

Characterization of ^{18}F -FPyKYNE-Losartan for Imaging AT_1 Receptors

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Most physiologic effects of the renin angiotensin system (RAS) are mediated via the angiotensin (Ang) type 1 receptor (AT_1R). The ^{18}F -FPyKYNE derivative of the clinically used AT_1R blocker losartan exhibits high binding selectivity for kidney AT_1R and rapid metabolism in rats. The aim of this study was to further assess the binding profile of this novel PET agent for imaging AT_1R in rats and pigs. **Methods:** In vitro binding assays were performed with ^{18}F -FPyKYNE-losartan in rat kidneys. Male Sprague-Dawley rats were used to assess dosimetry, antagonistic efficacy via blood pressure measurements, and presence of labeled metabolites in kidneys. Test-retest PET imaging, blocking with AT_1R antagonist candesartan (10 mg/kg), and plasma metabolism analysis were performed in female Yorkshire pigs. **Results:** ^{18}F -FPyKYNE-losartan bound with high affinity (dissociation constant of 49.4 ± 18.0 nM and maximal binding of 348 ± 112 fmol/mm²) to rat kidney AT_1R . It bound strongly to plasma proteins in rats (97%), and its labeled metabolites displayed minimal interference on renal AT_1R binding. FPyKYNE-losartan fully antagonized the Ang II pressor effect, albeit with 4-fold potency reduction (the effective dose inhibiting 50% of the Ang II-induced maximal pressor response of 25.5 mg/kg) relative to losartan. PET imaging exhibited high kidney-to-blood contrast and slow renal clearance, with an SUV of 14.1 ± 6.2 . Excellent reproducibility was observed in the calculated test-retest variability ($7.2\% \pm 0.75\%$). Only hydrophilic-labeled metabolites were present in plasma samples, and renal retention was reduced ($\sim 60\%$) at 10–15 min after blockade with candesartan. **Conclusion:** ^{18}F -FPyKYNE-losartan has a favorable binding profile and displays high potential for translational work in humans as an AT_1R PET imaging agent.

Key Words: RAS; AT_1R specific binding; metabolism; dosimetry; PET imaging studies in rats and pigs

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Hyperactivity of the renin angiotensin system (RAS), and specifically the pathologic alteration in angiotensin II (Ang II)

type 1 receptor (AT_1R), is thought to play a distinct role in the development of disease processes including hypertension, myocardial infarction, chronic kidney disease, and cardiorenal failure (1–3). Conflicting results have been reported on AT_1R levels measured under pathophysiologic conditions. Although previous PET studies have been performed with small (rodents) and larger (dogs, baboons, pigs) animals (4–6) using ^{11}C -labeled AT_1R ligands, no research has been published using an ^{18}F -labeled AT_1R ligand. ^{18}F -labeled tracers offer some key advantages over ^{11}C , including a longer half-life (109.6 vs. 20.4 min) (7), enabling multiple scans from a single formulation and shipment to other sites and higher image resolution due to lower energy and shorter positron range (0.23 vs. 0.39 mm in soft tissue) (8), thus enabling higher image resolution.

Previous structure–activity relationship studies demonstrated that large prosthetic groups can be introduced at the imidazole 5-position of losartan with minimal changes in binding properties (9). Moreover, addition of a leucine and a short linker composed of tetraethylene glycol and diglycolic acid between the losartan moiety and the tetra-amine chelator yielded to a losartan analog displaying high affinity (inhibitory constant = 6 nmol/L) for AT_1R (10). Recently, ^{18}F -FPyKYNE-losartan was synthesized by conjugation of ^{18}F -FPyKYNE with the azide-modified trityl losartan via click chemistry with high chemical and radiochemical purities (11). Preliminary in vivo evaluation of ^{18}F -FPyKYNE-losartan with PET imaging displayed a dose-dependent reduction of this radiotracer in rat kidney AT_1R s and binding selectivity for AT_1R over AT_2R . Additionally, ^{18}F -FPyKYNE-losartan is metabolized into mostly hydrophilic-labeled products in rat plasma, suggesting minimal interference of the ^{18}F -labeled metabolites with AT_1R binding (11).

