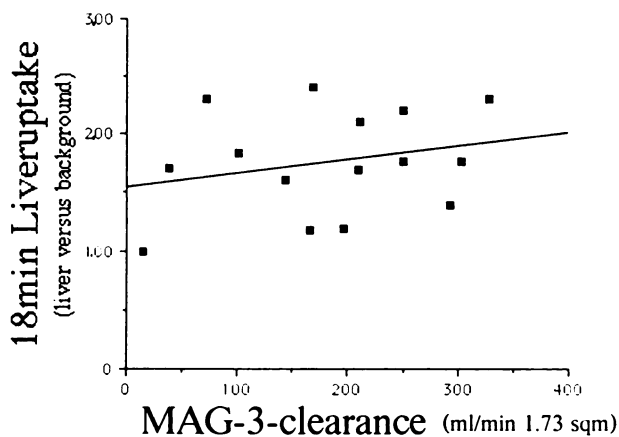
Uptake of [ $^{123}\text{I}$ ]OIH and  $\text{MAG}_3$  during the time after injection in liver and spleen. Both at 20 and 60 min postinjection, the  $\text{MAG}_3$  liver activity is significantly higher than 2–4 min postinjection compared to [ $^{123}\text{I}$ ]OIH. For [ $^{123}\text{I}$ ]OIH and  $\text{MAG}_3$ , there is a decrease in spleen activity with time, excluding specific uptake.

patients. For  $\text{MAG}_3$ , it was  $7.4\% \pm 1\%$  of the whole-blood activity as compared to  $26.9\% \pm 3.5\%$  for OIH ( $p < 0.01$ ).

### Gamma Camera Studies

Figure 2 shows analytic data of the acquired gamma camera studies. Both calculations of right kidney separate function (Fig. 2A), time-to-peak on renogram (Fig. 2B), and background ratio (Fig. 2D) were not significantly different between  $\text{MAG}_3$  and OIH. Elimination of the tracer as measured by  $T_{\text{max}1/2}$  was slower in  $\text{MAG}_3$  than in OIH, ( $p < 0.001$ ), seen as values above the line of identity in Figure 2C.

Figures 3, 4, and 7 show three examples from different patients. The images obtained with  $\text{MAG}_3$  or OIH looked quite similar. In Figure 3, a patient with hypertension shows normal images, renograms, and clear-



**FIGURE 6**

Absence of linear correlation ( $r = 0.25$ ) between the renal function, i.e.,  $\text{MAG}_3$  clearance (abscissa) and the 18–20-min liver uptake of  $\text{MAG}_3$  (ordinate).

