INM/ DIAGNOSTIC NUCLEAR MEDICINE

Tc-99m Methylene Diphosphonate Versus Tc-99m Pyrophosphate: Biologic and Clinical Comparison

Thomas G. Rudd, David R. Allen, and David E. Hartnett

University of Washington, Seattle, Washington

The biologic and imaging characteristics of Tc-99m MDP and Tc-99m PP_i were compared in animals and patients using freeze-dried bone-imaging kits. Biodistribution data in rabbits showed Tc-99m MDP had slightly higher bone uptake, significantly lower blood levels, and faster urinary excretion compared with Tc-99m PP_i. Duplicate studies performed on ten patients showed the following: (a) blood clearance of Tc-99m MDP was more prompt and complete, resulting in significantly lower blood levels at 4 hr; (b) urinary excretion was greater with Tc-99m MDP than with Tc-99m PP_i; and (c) Tc-99m PP_i showed significant red-cell labeling, whereas Tc-99m MDP did not. Image quality was generally better with Tc-99m MDP than with Tc-99m the two agents.

J Nucl Med 18: 872-876, 1977

Since Subramanian's description of Tc-99m polyphosphate for bone imaging in 1971 (1-3), a number of Tc-labeled phosphate and phosphonate boneimaging compounds have been introduced (4-6). The agents receiving the widest clinical use include Tc-99m complexes of polyphosphate, pyrophosphate (PP₁), ethylene hydroxy diphosphonate (HEDP), and methylene diphosphonate (MDP). The relative merits of certain of these agents have been compared clinically and in animal models (7-10). The most comprehensive comparisons have been done by Subramanian and McAfee (11,12) and Davis and Jones (13); they suggest that Tc-99m MDP has the best overall characteristics for bone imaging.

We have used an "in house" preparation of Tc-99m pyrophosphate in our laboratories for several years and it has been a reliable and satisfactory agent. Since there were relatively sparse clinical data directly comparing Tc-99m PP₁ and Tc-99m MDP, we decided to perform a study comparing the two agents before converting to Tc-99m MDP for clinical bone imaging.

Because MDP was not commercially available at the time of the study, an MDP bone-imaging kit was prepared.* This report consists of two integral parts: (a) formulation and preparation of an MDP boneimaging kit and comparison with a PP_i kit in an animal model, and (b) clinical comparison of the biologic and imaging characteristics of the two agents.

MATERIALS AND METHODS

Radiopharmaceuticals. Lots of 100 units of MDP and PP₁ were formulated, freeze-dried, and sterilized by similar processes. Stannous methylene diphosphonate kits were prepared by dissolving 1 g of methylene diphosphonic acid in 200 ml of sterile pyrogenfree water. The methylene diphosphonic acid solution was combined with a solution of 100 mg of stannous chloride dihydrate dissolved in 1 ml of N hydrochloric acid. The solution was adjusted to a volume of 300 ml and pH of 7.0 by the addition of N sodium hydroxide solution. The solution was filtered through

Received Jan. 24, 1977; revision accepted Apr. 9, 1977.

For reprints contact: Thomas G. Rudd, Dept. of Nuclear Medicine, University of Washington Hospital, Seattle, WA 98195.

	Urine			Avg.			
	Whole body	(0–3 hr)	Liver	Spleen	skeleton	Whole blood	Soft tissue
Tc-99m PPi	81.8 ± 8.1	18.2 ± 8.1	20.5 ± 3.4	1.39 ± 0.40	44.9 ± 6.3	1.74 ± 0.53	1.26 ± 0.23
Tc-99m MDP	68.1 ± 13.2	31.9 ± 13.2	3.44 ± 2.5	0.18 ± 0.14	47.8 ± 11.2	0.73 ± 0.64	1.13 ± 0.53



FIG. 1. Comparative blood clearance; activity expressed as percent dose/liter. PP₁—pyrophosphate; MDP—methylene diphosphonate.

a $0.22-\mu$ Millipore filter and dispensed into 3-ml units and freeze-dried in a Virtis Model 800M freezedryer for 48 hr. The final shelf temperature of the process was 80°F. Immediately after freeze-drying, the vials were sealed under vacuum. This freeze-drying process was critical to satisfactory kit preparation. Less than 48 hr of drying resulted in significant liver uptake in the animal model.

