First Clinical Evidence That Imaging with Somatostatin Receptor Antagonists Is Feasible

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Preclinical studies have indicated that somatostatin receptor (sst)-expressing tumors demonstrate higher uptake of radiolabeled sst antagonists than of sst agonists. In this study, we evaluated whether imaging with sst antagonists was feasible in patients. Methods: Biodistribution and tumor uptake of the sst antagonist ¹¹¹In-DOTA-pNO₂-Phe-c(DCys-Tyr-DTrp-Lys-Thr-Cys)DTyrNH₂ (¹¹¹In-DOTA-BASS) were studied in 5 patients with metastatic thyroid carcinoma or neuroendocrine tumors. Findings were compared with ¹¹¹In-pentetreotid (¹¹¹In-DTPA-octreotide) scan. Results: No adverse effects of ¹¹¹In-DOTA-BASS (20 μg) were observed. ¹¹¹In-DOTA-BASS detected 25 of 28 lesions, whereas ¹¹¹In-DTPA-octreotide detected only 17 of 28 lesions. In the same patient, ¹¹¹In-DOTA-BASS showed higher tumor and lower renal uptake than ¹¹¹In-DTPA-octreotide (3.5 \pm 2.8 percentage injected activity [%IA] vs. 1.0 \pm 0.99%IA and 1.5 \pm 0.3 %IA vs. 2.3 ± 0.7 %IA) at 4 h after injection. Conclusion: Imaging of neuroendocrine tumors with sst antagonists is clinically feasible. The favorable human biodistribution data suggest that sst antagonists could significantly affect peptide receptor-mediated imaging and therapy.

Key Words: somatostatin receptor targeting; neuroendocrine tumors; somatostatin receptor antagonists; antagonists

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 \mathbf{T} argeting of G-protein-coupled receptors with radiolabeled receptor agonists is a promising diagnostic and therapeutic approach in oncology (1). Radiolabeled somatostatin receptor (sst) agonists have become an integral part of diagnostic and therapeutic management of patients with neuroendocrine tumors (2,3). Although agonists readily internalize into tumor cells, permitting accumulation of radioactivity, radiolabeled antagonists do not and have therefore not been used clinically for tumor targeting.

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Ginj et al., however, showed in animal studies that radiolabeled sst antagonists are superior to agonists for the in vivo targeting of tumors expressing sst subtypes 2 (sst₂) and 3 (sst_3) (4). The in vivo uptake in sst_2 -positive tumors was twice as high with the antagonist ¹¹¹In-DOTA-pNO₂-Phe-c(DCys-Tyr-DTrp-Lys-Thr-Cys)DTyrNH₂ (¹¹¹In-DOTA-BASS) as with the highly potent agonist [¹¹¹In-DTPA⁰, Tyr³, Thr⁸]-octreotide (¹¹¹In-DTPA-TATE) (DTPA is diethylenetriaminepentaacetic acid). Scatchard analysis in human embryonic kidney (HEK)-sst₂ cells showed more than 10 times the number of binding sites for ¹¹¹In-DOTA-BASS (maximum number of sst₂ binding sites, 354 ± 14 pM) as for ¹¹¹In-DTPA-TATE (maximum number of sst₂ binding sites, 23 ± 1.0 pM) (4). The same observation has been made for 5-HT_{2A} receptors and corticotrophin-releasing factor receptors (5,6). Comparison of radiolabeled agonists and antagonists in a gastrin-releasing peptide receptor-positive prostate cancer animal model also showed the superiority of antagonists in an additional receptor system (7,8). If confirmed clinically, these observations could lead to a paradigm shift in the development of radiolabeled G-protein-coupled receptor ligands for diagnostic and therapeutic purposes.

The purpose of this pilot study was to evaluate the feasibility of imaging sst-expressing tumors in patients with the novel receptor antagonist ¹¹¹In-DOTA-BASS. Tumor detection rate and tracer uptake by tumors and normal organs were determined at various times and compared with recent ¹¹¹In-DTPA-octreotide scans. On the basis of the biodistribution of ¹¹¹In-DOTA-BASS, estimates were made for the expected radiation dosimetry of the therapeutic radioligand ⁹⁰Y-DOTA-BASS.