The specific aims of the present study were to evaluate the binding affinity of ^{18}F -FPyKYNE-losartan to renal AT_1R s and antagonistic efficacy on Ang II pressor effect in rats; the relative proportions of labeled metabolites (if any) in rat kidney and pig plasma, compared with unchanged tracer using column-switch high-performance liquid chromatography (HPLC); the biodistribution and radiation dosimetry; and the PET imaging profile of ^{18}F -FPyKYNE-losartan in pigs.

MATERIALS AND METHODS

Animals

All animal experiments were conducted in accordance with the Canadian Council on Animal Care guidelines and approved by the Animal Care Committee of the University of Ottawa. Male and female (for dosimetry) Sprague-Dawley rats (200–300 g) (Charles River) were housed in pairs and maintained on a 12-h light–dark cycle with

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free access to food and water. Female Yorkshire pigs (30–35 kg) (Panmure Farms) were kept for 1 wk on arrival before any studies were performed and fed with a standard diet.

Radiochemistry

^{18}F -FPyKYNE-losartan (specific activity, 7.4–155 GBq/ μmol) and unlabeled FPyKYNE-losartan (Fig. 1) were produced in high purity as reported previously (11).

Rat Studies

In Vitro Evaluation of Binding Parameters. After decapitation, rat kidneys were snap-frozen in 2-methylbutane at -40°C and stored at -80°C for autoradiography ($n = 5$ assays) (12–14). Briefly, 20- μm kidney sections mounted onto slides (3 per slide) were preincubated in 150 mM NaCl, 1 mM ethylenediaminetetraacetic acid, 50 mM Na_2PO_4 , 0.1% bovine serum albumin, and 0.1 mM bacitracin for 15 min, and then increasing concentrations of ^{18}F -FPyKYNE-losartan (1–500 nM) were added in the presence of AT_2R blocker PD123,319 (10^{-5} M; Sigma) for 1 h at room temperature. To determine nonspecific binding (NSB), parallel incubations were performed with the addition of 10 μM unlabeled Ang II (Sigma) (12–14). After 3 successive 15-min washes, dry slides were loaded into x-ray cassettes with a set of radioactivity standards and exposed to an imaging phosphor screen for 1 h. Screens were imaged with the Molecular Imager FX (Bio-Rad) and the binding densities quantified with a computer-assisted image analysis system (QuantityOne). The optical density value of each pixel of digitized image was converted into DPM/ mm^2 , and nonlinear curve-fitting of total binding and NSB isotherms was performed to determine the dissociation constant (K_D , nM) and maximal binding (fmol/ mm^2) of radiotracer using GraphPad Prism (version 6.02; GraphPad Software).

In Vivo Assessment of Antagonistic Activity. Rats were anesthetized with sodium pentobarbital (60 mg/kg, intraperitoneally), then the carotid artery was cannulated and connected to a fiber-optic pressure transducer for blood pressure measurement. Drugs were injected through a catheter in the tail vein. Animals were kept warm at 37°C .

To obtain a dose-pressor response curve for Ang II and determine its submaximal dose, Ang II (0.03–100 $\mu\text{g}/\text{kg}$) was administered intravenously in a cumulative manner ($n = 4$), and each successive injection was given immediately after the maximal effect of the preceding dose was obtained (10–20 s). Saline vehicle was given to animals 15 min before and after Ang II (15,16). All drugs were in-

jected in a 0.2- to 0.4-mL volume. Rats were euthanized by an overdose of sodium pentobarbital (90 mg/kg, intravenously) (17). The effects of FPyKYNE-losartan (3–50 mg/kg; $n = 1$ –3) and the AT_1R blockers candesartan (1–10 mg/kg; $n = 1$ –3) and losartan (1–30 mg/kg; $n = 1$ –3) were examined.

Doses were injected 15 min before the administration of Ang II at its submaximal dose, and then dose–response curves were plotted and the percentage decrease in diastolic blood pressure was calculated for each test compound in relation to that obtained in untreated animals. Inhibitory dose–response curves were best-fitted with a 4-parameter logistic equation under a shared Hill slope using GraphPad Prism 6.02 to determine the effective dose inhibiting 50% of the Ang II–induced maximal pressor response and the percentage antagonist-mediated maximal inhibition in vivo.

Plasma-Protein Binding. Plasma-protein binding of ^{18}F -FPyKYNE-losartan was measured in quintuplicate using the Centrifree Ultrafiltration device (Millipore; molecular weight cutoff, 30,000 Da). Unchanged ^{18}F -FPyKYNE-losartan was added to plasma obtained from rats ($n = 5$) sacrificed by decapitation without anesthesia. Plasma was centrifuged (2,000g, 30 min) and the filtrate (protein-free fraction) counted in the γ -counter (18,19).

