

# Monitoring and Predicting Response to Therapy with $^{18}\text{F}$ -FDG PET in Colorectal Cancer: A Systematic Review

Lioe-Fee de Geus-Oei<sup>1</sup>, Dennis Vriens<sup>1</sup>, Hanneke W.M. van Laarhoven<sup>2</sup>, Winette T.A. van der Graaf<sup>2</sup>, and Wim J.G. Oyen<sup>1</sup>

<sup>1</sup>Department of Nuclear Medicine, Radboud University, Nijmegen Medical Centre, Nijmegen, The Netherlands; and

<sup>2</sup>Department of Medical Oncology, Radboud University, Nijmegen Medical Centre, Nijmegen, The Netherlands

Molecular imaging with  $^{18}\text{F}$ -FDG PET has been proven useful in the management of colorectal cancer.  $^{18}\text{F}$ -FDG PET plays a pivotal role in staging before surgical resection of recurrent colorectal cancer and metastases, in the localization of recurrence in patients with an unexplained rise in serum carcinoembryonic antigen levels, and in the assessment of residual masses after treatment. Currently, there is increasing interest in the role of  $^{18}\text{F}$ -FDG PET beyond staging. The technique appears to have significant potential for the characterization of tumors and for the prediction of prognosis in the context of treatment stratification and early assessment of tumor response to therapy. This systematic review provides an overview of the literature on the value of  $^{18}\text{F}$ -FDG PET for monitoring and predicting the response to therapy in colorectal cancer. The review covers chemotherapy response monitoring in advanced colorectal cancer, monitoring of the effects of local ablative therapies, and preoperative radiotherapy and multimodality treatment response evaluation in primary rectal cancer. Given the added value of  $^{18}\text{F}$ -FDG PET for these indications, implementation in clinical practice and systematic inclusion in therapeutic trials to exploit the potential of  $^{18}\text{F}$ -FDG PET are warranted.

**Key Words:**  $^{18}\text{F}$ -FDG PET; colorectal carcinoma; prediction; response monitoring; therapy monitoring

**J Nucl Med 2009; 50:43S–54S**

DOI: 10.2967/jnumed.108.057224

Colorectal cancer was the third most common cancer in men and women in 2008 and is the third leading cause of cancer-related deaths in the United States. Although colorectal cancer incidence rates decreased from 1998 through 2004, this cancer continues to be a major health problem worldwide. About 148,800 cases of colorectal cancer are expected to be newly diagnosed in the United States in 2008, the cumulative lifetime risk is approxi-

mately 5%, and the current 5-y survival rate approaches 66% (1). The prognosis for patients with this disease has improved substantially, mainly because of earlier detection and the introduction of effective systemic (chemo)therapeutic agents (2–4).

Molecular imaging with  $^{18}\text{F}$ -FDG PET has been shown to be useful in the management of colorectal cancer.  $^{18}\text{F}$ -FDG PET already plays a pivotal role in staging before surgical resection of locally recurrent cancer and metastases, in the localization of recurrence in patients with an unexplained rise in serum carcinoembryonic antigen levels, and in the assessment of residual masses after treatment. This diagnostic tool seems to be very promising for therapy stratification as well (5,6).

Currently, there is growing interest in the role of  $^{18}\text{F}$ -FDG PET beyond staging, in particular, for the prediction of tumor response to therapy. Hence, the number of clinical applications for  $^{18}\text{F}$ -FDG PET in colorectal cancer continues to increase. This systematic review discusses the emerging role of  $^{18}\text{F}$ -FDG PET in the prediction and evaluation of responses to treatment, such as monitoring chemotherapy responses in advanced colorectal cancer, monitoring responses after local ablative therapy of liver metastases, and monitoring radiotherapy and multimodality treatment responses in primary rectal cancer.

## SEARCH STRATEGY AND LITERATURE SELECTION CRITERIA

Data for this review were identified by searches of PubMed, MEDLINE (OvidSP), EMBASE (OvidSP), and the Cochrane Library up to December 2008 with the search terms reported by Mijnhout et al. (7) for identifying clinical  $^{18}\text{F}$ -FDG PET studies and the following 2 groups of search terms: “colorectal carcinoma” or “colorectal cancer” or “colon cancer” or “rectal cancer” or “rectal carcinoma” and “prediction” or “therapy monitoring” or “response monitoring” or “response.” Search results were evaluated for adequacy. Only articles in English were included. We omitted articles that merely dealt with staging before surgical resection of recurrent cancer and metastases,

Received Nov. 4, 2008; revision accepted Jan. 28, 2009.

For correspondence or reprints contact: Lioe-Fee de Geus-Oei, Department of Nuclear Medicine, Internal Postal Code 444, Radboud University, Nijmegen Medical Centre, P.O. Box 9101, 6500 HB Nijmegen, The Netherlands.

E-mail: L.deGeus-Oei@nuccmed.umcn.nl

COPYRIGHT © 2009 by the Society of Nuclear Medicine, Inc.