MATERIALS AND METHODS

Patients

Five consecutive patients with progressive disease after standard therapy were prospectively recruited. Their tumors demonstrated low or moderate uptake of ¹¹¹In-DTPA-octreotide (tumor uptake less than or slightly above liver uptake) limiting the efficacy of radionuclide therapy with sst agonists. Patient characteristics are summarized in Table 1. Exclusion criteria were renal insufficiency, treatment with short-acting somatostatin analogs less than 3 d, and sandostatin depot injection less than 4 wk before scanning. The use

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 TABLE 1

 Patient Characteristics

Patient no.	Age (y)	Sex	Tumor type	Previous treatment*
1	59	М	Follicular thyroid cancer	Isotretinoin redifferentiation therapy
2	66	М	Neuroendocrine tumor	⁹⁰ Y-DOTATOC therapy
3	60	М	Neuroendocrine tumor	Sandostatin therapy
4	56	М	Neuroendocrine tumor	Transabdominal chemoembolization
5	74	М	Neuroendocrine tumor	Hemihepatectomy

*111In-DTPA-octreotide and 111In-DOTA-BASS scans were performed more than 3 mo after termination of treatment.

of ¹¹¹In-DOTA-BASS in this setting was approved by the local institutional review board, and written informed consent was obtained in accordance with the Declaration of Helsinki.

Radiochemistry

The peptide-chelator conjugate DOTA-pNO₂-Phe-c(DCys-Tyr-DTrp-Lys-Thr-Cys)DTyrNH₂ (DOTA-BASS) (Fig. 1) was synthesized under good manufacturing practice conditions by standard 9-fluorenylmethoxycarbonyl solid-phase synthesis as previously described (9). The pure compound was characterized by electrospray ionization mass spectrometry and analytic reversed-phase high-performance liquid chromatography.

For the preparation of ¹¹¹In-DOTA-BASS, 500 μ L of ammonium acetate buffer (0.4 M, pH 5.0) were added to the vial containing 25 μ g of DOTA-BASS and incubated with approximately 200 MBq of ¹¹¹InCl₃ (Mallinckrodt) for 30 min at 95°C. Quality control of the final product was performed by analytic reversedphase high-performance liquid chromatography and instant thinlayer chromatography. ¹¹¹In-DTPA-octreotide was prepared with the commercially available kit Octreoscan (Covidien). The labeling yield of ¹¹¹In-DOTA-BASS and ¹¹¹In-DTPA-octreotide was 95% or greater at a specific activity of 12 GBq/µmol and 26 GBq/µmol, respectively. The radiochemical purity of ¹¹¹In-DOTA-BASS was greater than 95%.

Imaging

Total-body and SPECT ¹¹¹In-DOTA-BASS scans were obtained 11–13 d after ¹¹¹In-DTPA-octreotide scans. All studies were done with the same dual-head γ -camera (Skylight; Phillips) equipped with medium-energy, parallel-hole collimators. ¹¹¹In-DTPAoctreotide planar scans were obtained at 4 and 24 h after injection of approximately 10 μ g (167–192 MBq) of ¹¹¹In-DTPA-octreotide (European Association of Nuclear Medicine guidelines (10)), and sst antagonist total-body scans were obtained at 0.5, 1, 4, 24, 48, and 72 h after injection of 20 μ g (145–176 MBq) of ¹¹¹In-DOTA-BASS. For both tracers, SPECT images of the abdomen and thorax were acquired at 5 h after injection (2 × 180° rotation; 64 projections; 128 × 128 matrix; 30-s acquisition time per projection). Patients' vital parameters such as blood pressure, pulse rate, oxygen saturation, and electrocardiogram were monitored for at least 2 h after injection of ¹¹¹In-DOTA-BASS.

¹¹¹In-DOTA-BASS and ¹¹¹In-DTPA-octreotide scans were assessed independently by 2 experienced nuclear medicine physicians. The physicians were unaware of patients' identity, type of scan, and results of other imaging modalities. Afterward, the image findings were compared lesion by lesion with each other and with contrast-enhanced CT studies of the chest and abdomen. These scans had been acquired less than 5 wk before the ¹¹¹In-DTPA-octreotide scans using various multislice CT scanners.

Pharmacokinetics

For all imaging time points, regions of interest were manually drawn around the whole body and each organ with visible uptake (kidneys, spleen, liver, bladder, and tumor lesions), on both the anterior and the posterior scans. Background regions were placed close to the regions of interest for background correction. The geometric mean value, between anterior and posterior, was taken and corrected for attenuation using the conjugate-view method (*11*). Whole-body activity acquired 30 min after injection of ¹¹¹In-DOTA-BASS was set as 100 percentage injected activity (%IA).