Radiolabeled Metabolite Analysis in Kidneys. ^{18}F -FPyKYNE-losartan metabolism was examined in normal rat kidney homogenates at 5, 10, 20, and 30 min ($n = 3$ per time point) after the injection of 74–148 MBq as previously reported (11). Blocking studies were performed with candesartan (5 mg/kg, $n = 4$; and 10 mg/kg, $n = 5$) via the tail vein immediately before ^{18}F -FPyKYNE-losartan, and rats were sacrificed at 10 min after tracer injection. Tissue samples were homogenized in ethanol/water 80/20 (v/v) using a polytron homogenizer and ultracentrifuged for 15 min. Supernatant was collected and evaporated, reconstituted in acetonitrile/water 1/99 (v/v) containing urea (0.4 g/mL) to disrupt plasma-protein binding, filtered (0.2- μm Nylon Membrane [Acrodisc]), and injected into the HPLC system. Tissue homogenates ($n = 3$ at -5 min) spiked with approximately 1.85 MBq of ^{18}F -FPyKYNE-losartan were also tested by column-switch HPLC as controls.

Estimation of Human Effective Doses. Human dosimetry for ^{18}F -FPyKYNE-losartan was calculated using whole-body biodistribution data extrapolated from rats, following the Radiation Dose Assessment Resource method as implemented in the OLINDA/EXM 1.1 program (Vanderbilt University), and using tissue-weighting factors from the International Commission on Radiological Protection (ICRP) protocols (20,21). A detailed description of the methods is provided in the supplemental materials (available at <http://jnm.snmjournals.org>).

Pig Studies

PET Imaging. Pigs were anesthetized with Telazol (tiletamine-zolazepam, 0.044 mg/kg; Zoetis Inc.), glycopyrrolate (0.01 mg/kg), and isoflurane and placed supine for PET imaging using a Discovery D690 PET-VCT scanner (GE Healthcare). A CT scout scan was obtained, followed by a low-dose CT for attenuation correction and a 10-min ^{82}Rb scan to ensure that both kidneys were in the scanner field of view. ^{18}F -FPyKYNE-losartan was administered via the ear vein at a dose of 5 MBq/kg. Vital data were monitored continuously. Pigs were rescanned the following week to ensure repeatability of results ($n = 3$). On the third week of each pig study, blocking PET scans were obtained by injecting (intravenously) a saturating dose of candesartan (10 mg/kg) 15 min before tracer injection. PET imaging was performed for 90 min and images reconstructed into 12×10 s, 6×20 s, and 16×5 min frames, using the vendor ordered-subset expectation maximization method with 8-mm postfiltering.

PET Image Analysis. Siemens Inveon Research Workplace software was used to analyze reconstructed dynamic images and generate time–activity curves for the arterial blood input function (derived from aorta regions) and right kidney tissue uptake. Renal CT angiography images

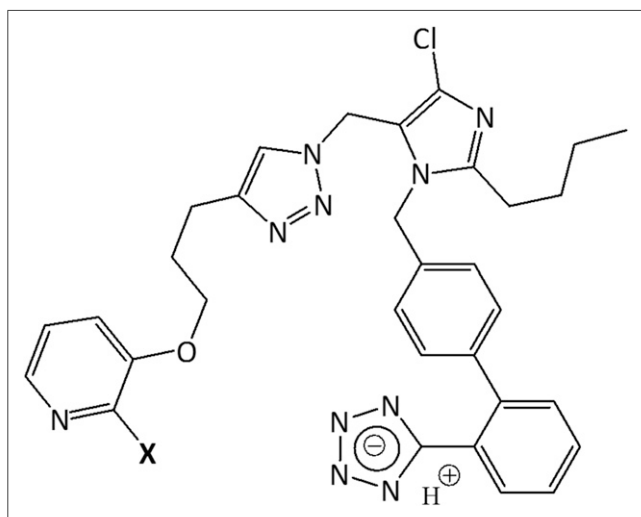


FIGURE 1. Chemical structures of ^{18}F -FPyKYNE-losartan (X: F-18) and cold standard, FPyKYNE-losartan (X: F-19).