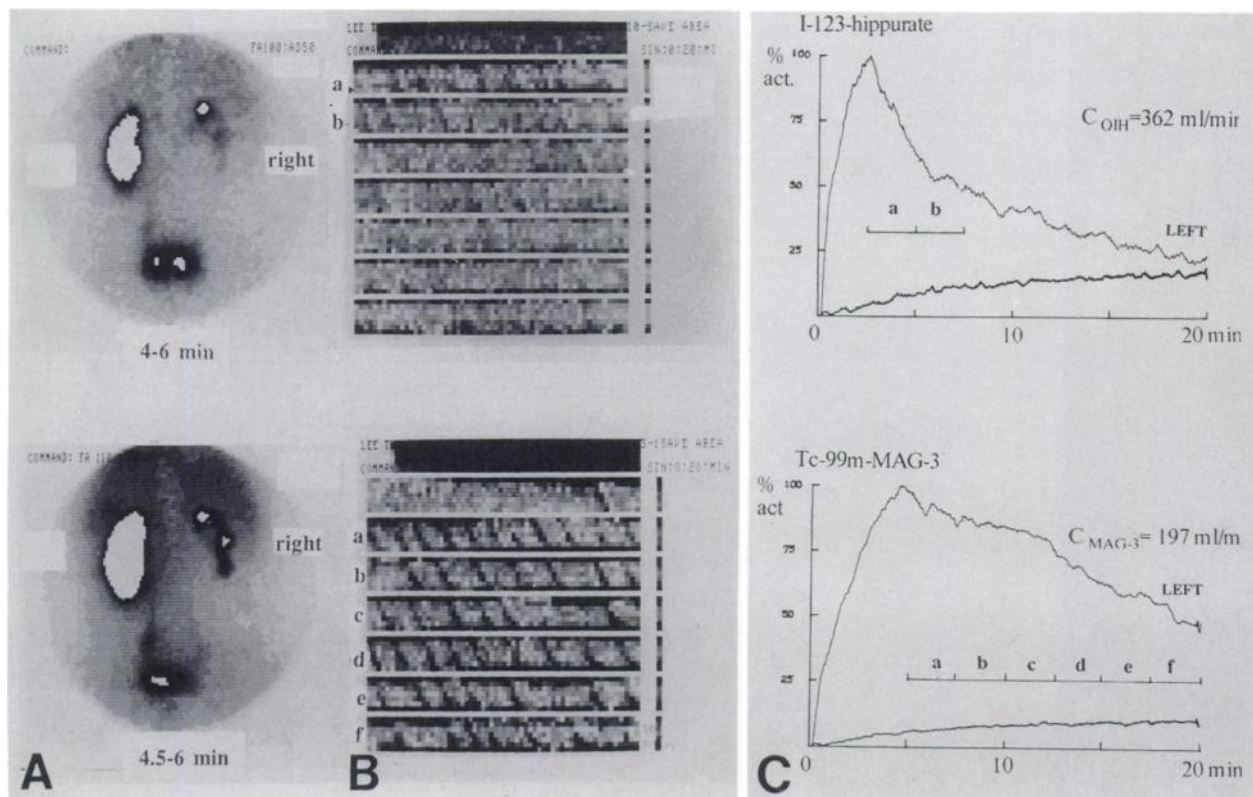
ances for both substances. Figure 4, a study of a patient with end-stage renal failure due to recurrent infections after spinal cord injury 15 yr earlier, demonstrates considerable liver uptake of both OIH and  $\text{MAG}_3$ , but particularly  $\text{MAG}_3$ . Figure 5 gives the mean values  $\pm$  s.e.m. of spleen and liver uptake versus time in all patients and demonstrates a specific  $\text{MAG}_3$  liver uptake,  $\sim 1.5$ – $2.0$  times background at 20–120 min postinjection which was not the case for OIH. This uptake was not correlated to kidney function (Fig. 6).

Urinary peristalsis was visualized in patients with tracer elimination from the kidney to the bladder on screen in the “cine-mode” as well in the condensed form as a time-space matrix (16). In most patients, urinary peristalsis was visible more clearly using  $\text{MAG}_3$  than OIH, as demonstrated in Figure 7. This patient had earlier pyelonephritis with very low uptake function in the right kidney. The peristalsis of the left ureter is visualized more clearly by  $\text{MAG}_3$  than by OIH (compare Fig. 7B lower panel, row a–f corresponding to 5–20 min postinjection for  $\text{MAG}_3$ , with Fig. 7B upper panel, row a–b corresponding to 2.5–7.5 min for OIH).

### DISCUSSION

The present study sought to investigate the clinical value of a new radiopharmaceutical,  $\text{MAG}_3$ , recently introduced for renal studies (1,2) in patients with different degrees of renal function impairment with ERPF varying between 10 and 654 ml/min 1.73 sqm. We used OIH as reference substance for evaluation since  $\text{MAG}_3$  has been recommended as a substitute for hippurate. In the first reports, no clinically significant differences were found (9,10) besides the observation that  $\text{MAG}_3$  clearance was found to be lower than that for hippurate. Some authors recommend the use of a simple correction factor to obtain a value equal to the hippurate clearance, i.e., the ERPF (17), whereas others found  $\text{MAG}_3$  clearance not suitable for renal plasma flow measurements (9,18). In both the present study and others (3,9,12,13), there is evidence for extrarenal clearance of  $\text{MAG}_3$ . However, the exact amount of such an eventual extra renal clearance has not been measured, since it demands both plasma renal clearance measurements simultaneously—which has been done neither in the present study nor in other studies. Thus, the use of a simple correction factor of  $\text{MAG}_3$  plasma clearance to obtain renal plasma flow may be incorrect for that reason and also because of estimating errors (7,9,18).