Because the freeze-drying process is nonsterile, final sterilization was accomplished by cobalt-60 irradiation of the freeze-dried kits. A dose of 4 million rads was given in a single exposure. Comparison of irradiated and nonirradiated kits showed no detectable difference in labeling efficiency or biologic behavior.

Stannous pyrophosphate kits were prepared similarly, using 10 g of sodium pyrophosphate. The final molar ratios of phosphate to stannous chloride were 12.8:1 for methylene diphosphonate and 50.6:1 for pyrophosphate.

Routine quality control on all batches included pyrogen testing (USP rabbit pyrogenicity test and Limulus testing), determination of labeling efficiency (thin layer chromatography), and sterility tests (culture and colony count).

Animal studies. Biologic behavior of Tc-99m MDP and Tc-99m PP, was compared in New Zealand juvenile (1.5-2.3 kg) albino female rabbits. Six rabbits were given a quantitative intravenous injection of each radiopharmaceutical and killed 3 hr later. Doses were 0.5 mg MDP or 3.0 mg PP_i with 0.05 mg stannous chloride and 1 mCi Tc-99m/ kg body weight. Whole-body retention was estimated by a whole-body count immediately following injection and again after the bladder had been removed at necropsy. A flat-field collimated sodium iodide crystal was used at a distance of 1 m. A standard was counted at the time of whole-body counting to correct for physical decay. The liver was counted in similar fashion. The entire spleen, weighed samples of muscle (thigh) and bone (femur, with dried marrow removed), and 1 ml of blood were counted in a standard well counter along with appropriate standards. Activity was expressed as percent of administered dose. The sample activities were then extrapolated to whole-body retention based on the following fractions of body weight: blood 8%, bone 10%, and soft tissue 40%.

Patient studies. Ten cooperative patients referred for bone imaging were asked to participate, with no attempt made to select patients with or without bone disease. Paired bone studies were performed; one with Tc-99m PP₁ and one with Tc-99m MDP within a week. Technetium-99m PP₁ was the first agent used in five patients and Tc-99m MDP the first in the remaining five. Each patient received a calibrated injection of 15–20 mCi of Tc-99m-labeled MDP or PP₁; appropriate standards were prepared at the time of injection.

Urine collection for the 4 hr following injection and a blood sample at 4 hr after injection were obtained on all patients, and activity was expressed as percent administered dose and percent dose per liter, respectively. In five patients we also determined

	Urinary excretion: % dose 0-4 hr			Whole-blood activity: % dose/l @ 4 hr		
Patient No.	PPi	MDP	PP1/MDP	PP ₁	MDP	PP1/MD
1	36	44	0.82	1.1	0.3	3.7
2	32	47	0.68	2.1	1.0	2.1
3	45	51	0.88	2.1	0.7	3.0
4	18*	66		1.8	0.6	3.0
5	40	47	0.83	1.9	0.8	2.4
6	48	65	0.74	2.5	1.0	2.5
7	43	56	0.77	1.5	0.6	2.5
8	37*	30*		2.0	1.0	2.0
9	27*			2.5	1.2	2.1
10	47	63	0.75	1.4	0.5	2.8
Avg.	41	55	0.78†	1.9	0.75	2.6†

blood clearance of radioactivity from immediately following injection up to 4 hr.

Blood and urine samples were counted in a standard well counter with a sodium iodide crystal and single-channel analyzer, calibrated for the 140-keV peak of Tc-99m. Determination of plasma: RBC partition of radioactivity at 4 hr was done by measuring the hematocrit and counting whole blood and plasma.

Each patient received a standard whole-body scan (scintillation camera with whole-body imaging table) 4 hr after injection. In addition, serial images of the left shoulder and lumbar spine were obtained



FIG. 2. Shaded area indicates average fraction of blood radioactivity in plasma. Open area is RBC fraction. Values represent average of ten patients. Brackets indicate range.

at 1, 2,3, and 4 hr. The images were graded subjectively for overall image quality by three independent observers in two different ways. First, the 20 scans were graded for image quality on a scale from 1 (= poor) to 5 (= excellent), without knowledge of the patient or agent. Second, the two scans for each patient were compared without knowledge of which was MDP and which was PP₁ and were graded for relative image quality.