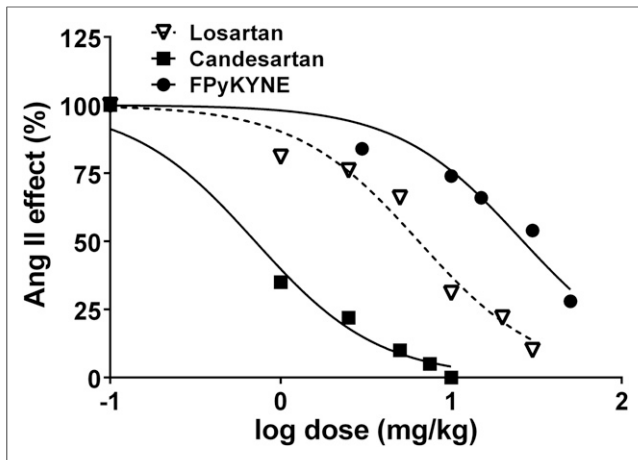


FIGURE 2. Dose–response curves of AT₁R blockers candesartan, losartan, and FPYKYNE-losartan displaying their effect in blocking Ang II pressor response (submaximal dose, 3 μ g/kg intravenously) in anesthetized rats. Curves were analyzed with nonlinear curve-fitting program (GraphPad Prism 6.02) using 4-parameter logistic equation. Curves were fitted with shared Hill slope factor.

were coregistered with their corresponding PET scans, to ensure accurate localization of regions of interest. Arterial time–activity curves were corrected for partial-volume losses based on a computer simulation of the PET image resolution and the aorta radial dimension measured on the CT angiography images. ¹⁸F-FPYKYNE-losartan renal activity was measured as SUV (g/mL) at 10–15 min after injection (frame 20) (22). The specific binding index was expressed as the reduced peak retention in the renal cortex and was calculated from the difference in peak SUVs between baseline and blocking conditions.

Radiolabeled Metabolite Analysis in Plasma. During the PET scans, blood samples were collected from the femoral artery at –5 (control), 1, 2, 5, 10, 20, and 40 min after injection ($n = 3$). Plasma samples were prepared as described previously (11,19). Control plasma was spiked with 0.37 MBq of tracer, filtered, and then injected onto the HPLC system.

Arterial Input Function Corrections. The first step toward generating an arterial input function was correcting for plasma radioactivity. To correct for binding to red blood cells, activity in whole blood and in plasma was measured at serial time points, and the plasma–to–whole-blood ratio was determined as a function of time. ¹⁸F-FPYKYNE-losartan (5 MBq/kg) was injected via the ear vein and arterial blood (5–6 mL) collected from the femoral artery at 0.5, 1, 2, 5, 10, 20, 40, 60, and 90 min. Blood samples were weighed and counted for activity. Samples were centrifuged to obtain plasma and then weighed and counted to determine activity per gram (19).

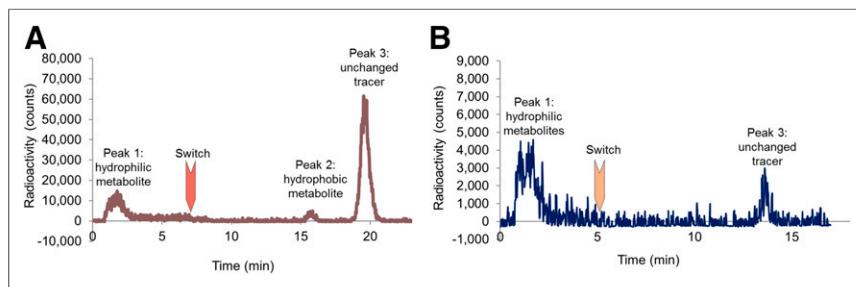


FIGURE 3. Representative high-performance liquid chromatograms displaying unchanged ¹⁸F-FPYKYNE-losartan (peak 3) and its labeled hydrophilic (peak 1) and hydrophobic (peak 2) metabolites in rat kidney (A) and pig plasma (B) at 10-min time point.

Statistical Analysis

All results are expressed as mean \pm SD or \pm SE as indicated. Means were compared using the *t* test (paired or unpaired). The test–retest variability was calculated by taking the ratio of the absolute value of the difference between repeated measurements and the mean of the repeated measurements as described by Lortie et al. (19).