Data from animal studies suggest that  $\text{MAG}_3$  and hippurate are secreted by the same transport system and are competitively inhibited by probenecid (2,19) and PAH (20). The lower total renal clearance of  $\text{MAG}_3$  is partly a result of lower glomerular filtration as demonstrated by glomerular micropuncture (21) due to higher protein binding. Also, a lower renal secretory



**FIGURE 7**

(A) Images, (B) time-space matrix, and (C) renograms of a patient with earlier pyelonephritis referred for evaluation of right kidney separate function, which is 3% in both the  $^{123}\text{I}$  study (upper panels) and the  $\text{MAG}_3$  study (lower panels). Activity lines in the time-space-matrix (B) indicate peristaltic contractions of the left ureter in both studies, which is more clearly seen with  $\text{MAG}_3$  (see rows a-f corresponding to 5–20 min postinjection, lower panel) than with [ $^{123}\text{I}$ ]OIH (see rows a and b, i.e., 2.5–7.5 min postinjection, upper panel). Also, elimination of the tracer is slower for  $\text{MAG}_3$ .

transport capacity for  $\text{MAG}_3$  (5,20,21) contributes to this difference. Thus, since more than tubular secretion rate determines the  $\text{MAG}_3$  clearance, it may not be appropriate to use it for measuring renal tubular excretion rates only, as recommended by Brandau et al. (20), especially since we do not know how the different factors are abolished.

Some authors characterize the renal handling of  $\text{MAG}_3$  by measuring the excretion rate, i.e., the amount of injected activity in the urine at a certain time post-injection or by whole-blood clearance. Whole-blood clearance for  $\text{MAG}_3$  was found to be the same or even higher (2,4) than that for hippurate. However, whole-blood clearance is determined both by the penetration process of the tracer into and from the blood cells and by the clearance of the tracer by the kidney. Calculation of renal clearance according to its definition by Smith, i.e., the amount of plasma which is cleared per unit of time, revealed, on the other hand, that  $\text{MAG}_3$  clearance is ~50% that of hippurate. This is in good agreement with earlier studies (3,4,7,9,10,13,18,20). The excretion rate is also influenced by other factors like volume distribution and the time course of exchange between distribution volume and transport process of excretion. The excretion rate in the present study and in the

literature was not significantly different for  $\text{MAG}_3$  and OIH, which is a result of a low renal clearance and low distribution volume for  $\text{MAG}_3$  and a high renal clearance and high distribution volume for OIH. Higher plasma protein binding and less penetration of  $\text{MAG}_3$  into cells as found in the present study and in earlier rat experiments by ourselves and others (10) are factors contributing to the lower distribution volume of  $\text{MAG}_3$ . Russell et al. (10) demonstrated no penetration of  $\text{MAG}_3$  into blood cells using the American kit of  $\text{MAG}_3$ , whereas we found some percentage within the cells using the European kit. A technical error, caused by not taking into account the activity in the plasma trapped between the cells using routine blood centrifugation in our case, may be one explanation for this difference. A very low distribution volume could be advantageous for perfusion studies in that  $\text{MAG}_3$  gives higher plasma activity in relation to the injected amount than OIH. The high spleen-liver activity of  $\text{MAG}_3$  at 2–4 min postinjection may be due to a high blood supply (see Fig. 5). However, in the present study we were unable to investigate perfusion properly since our dosage of  $\text{MAG}_3$  and OIH was too low. Transplanted kidneys with abdominal views and a higher dosage may be more suitable for that purpose.