FIGURE 1. Structural formula of sst antagonist DOTA-BASS.

¹¹¹In-DOTA-BASS



4 h p.i. 24 h p.i. 72 h p.i. ¹¹¹In-DTPA-OC



4 h p.i. 24 h p.i.

FIGURE 2. ¹¹¹In-DOTA-BASS (4, 24, and 72 h after injection) and ¹¹¹In-DTPA-octreotide (4 and 24 h after injection) planar posterior whole-body scans from patient 4. Large arrow shows increased uptake of ¹¹¹In-DOTA-BASS and ¹¹¹In-DTPA-octreotide in liver, corresponding to multiple liver metastases. Tumor-to-kidney uptake ratio was several times higher for ¹¹¹In-DOTA-BASS (6.1 at 4 h after injection and 3.3 at 24 h after injection) than for ¹¹¹In-DTPA-octreotide (1.18 at 4 h after injection and 0.88 at 24 h after injection). Small arrow shows spleen. OC = octreotide; p.i. = after injection.

Blood samples were taken at 1, 2, 3, 4, 5, 10, 20, 30, and 60 min and 1.5, 4, and 24 h after injection.

Dosimetry

Time–activity curves derived from the whole-body studies were analyzed with the OLINDA/EXM 1.0 software (Vanderbilt University) (*12*) to calculate the radiation dose after administration of ¹¹¹In-DOTA-BASS. For the latter calculation, it was assumed that ¹¹¹In-DOTA-BASS and ⁹⁰Y-DOTA-BASS demonstrate an identical biodistribution. Organ doses were calculated for the wholebody male phantom. Blood marrow doses were determined by the blood-based red marrow dose methodology, which assumes a linear relation between the blood residence time and the red marrow residence time (*13*).

RESULTS

Pharmacokinetics and Dosimetry

None of the patients experienced adverse reactions after intravenous injection of 20 µg of ¹¹¹In-DOTA-BASS. Blood sampling revealed a biexponential blood clearance of ¹¹¹In-DOTA-BASS: α -half-life, 12.5 ± 1.9 min and β -half-life, 6.1 ± 1.1 h; approximately 65% of the administered activity was cleared in the α -phase. The clearance occurred predominantly via the kidneys. The longest residence times of ¹¹¹In-DOTA-BASS were observed in the tumor and kidneys (Figs. 2 and 3).

¹¹¹ In-DOTA-BASS



¹¹¹ In-DTPA-OC



FIGURE 3. ¹¹¹In-DOTA-BASS (4, 24, and 72 h after injection) and ¹¹¹In-DTPA-octreotide (4 and 24 h after injection) planar anterior whole-body scans from patient 2. Images illustrate better tumor-to-background uptake ratio of ¹¹¹In-DOTA-BASS (e.g., tumor-to-liver uptake ratio of 6.7 at 4 h after injection and 3.5 at 24 h after injection) than of ¹¹¹In-DTPA-octreotide (1.2 at both time points). In this patient, ¹¹¹In-DOTA-BASS detected 16 metastases, whereas ¹¹¹In-DTPA-octreotide detected only 11 metastases. OC = octreotide; p.i. = after injection.



FIGURE 4. Comparison of ¹¹¹In-DOTA-BASS and ¹¹¹In-DTPA-octreotide uptake in kidneys, spleen, and tumors. Values are expressed as mean \pm SD of 5 patients. OC = octreotide; p.i. = after injection.

In the quantitative analysis, there were relevant differences in the biodistribution between the sst antagonist ¹¹¹In-DOTA-BASS and sst agonist ¹¹¹In-DTPA-octreotide (Fig. 4). The antagonist showed up to 4.1 times higher uptake in the tumor (¹¹¹In-DOTA-BASS, 3.5 ± 2.8 %IA, and ¹¹¹In-DTPA-octreotide, 1.0 ± 0.99 %IA) 4 h after injection. At the same time, the uptake in the kidneys was lower (¹¹¹In-DOTA-BASS, 1.5 ± 0.3 %IA, and ¹¹¹In-DTPA-octreotide, 2.3 ± 0.7 %IA), resulting in up to 5.2 times higher tumor-to-kidney uptake ratio in favor of the antagonist (Supplemental Table 1 [supplemental materials are available online only at http://jnm.snmjournals. org]; Figs. 2 and 4). The antagonist also showed lower uptake in organs such as liver (¹¹¹In-DOTA-BASS, 1.4 \pm 0.2 %IA, and ¹¹¹In-DTPA-octreotide, 1.7 \pm 0.6 %IA) and spleen (¹¹¹In-DOTA-BASS, 1.0 \pm 0.2 %IA, and ¹¹¹In-DTPA-octreotide, 2.3 \pm 1.0 %IA) 4 h after injection.