RESULTS

In Vitro Binding in Rat Kidneys

¹⁸F-FPYKYNE-losartan binding displayed a discrete localization in the kidney. The highest level of total binding relative to NSB was detected in the kidney cortex. Global curve-fitting analyses of paired total binding and NSB curves revealed that ¹⁸F-FPYKYNE-losartan bound to kidney cortex with a K_D of 49.4 ± 18.0 nM and maximal binding of 348 ± 112 fmol/mm².

In Vivo Antagonism in Rats

Ang II caused a dose-dependent increase in blood pressure, and the submaximal dose was determined graphically to be 3 μ g/kg. Losartan and candesartan exhibited a dose-dependent antagonism of AT₁Rs and virtually abolished the Ang II effect at doses of 30 and 10 mg/kg, respectively. The blocking effect of FPYKYNE-losartan obtained at a dose of 50 mg/kg was 72%. The computed effective doses inhibiting 50% of the Ang II–induced maximal pressor response values (in mg/kg) obtained with the curve-fitting analysis of inhibitory dose–response curves were 0.7 (candesartan), 6.4 (losartan), and 25.5 (FPYKYNE-losartan) (Fig. 2). FPYKYNE-losartan also displayed full antagonism but with less potency than losartan and candesartan.

Radiolabeled Metabolite Analysis in Rat Kidneys

¹⁸F-FPYKYNE-losartan was bound at 97% to plasma proteins. As described in rat plasma (11), kidney samples revealed 3 major HPLC peaks (Fig. 3A). The hydrophilic-labeled metabolite (peak 1) increased slowly over time, whereas the hydrophobic metabolite (peak 2) had a negligible contribution to the total activity. Unchanged ¹⁸F-FPYKYNE-losartan (peak 3) accounted for more than 50% of total radioactivity at all time points (Table 1). Blockade of AT₁Rs with candesartan produced a reduction in unchanged tracer proportion by 74% and 88% with the 5 and 10 mg/kg doses, respectively. The proportion of hydrophobic metabolite decreased by half at a blocking dose of 10 mg/kg, whereas hydrophilic metabolite proportion increased by 86% at the same blocking dose (Table 1).

Rat Dosimetry Studies

The decay-corrected accumulation of radioactivity in whole tissues is depicted in Supplemental Table 1. Most tissues demonstrated a rapid tracer uptake, with levels gradually decreasing after 5 min. Similar trends were observed for both men and women. Elevated radioactivity accumulation with time occurred within the metabolism-related tissues (liver, small intestine contents, upper and lower intestine contents, and urine). The liver showed the highest uptake and accounted for around 30% of the effective dose (ED) in both sexes (Supplemental Table 2). The sex-averaged ED calculated using both ICRP 60 and 103 was 0.031 mSv/MBq. A mean of 85% of the total injected dose was recovered in the organs and carcass at the time points studied.

TABLE 1
Proportions of Unchanged ^{18}F -FPyKYNE-Losartan and Its Labeled Metabolites in Rat Kidney Homogenate at Respective Time Points

Peak	Identity	0 min	5 min	10 min*			20 min	30 min
				0 mg/kg	5 mg/kg	10 mg/kg		
1	Hydrophilic metabolite	0 ± 0	5 ± 4	12 ± 1	76 ± 20	88 ± 15 ^{†‡}	17 ± 8	39 ± 20
2	Hydrophobic metabolite	0 ± 0	2 ± 1	4 ± 2	2 ± 2 [†]	2 ± 0 [†]	5 ± 2	3 ± 2
3	Unchanged tracer	100 ± 0	93 ± 4	84 ± 2	22 ± 20 [†]	10 ± 14 [†]	78 ± 10	58 ± 19

*Data are given for 3 different candesartan blocking doses.

[†] $P < 0.05$, compared with baseline.

[‡]Compared with previous dose.

PET Imaging of Pig Kidneys

Time-activity curves derived from the right kidney exhibited high kidney-to-blood contrast (ratio of approximately 3) and slow clearance from the kidneys (Fig. 4A). The highest renal uptake was obtained at 10–15 min after injection, with an SUV of 14.1 ± 6.15 . Test and retest scans were reproducible with a test-retest variability of $7.2\% \pm 0.75\%$ and a P value of 0.89, showing no significant difference between the datasets. Blocking AT_1Rs with 10 mg/kg of candesartan resulted in reduced renal ^{18}F -FPyKYNE-losartan retention (Fig. 4B). SUV_{max} (at 10–15 min) in the right kidney was significantly decreased from 14.1 ± 6.15 to 5.8 ± 4.60 ($P = 0.03$). This 60% reduction demonstrates specific binding of ^{18}F -FPyKYNE-losartan to AT_1Rs in pig kidneys. The arterial plasma-to-whole-blood ratio (1.4) was almost constant over time (0–90 min), facilitating input function correction. However, further corrections are necessary for accurate quantification, including correction for labeled metabolites and plasma-protein binding.