RESULTS

Animals. Images of the animals showed satisfactory bone labeling with both agents. Biologic data are shown in Table 1. Whole-body retention of MDP is lower, and urinary excretion higher, than that of PP₁. Soft-tissue activity (muscle) was similar with the two agents, but blood activity is significantly lower with MDP. The skeletal data suggest slightly higher MDP bone deposition, but the difference may not be significant. The relatively high PP₁ uptake by the liver is a characteristic of this agent in the rabbit.

Patients. The whole-blood clearances are shown in Fig. 1. The curves represent average values, with bars indicating range. Blood levels at 4 hr and cumulative 4-hr urinary excretion data are shown in Table 2. As expected, there was considerable interpatient variability, but in any given patient Tc-99m MDP blood activity was lower and urinary excretion higher than with Tc-99m PP₁. The plasma:RBC partitions at 4 hr are shown in Fig. 2. A significant fraction of Tc-99m PP₁ activity is associated with the red cells, whereas Tc-99m MDP is confined primarily to the plasma.

Both agents produced satisfactory bone images. Analysis of relative image quality was necessarily subjective, and although the differences were less impressive than in the measured data, the Tc-99m



FIG. 3. Comparative hourly images up to 4 hr. PP1—pyrophosphate; MDP—methylene diphosphonate.

MDP images were generally slightly sharper. Figure 3 compares serial images of the shoulder up to 4 hr, and Fig. 4 compares anterior whole-body images at 4 hr. Tables 3 and 4 show the results of the whole-body images at 4 hr. Tables 3 and 4 show the results of the whole-body image evaluation. Whether compared independently or one against the other, the Tc-99m MDP images were generally judged to be equal or superior to Tc-99m PP₁ images, although the independent evaluation differences (Table 3) were not statistically significant. Relative diagnostic sensitivity was not critically evaluated because of the small series and unavoidable differences in imaging technique. But there was no apparent differences in diagnostic sensitivity between the two agents.

DISCUSSION

The biologic behavior of Tc-99m PP₁ and Tc-99m MDP found in this study support the findings of Subramanian and McAfee using normal volunteers (11). They have indicated that the lower Tc-99m MDP blood levels and increased urinary excretion are due to reduced plasma protein labeling, compared with Tc-99m PP_1 (12). Our data indicate that these differences may be due primarily to absence of red-cell labeling by Tc-99m MDP, whereas Tc-99m PP₁ shows significant red-cell labeling. The phenomenon of in vivo red-cell labeling by pertechnetate following pyrophosphate administration has been reported previously (14-17), and is felt to be due to reduction of the pertechnetate by excess circulating tin complexes. Our experience suggests that a small fraction of Tc-99m pyrophosphate activity labels red cells rather promptly following intravenous administration. Whether this is due to direct Tc-99m pyrophosphate labeling or to reduction of free pertechnetate remains to be determined, although the former seems more likely.

Weber and Keyes (10) recently performed a comparative study and concluded Tc-99m pyrophosphate



FIG. 4. Comparative anterior whole-body images at 4 hr. PP₁pyrophosphate; MDP-methylene diphosphonate.

	No. scans	No. observations*	Avg. grade†‡
Tc-99m MDP	10	30	3.43 ± 0.81
Tc-99m PP1	10	30	2.93 ± 0.79

† Scale: 1 (poor) to 5 (excellent).

‡ Differences were not significant.



had the best overall characteristics for bone imaging. Unfortunately, Tc-99m methylene diphosphonate was not included in the agents studied. Their work is further compromised in that the patient studies were in series and not in duplicate, and the well-known interpatient biologic variability of these agents was not controlled. We feel that meaningful results can be obtained only if this variable is eliminated. For this reason we limited our study to two agents and performed paired studies with the patient serving as his own control. Krishnamurthy (7,8) and Citrin (9,18) have previously used this same approach when making clinical comparisons of boneimaging agents. Citrin (9,18) considers Tc-99m HEDP to be the bone-imaging agent of choice, but he has not compared it with Tc-99m MDP. We intend to perform such a clinical comparison shortly.

The advantages of Tc-99m MDP are several. The rapid blood clearance allows earlier imaging following injection, and we now routinely image at 3 hr with this agent. The lower blood levels offer improved target-to-background ratios and should increase diagnostic sensitivity. This was not apparent in our study, but the series is small. Because of the more rapid clearance and lower background, highresolution imaging of small bones and joints should be more effective with Tc-99m MDP. Considering the findings of this study, we have converted to Tc-99m MDP for routine bone imaging in our laboratories.