The calculated effective radiation dose was 0.049 \pm 0.0076 mSv/MBq for 111 In-DOTA-BASS and 0.25 \pm 0.031 mSv/MBq for 90 Y-DOTA-BASS (Table 2). The crit-

TABLE 2
Absorbed Radiation Dose of ¹¹¹ In-DOTA-BASS and ⁹⁰ Y-DOTA-BASS

	Wild et al.,	this study	Cremonesi et al. (<i>17</i>), ⁹⁰ Y-DOTATOC (mGy/MBq)
Organ/tissue	¹¹¹ In-DOTA-BASS (mGy/MBq)	⁹⁰ Y-DOTA-BASS (mGy/MBq)	
Adrenals	0.054 ± 0.0092	0.19 ± 0.074	NA
Brain	0.031 ± 0.0074	0.19 ± 0.074	NA
Gallbladder wall	0.053 ± 0.0072	0.19 ± 0.074	NA
Gastrointestinal tract			
Lower large intestine wall	0.049 ± 0.010	0.19 ± 0.074	NA
Small intestine	0.049 ± 0.010	0.19 ± 0.074	NA
Stomach wall	0.046 ± 0.010	0.19 ± 0.074	NA
Upper large intestine wall	0.048 ± 0.0098	0.19 ± 0.074	NA
Heart wall	0.043 ± 0.0090	0.19 ± 0.074	0.22 ± 0.12
Kidneys	0.18 ± 0.018	1.92 ± 0.21	3.31 ± 2.22
Liver	0.054 ± 0.0078	0.26 ± 0.098	0.72 ± 0.57
Lungs	0.038 ± 0.0078	0.19 ± 0.074	0.31 ± 0.18
Muscle	0.036 ± 0.0077	0.19 ± 0.074	NA
Pancreas	0.057 ± 0.011	0.21 ± 0.068	NA
Red marrow	0.041 ± 0.0076	0.18 ± 0.041	0.03 ± 0.01
Osteogenic cells	0.086 ± 0.019	0.30 ± 0.067	NA
Skin	0.024 ± 0.0054	0.16 ± 0.042	NA
Spleen	0.12 ± 0.034	1.01 ± 0.36	7.62 ± 6.30
Thymus	0.037 ± 0.0087	0.19 ± 0.074	NA
Thyroid	0.037 ± 0.0089	0.19 ± 0.074	NA
Urinary bladder wall	0.12 ± 0.028	0.98 ± 0.36	2.21 ± 0.31
Total body	0.039 ± 0.0079	0.22 ± 0.086	0.14 ± 0.06
Effective dose (mSv/MBq)	0.049 ± 0.0076	0.25 ± 0.031	NA

Data are mean \pm SD.

NA = not applicable.

Organ receiving highest dose was kidney. For therapeutic approach, it is likely that internalization and intracellular accumulation of short-range radionuclides (e.g., Auger emitters such as ¹¹¹In) is necessary because intracellular structures, for example, nucleus, are most radiosensitive part of cell. Because of cross-fire effect of medium- and long-range radionuclides (all β - and most α -emitters), internalization of such radionuclides is less likely to be relevant.

ical normal organs receiving the highest radiation dose were kidneys, spleen, and urinary bladder wall.

Detection of Tumor Lesions

¹¹¹In-DOTA-BASS, ¹¹¹In-DTPA-octreotide, and CT scans detected a total of 28 lesions; 25 were positive with ¹¹¹In-DOTA-BASS and 17 with ¹¹¹In-DTPA-octreotide (Supplemental Table 2). All lesions visible on ¹¹¹In-DTPA-octreotide scans were also detected by ¹¹¹In-DOTA-BASS, whereas there were 8 lesions on ¹¹¹In-DOTA-BASS that were not visible on ¹¹¹In-DTPA-octreotide. There were 3 bone lesions that were negative on both ¹¹¹In-DOTA-BASS and ¹¹¹In-DTPA-octreotide scans, but bone metastases were confirmed by serial CT.