The relative renal uptake was found to be similar for OIH and MAG<sub>3</sub> in accordance with earlier reports (2, 7,8,9,13), as was the time-to-maximum peak activity on the renogram since both substances are predominantly excreted by tubular secretion. The elimination of the tracer from the kidneys was, however, in the present study slower for MAG<sub>3</sub> than for OIH. This was evidently not explained by different urine flow rates. The lower renal clearance of MAG<sub>3</sub> might have been of importance.

Slow elimination together with high target-to-background ratios and the use of a high dose of MAG<sub>3</sub> may be factors which made it possible to visualize the ureters and their peristalsis. In most patients, ureteral peristalsis in the time-space matrix (16) could be demonstrated and followed more clearly with MAG<sub>3</sub> than with OIH (Fig. 7). Another difference in imaging quality was the higher liver uptake of MAG<sub>3</sub>, which has been observed earlier in rat experiments (3) and described in patients (9,7,13). Brandau et al. (20) did not observe liver uptake using HPLC-purified MAG<sub>3</sub> in humans. Thus, kit-prepared MAG<sub>3</sub> seems to differ in this respect, although bile excretion of HPLC-purified MAG<sub>3</sub> has been demonstrated to do so (3), and other possible factors may be involved. Since the specific liver uptake is slow and seen only in late pictures and not more than twice in background activity, it does not jeopardize reliable calculation of separate function of the right kidney and is, therefore, of limited clinical significance.

In summary, there are some significant differences in the renal handling, the plasma distribution, and the cell penetration of MAG<sub>3</sub> and OIH. MAG<sub>3</sub>, however, seems to have particular qualifications as a radionuclide for dynamic renal scintigraphy since it is easily available by kit for labeling with <sup>99m</sup>Tc and has a higher renal excretion rate than a glomerular marker like DTPA. Despite the lower clearance of MAG<sub>3</sub> in comparison to hippurate, its lesser cell penetration and its higher target-to-background ratio results in renal dynamic scintigrams of high quality. Thus, from a clinical point of view, MAG<sub>3</sub> has advantages over other radiopharmaceuticals especially in patients with low renal function who require acute investigations.

#### ACKNOWLEDGMENTS

This study was partly supported by research grants from the Karolinska Institute and from Mallinckrodt, Petten, Netherlands. The typing of the manuscript by Karin Hultberg is gratefully acknowledged. We are also grateful to all patients who made this study possible.

#### REFERENCES

1. Fritzbeg AR, Kasina S, Eshima D, Johnson DL. Synthesis and biologic evaluation of technetium-99m-MAG<sub>3</sub> as a hippuran replacement. *J Nucl Med* 1986; 27:111-116.
2. Taylor A, Eshima D, Fritzbeg AR, Christian PE, Kasina S. Comparison of I-131-OIH and technetium-99m-MAG<sub>3</sub> renal imaging in volunteers. *J Nucl Med* 1986; 27:795-803.