ACKNOWLEDGMENTS

The authors wish to thank Dan H. Chadwick for his technical assistance, and Drs. Wil B. Nelp and Michael J. Daly for reviewing the whole-body images.

FOOTNOTE

* This was done for us by the University of Washington Nuclear Pharmacy.

REFERENCES

1. SUBRAMANIAN G, MCAFEE JG: A new complex of ^{50m}Tc for skeletal imaging. *Radiology* 99: 192–196, 1971

2. SUBRAMANIAN G, MCAFEE JG, BELL EG, et al: ^{som}Tc labeled polyphosphate as a skeletal imaging agent. *Radiology* 102: 701-704, 1972

3. SUBRAMANIAN G, MCAFEE JG, O'MARA RE, et al: ⁹⁹TC polyphosphate PP₄₆: A new radiopharmaceutical for skeletal imaging. J Nucl Med 12: 399–400, 1971 (Abst)

4. WEBER DA, KEYES JW, BENDETTO WJ, et al: ^{®BET}C pyrophosphate for diagnostic bone imaging. *Radiology* 113: 131-137, 1974

5. YANO Y, MCRAE J, VANDYKE DC, et al: ^{90m}Tc labeled Sn(II)-diphosphonate: a bone scanning agent. J Nucl Med 13: 480, 1972 (Abst)

6. SUBRAMANIAN G, BLAIR RJ, KALLFELZ EA, et al: ^{****}Tc-MDP (Methylene Diphosphonate): A superior agent for skeletal imaging. J Nucl Med 14: 640, 1974 (Abst)

7. KRISHNAMURTHY GT, TUBIS M, ENDOW JS, et al: Clinical comparison of the kinetics of ^{90m}Tc labeled polyphosphate and diphosphonate. J Nucl Med 15: 848-855, 1974

8. KRISHNAMURTHY GT, HUEBOTTER RJ, WALSH CF, et al: Kinetics of ^{som}Tc labeled pyrophosphate and polyphosphate in man. J Nucl Med 16: 109-115, 1975

9. CITRIN DL, BLESSENT RG, TUOHY JB, et al: A comparison of phosphate bone scanning agents in normal subjects and patients with malignant disease. Br J Rad 48: 118-121, 1975

10. WEBER DA, KEYES JW, WILSON GA, et al: Kinetics and imaging characteristics of ^{som}Tc labeled complexes used for bone imaging. *Radiology* 120: 615-621, 1976

11. SUBRAMANIAN G, MCAFEE JG, BLAIR RJ, et al: Technetium-99m methylene diphosphonate—a superior agent for skeletal imaging: Comparison with other technetium complexes. J Nucl Med 16: 744-755, 1975

12. SUBRAMANIAN G, MCAFEE JG, BLAIR RJ, et al: An evaluation of ^{®m}Tc-labeled phosphate compounds as bone imaging agents. In *Radiopharmaceuticals*, New York, Society of Nuclear Medicine, 1975, pp 319-328

13. DAVIS MA, JONES AG: Comparison of ^{99m}Tc labeled phosphate and phosphonate agents for skeletal imaging. Semin Nucl Med 6: 19-31, 1976

14. STOKELY EM, PARKEY RW, BONTE FJ, et al: Gated blood pool imaging following ^{60m}Tc stannous pyrophosphate imaging. *Radiology* 120: 433–434, 1976

15. CHANDLER WM, SHUCK LD: Abnormal technetium-99m pertechnetate imaging following stannous pyrophosphate bone imaging. J Nucl Med 16: 518, 1975 (Abst)

16. KHENTIGAN A, GARRETT M, LUM D, et al: Effect of prior administration of Sn(II) complexes used in nuclear medicine on in vivo distribution of subsequently administered Tc-99m pertechnetate and technetium-99m compounds. J Nucl Med 16: 541, 1975 (Abst)

17. ZIMMER AM, PAVEL DG, PATTERSON VN: In vivo red blood cell labeling using consecutive injections of stannous pyrophosphate and technetium-99m pertechnetate. J Nucl Med 17: 566, 1976 (Abst)

18. CITRIN DL, BESSENT RG, GREIG WF: Clinical evaluation of ^{60m}Tc labeled monofluorophosphate: A comparison with ethane-hydroxy-diphosphonate. J Nucl Med 15: 1110-1112, 1974