DISCUSSION

Our pilot study provides the first clinical evidence that radiolabeled sst antagonists not only detect sst₂-expressing neuroendocrine tumors but also may even be superior to sst agonists for imaging and therapy of neuroendocrine tumors. The studied receptor antagonist ¹¹¹In-DOTA-BASS had a favorable biodistribution profile (higher tumor uptake and lower organ uptake) than the agonist ¹¹¹In-DTPA-octreotide, resulting in a higher tumor detection rate. The higher tumor uptake of ¹¹¹In-DOTA-BASS is consistent with preclinical studies that have indicated that ¹¹¹In-DOTA-BASS can bind to a larger number of sst₂ conformations than sst agonists (4). The slightly better receptor affinity of ¹¹¹In-DOTA-BASS than of ¹¹¹In-DTPA-octreotide (inhibitory concentration of 50%, 9.4 \pm 0.4 nM (4) vs. 22 \pm 3.6 nM (14)) may also have contributed to the higher tumor uptake of ¹¹¹In-DOTA-BASS.

With respect to radionuclide therapy, it is particularly encouraging that tumor-to-kidney ratios were up to 5.2 times higher for the antagonist than for the agonist. The kidneys are the major dose-limiting organ in peptide receptor radionuclide therapy, especially for ⁹⁰Y-labeled receptor agonists (3,15). On one hand, the severalfold higher tumor-to-kidney uptake ratio could significantly improve the efficacy and toxicity profile of radionuclide therapy with ⁹⁰Y-labeled sst ligands, especially in patients with moderate ¹¹¹In-DTPA-octreotide-avid tumor tissue (e.g., our patient collective). On the other hand, sst antagonists may have a negative effect on tumor growth. In vivo studies in rats showed a dose-dependent release of growth hormone growth hormone (GH) after injection of sst antagonists (16). The minimum effective dose necessary to achieve growth hormone release was 0.02 mg/kg (17.4 nmol/kg). The GH-releasing effect lasted about 15 min. It is therefore unlikely that 20-200 µg (14-140 nmol) of DOTA-BASS have any effects on tumor growth. To further evaluate the potential of somatostatin receptor antagonists for radionuclide therapy, we estimated the organ doses of ⁹⁰Y-DOTA-BASS. For these calculations, we assumed that ⁹⁰Y-DOTA-BASS shows the same biodistribution as ¹¹¹In-DOTA-BASS. ¹¹¹In has been used extensively as surrogate

for the pure β -emitter ⁹⁰Y because it has similar coordination chemistry, a similar physical half-life, and a similar metabolic handling (*1*,*17*). Compared with literature data for ⁹⁰Y-DOTATOC (*18*,*19*), ⁹⁰Y-DOTA-BASS showed lower renal, hepatic, and splenic radiation doses (Table 2).

The lower renal and liver doses can be explained by a charge difference between the 2 compounds. Negatively charged compounds such as ¹¹¹In-DTPA-octreotide show often higher renal and liver doses than neutral compounds such as ¹¹¹In- or ⁹⁰Y-DOTA-BASS (*20*). The reasons for the low splenic uptake of ¹¹¹In-DOTA-BASS are currently not clear.

The following limitations of the study should be noted. We have chosen the only Food and Drug Administration– and European Medicines Agency–approved sst imaging agent (¹¹¹In-DTPA-octreotide) for comparison with our sst₂ antagonist ¹¹¹In-DOTA-BASS. Comparison with unlicensed second-generation sst analogs—for example, ¹¹¹In-DOTATOC—was not possible because of legal reasons. However, choosing ¹¹¹In-DTPA-octreotide is feasible because ¹¹¹In-DTPA-octreotide and ¹¹¹In-DOTATOC show similar biodistribution in humans, without significantly different tumor and kidney uptake (*21*).

Only 5 patients were included in this pilot study because the aim of the study was to obtain first clinical data on the potential usefulness of imaging sst-positive tumors with sst antagonists and not to determine the sensitivity and specificity of ¹¹¹In-DOTA-BASS SPECT in a specific population.

CONCLUSION

This is the first proof of concept showing that imaging of neuroendocrine tumors with sst antagonists is feasible in patients. As suggested by preclinical studies, ¹¹¹In-DOTA-BASS provided a higher tumor uptake and better visualization of metastatic neuroendocrine tumors than ¹¹¹In-DTPA-octreo-tide. We anticipate that the antagonist approach will have significant impact on peptide receptor–mediated imaging and therapy.

DISCLOSURE STATEMENT

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