Radiolabeled Metabolite Analysis in Pig Plasma

Only peak 1 (hydrophilic metabolite) and peak 3 (unchanged tracer) were present in pig plasma in both normal and blocking conditions (Fig. 3B). At 20 min, less than 10% of radioactivity (noise- and decay-corrected) in normal plasma was associated with unchanged tracer (Table 2). Blocking AT_1Rs resulted in a faster metabolism in pig plasma at 20 min (Table 2). A significant difference in the proportion of unchanged ^{18}F -FPyKYNE-losartan between normal and blocking conditions was identified at 5, 10, and 20 min ($P = 0.008, 0.02, \text{ and } 0.04$, respectively).

DISCUSSION

Successful development of a new noninvasive ^{18}F -labeled molecular imaging probe for quantifying AT_1Rs would present a

unique opportunity to advance understanding and contribution of AT_1Rs to the progression of cardiovascular and renal diseases and a means to directly measure therapy responses to optimize patient outcomes. ^{18}F -FPyKYNE-losartan small-animal PET images obtained previously (11) displayed high tissue contrast in the kidney cortex and outer medulla, which correlates well with the known physiologic distribution of AT_1R (24). Nonetheless, the cortical and medullary binding could not be differentiated, which can be attributed to the image resolution of the small-animal PET. In the current study, FPyKYNE-losartan exhibited high in vitro binding affinity to AT_1R in the renal cortex and full in vivo antagonism, similar to its parent compound losartan. However, a higher dosage of FPyKYNE-losartan was required for blocking Ang II pressor response relative to losartan and candesartan (4 and 36 times more potent, respectively). The order of potency of candesartan and losartan in vivo is consistent with the in vitro binding affinities of these AT_1R blockers reported previously (13,25).

Binding of a drug to plasma and tissue proteins inhibits drug disposition and has important effects on drug dynamics because only the free (unbound) drug interacts with receptors. ^{18}F -FPyKYNE-losartan exhibits similar plasma-protein binding to its parent compound losartan (97% and 98%, respectively). Thus, only 3% of ^{18}F -FPyKYNE-losartan is freely available for binding to tissue AT_1R . ^{18}F -FPyKYNE-losartan kidney metabolite analysis displayed slow accumulation of the hydrophilic-labeled metabolite and the presence of almost 60% unchanged tracer at 30 min after injection. Blocking AT_1R with candesartan at 10 min reduced the proportion of the parent compound, whereas a significant increase was observed in the proportion of the hydrophilic metabolite. These results indicate that the portion of the signal corresponding to AT_1R -specific binding is mostly from the unchanged tracer because of the proportion of the labeled metabolites that is not reduced after AT_1R blockade in rat kidneys.

Rat dosimetry data suggest that the hepatobiliary system is primarily responsible for eliminating ^{18}F -FPyKYNE-losartan from the body, whereas the kidneys are the secondary route of excretion. Losartan is known to be primarily eliminated via the cytochromes-P450 system in the liver (26,27). As a major site for drug metabolism, the liver aids in the elimination

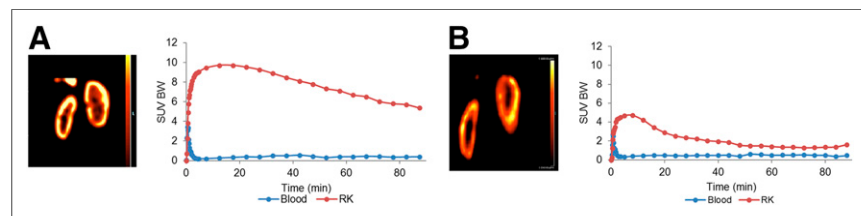


FIGURE 4. Representative PET image (coronal view) of pig kidneys displaying ^{18}F -FPyKYNE-losartan uptake in normal (A) and blocking (B) conditions. Tracer time-activity curves for blood input (aorta) and right kidney (RK) are presented as SUVs normalized to body weight (SUV_{BW}) from 0 to 90 min.