3. Müller-Suur R, Müller-Suur C. Renal and extrarenal handling of a new imaging compound (<sup>99m</sup>Tc-MAG<sub>3</sub>) in the rat. *Eur J Nucl Med* 1986; 12:438-442.
4. Coveney JR, Robbins MS. Comparison of technetium-99m-MAG<sub>3</sub> kit with HPLC-purified technetium-99m-MAG<sub>3</sub> and OIH in rats. *J Nucl Med* 1987; 12:1881-1887.
5. Coveney JR, Pilcher GD, Robbins MS. Tc-99m-MAG<sub>3</sub> and I-131-o-iodohippurate (OIH) transport kinetics in rat kidney slices. *Eur J Nucl Med* 1988; 14:256.
6. de Jong R, Nielsen J, van Steenberg J, et al. Results of ERBF and ERPF measurements in healthy dogs with two new radiopharmaceutical principles. *Contr Nephrol* 1987; 56:49-52.
7. Taylor A, Ziffer JA, Stevens A, et al. Clinical comparison of I-131-orthoiodohippurate and the kit formulation of Tc-99m-mercaptoacetyltriglycine. *Radiology* 1989; 170:721-725.
8. Bubeck B, Brandau W, Steinbaeher M, et al. Technetium-99m-labeled renal functioning and imaging agents. II. Clinical evaluation of <sup>99m</sup>Tc-MAG<sub>3</sub> (<sup>99m</sup>Tc-mercaptoacetyl-glycylglycine). *Nucl Med Biol* 1988; 15:109-118.
9. Jafri RA, Britton KE, Nimmon CE, et al. Technetium-99m-MAG<sub>3</sub>: a comparison with iodine-123 and iodine-131-orthoiodohippurate in patients with renal disorders. *J Nucl Med* 1988; 29:147-158.
10. Russell CD, Thorstad B, Yester MV, et al. Comparison of Tc-99m-MAG<sub>3</sub> with I-131-hippuran by a simultaneous dual-channel technique. *J Nucl Med* 1988; 29:1189-1193.
11. Thorstad BL, Russell CD, Dubovsky EV, et al. Abnormal Captopril renogram and a technetium-99m-labeled hippuran analog. *J Nucl Med* 1988; 29:1730-1737.
12. Martin WG, Wolf IH, Kempken K, et al. Tc-99m-MAG<sub>3</sub> versus I-123-OIH: comparison of quantitative parameters in renal function. In: Schmidt HEA, Buraggi GL, eds. *Nuclear medicine*. Stuttgart-New York: Schattauer Verlag; 1989:413-416.
13. Al-Nahhas AA, Jafri RA, Britton KE, et al. Clinical experience with <sup>99m</sup>Tc-MAG<sub>3</sub>, mercaptoacetyltriglycine, and a comparison with <sup>99m</sup>Tc-DTPA. *Eur J Nucl Med* 1988; 14:453-462.
14. Ensing GJ, Nielsen J, Panek-Finda H, Panek K. Development of a kit formulation for the labelling of mercaptoacetyl triglycine (MAG<sub>3</sub>) with <sup>99m</sup>Tc. *Eur J Nucl Med* 1988; 14:303.
15. Tauxe WN, Dubovsky EV, Kidd TE. Comparison of measurement of effective renal plasma flow by single plasma sample and plasma disappearance slope/volume methods. *Eur J Nucl Med* 1985; 9:443-445.
16. Müller-Schauenburg W, Hofmann U, Feine U, et al. Criteria for ureteral obstruction by functional imaging of the upper urinary tract. *Contr Nephrol* 1987; 56:225-231.
17. Jafri RA, Nimmon CC, Britton KE, et al. Tc-99m-mercaptoacetyl-triglycine, MAG<sub>3</sub>, a comparison with <sup>131</sup>I- and I-123-orthoiodohippurate for routine renal work. *J Nucl Med* 1987; 28:647.
18. Schaap GH, de Jong RBJ, Alferink THR, et al. <sup>99m</sup>Tc-MAG<sub>3</sub> is not suitable for determination of the effective renal plasma flow. *Kidney Int* 1987; 32:775.
19. Müller-Suur R, Müller-Suur C. Renal and extrarenal excretion of a new renal scanning agent: <sup>99m</sup>Tc-MAG<sub>3</sub> in comparison to <sup>125</sup>I-hippurate (OIH) and <sup>51</sup>Cr-EDTA. *Abstract Eur Nucl Med Congr* 1986.
20. Brandau W, Bubeck B, Berger J, et al. Excretion characteristics of Tc-99m-MAG<sub>3</sub> in patients. In: Schmidt HEA, Buraggi GL, eds. *Nuclear medicine*. Stuttgart-New York: Schattauer Verlag; 1989:409-412.
21. Müller-Suur R, Müller-Suur C. Glomerular filtration and tubular secretion of MAG<sub>3</sub> in the rat. *J Nucl Med* 1989; in press.