TABLE 2
Proportions of ¹⁸F-FPyKYNE-Losartan and Its Hydrophilic-Labeled Metabolites in Pig Plasma in Normal and Blocking Conditions

Condition	Identity	Time point (min)						
		0	1	2	5	10	20	40
Baseline	Hydrophilic metabolite (%)	0 ± 0	0 ± 0	0 ± 0	32 ± 6	83 ± 2	94 ± 4	100 ± 0
	Unchanged tracer (%)	100 ± 0	100 ± 0	100 ± 0	68 ± 6	17 ± 2	6 ± 4	0 ± 0
Blocking	Hydrophilic metabolite (%)	0 ± 0	0 ± 0	3 ± 13	68 ± 21	96 ± 0	99 ± 3	100 ± 0
	Unchanged tracer (%)	100 ± 0	100 ± 0	97 ± 12	32 ± 15*	4 ± 1*	1 ± 2*	0 ± 0

**P* < 0.05 compared with baseline.

n = 3 per time point.

process by converting lipid-soluble substances into more hydrophilic compounds to be easily excreted by the kidneys (28). Hence, for hydrophobic compounds such as FPyKYNE-losartan, the hepatobiliary pathway is the main elimination route. The sex-averaged ED value calculated using ICRP 60 and 103 (0.031 mSv/MBq) is comparable to other ¹⁸F tracers such as 2-¹⁸F-fluoro-A-85380 (ED = 0.028 mSv/MBq), *O*-(2-¹⁸F-fluoroethyl)-L-tyrosine (ED = 0.016 mSv/MBq), and ¹⁸F-FDG (ED = 0.024 mSv/MBq) (29–31). EDs calculated for ¹⁸F-FPyKYNE-losartan are well below the regulatory limits recommended by the Food and Drug Administration (28). Although the RAS is known to differ between rats, pigs, and humans, the dose-limiting organs associated with common hepatobiliary metabolism and clearance pathways are likely to remain similar between species. Therefore, the dosimetry profile of ¹⁸F-FPyKYNE-losartan is within safe limits for approval of use in human studies.

PET imaging studies in pigs revealed higher image contrast and slower clearance of the tracer from the kidneys than in rats. The long residence time at the receptor and high tissue retention are expected with the long dissociation time of losartan in humans (67 min) (25,32,33). Reproducible results and imaging procedure provide evidence that the quantification of tracer uptake using SUVs is reliable in large animals. For accurate quantification, further arterial input corrections are required, namely metabolite correction for authentic parent tracer in plasma from arterial blood. Blockade of AT₁Rs with candesartan reduced peak retention (~ 60%) at 10–15 min in the renal cortex, further confirming binding specificity of ¹⁸F-FPyKYNE-losartan to AT₁Rs. Thereby, the PET signal in pigs is expected to represent unchanged ¹⁸F-FPyKYNE-losartan and NSB. Normal pig plasma exhibited faster ¹⁸F-FPyKYNE-losartan metabolism than rat plasma, because complete clearance from pig plasma was detected at 30 min whereas 23% of unmetabolized tracer was still present in rat plasma at that time (13). When AT₁Rs were blocked with candesartan, this process was even faster, with only 4% of the total radioactivity from unchanged tracer at 10 min and complete clearance of ¹⁸F-FPyKYNE-losartan from plasma at 20 min. The presence of hydrophilic metabolites only, not binding to AT₁Rs, should facilitate accurate blood input function corrections and kinetic modeling calculations for quantification of AT₁Rs in future studies. Confirmation of similar binding characteristics and high signal-to-noise ratio of the ¹⁸F-FPyKYNE structural analog of the clinically used losartan will facilitate translational work to humans.

CONCLUSION

¹⁸F-FPyKYNE-losartan binds with high affinity to renal AT₁Rs, and this novel radioligand displays full antagonism for Ang II pressor effect. The metabolism studies in small and large animals revealed that the labeled metabolites do not bind to renal AT₁Rs facilitating quantitative PET imaging. The sex-averaged ED of ¹⁸F-FPyKYNE-losartan is within an acceptable range compared with other ¹⁸F-labeled tracers. Reproducible PET images obtained in pigs combined with favorable binding profile support the potential of ¹⁸F-FPyKYNE-losartan for translational work in humans as an AT₁R PET imaging agent.

DISCLOSURE

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