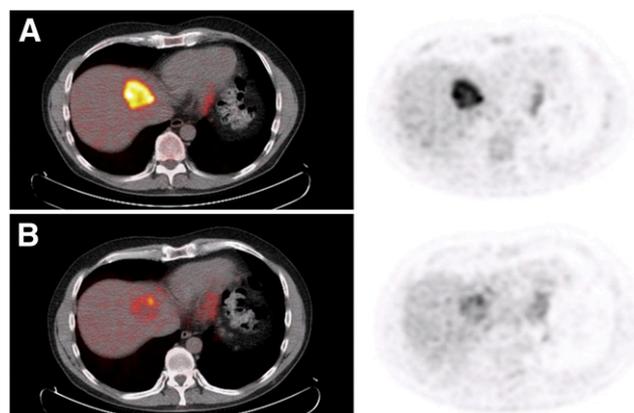
localization of recurrence in patients with an unexplained rise in serum carcinoembryonic antigen levels, and residual masses after treatment; with malignancies other than colorectal carcinoma or liver metastases of mixed primary origin; and with radiopharmaceuticals other than  $^{18}\text{F}$ -FDG. Descriptions of treatment responses without fixed outcome parameters (e.g., histologic or morphologic response, overall survival, or progression-free survival) were also omitted. References from included articles were checked for studies not retrieved by the search strategy. Abstracts, reports from meetings, editorial comments, and letters to the editor were excluded.

#### CHEMOTHERAPY RESPONSE MONITORING IN ADVANCED COLORECTAL CANCER

Tumor response to therapy is traditionally assessed by comparison of tumor sizes determined by morphologic imaging methods (CT) before and after treatment. According to the response evaluation criteria in solid tumors (RECIST), the current definition of tumor response is a decrease in the sum of the largest diameters of tumor lesions of at least 30% (8). The RECIST are widely accepted, but the correlation between morphologic tumor response and patient outcome is rather weak (9). Moreover, residual benign masses may persist, despite the fact that disease activity may have completely resolved after successful therapy.

$^{18}\text{F}$ -FDG PET yields data independent of associated structural characteristics and therefore allows the detection or monitoring of specific metabolic changes that are not associated with or that precede therapy-induced anatomic changes. The strength of  $^{18}\text{F}$ -FDG PET is that it permits whole-body imaging in a noninvasive way. In contrast to histopathologic analysis of biopsy material,  $^{18}\text{F}$ -FDG PET is not limited to the characterization of one or a few (sometimes very heterogeneous) target lesions; it can evaluate multiple tumor sites at the same time. Furthermore, serial scanning can be performed, allowing the measurement of functional changes over time during therapeutic interventions. PET not only can visualize (Fig. 1) but also can quantify  $^{18}\text{F}$ -FDG uptake and is able to provide several highly reproducible quantitative parameters of tumor glucose metabolism. However, the disadvantages of molecular imaging techniques (limited resolution and radiation burden) relative to techniques such as MRI should not be ignored. Therefore, these techniques should not be considered competitive but rather should be considered complementary because they aim to visualize and measure different processes in the human body.

However, it is obvious that the introduction of targeted therapies, such as the angiogenesis inhibitor bevacizumab, requires new tools for monitoring therapeutic effects, because these agents exhibit a cytostatic effect rather than the cytoreductive effect of classic chemotherapy. These new agents inhibit the growth of new blood vessels in cancer tissue; this effect does not immediately lead to a



**FIGURE 1.** Typical example of colorectal cancer patient with nonresectable liver metastases that responded to chemotherapy. Relative to baseline (A), 85% decrease in  $\text{MR}_{\text{glu}}$  was seen on  $^{18}\text{F}$ -FDG PET after 2 mo of chemotherapy (B). Transversal fused PET/CT scans are shown on left; transversal PET scans are shown on right. (Courtesy of Dr. W.V. Vogel.)

decrease in tumor size and thus places new demands on imaging modalities. When anticancer treatment becomes more individualized, it is increasingly important to identify a response at earlier time points. Early identification of the lack of a response would prevent side effects, reduce the costs of futile treatment, and prevent delays in instituting a second-line, potentially effective, therapy.

Five studies (Table 1) reported the predictive value of  $^{18}\text{F}$ -FDG PET in patients treated with chemotherapy for nonresectable colorectal cancer liver metastases (10–14). As early as 1996, Findlay et al. (10) studied 18 patients before, at 1–2 wk after, and at 4–5 wk after 5-fluorouracil (5-FU) chemotherapy with or without interferon- $\alpha$  by using a nondedicated PET system with a limited resolution. Responding lesions showed a significantly greater reduction in  $^{18}\text{F}$ -FDG uptake, compared with the baseline value, than nonresponding lesions (–33% vs. –1%;  $P < 0.001$ ). The 4- to 5-wk tumor-to-liver ratio was able to discriminate responders from nonresponders in both a lesion-by-lesion assessment and an overall patient response assessment with a sensitivity of 100% and specificities of 90% and 75%, respectively. A clear correlation was observed between the reduction in tumor glucose metabolism at 5 wk after the start of chemotherapy and treatment outcome; such a correlation was not observed at 1–2 wk after the start of treatment. These results showed the importance of the correct timing of  $^{18}\text{F}$ -FDG PET after the beginning of chemotherapy. The authors mentioned the so-called flare phenomenon that occurs at 1–2 wk after the initiation of chemotherapy and that can be observed as a marked increase in  $^{18}\text{F}$ -FDG metabolism in lesions that show a response later on.

Bender et al. (11) showed that the flare phenomenon probably does not play a role as early as 72 h after the initiation of chemotherapy. Their preliminary results demonstrated that therapy-sensitive metastases show a reduc-

**TABLE 1.** Chemotherapy Response Monitoring in Advanced Colorectal Cancer

Authors	Year	Reference	No. of patients	Therapy	Timing of PET evaluation	PET response criteria	Outcome measures	Results	P	
Findlay et al.	1996	10	18	5-FU chemotherapy	1–2 wk, 4–5 wk	– $\Delta T:L > 15\%$ (at 4–5 wk)	Morphologic response on CT scan at 12 wk (WHO criteria)	Sensitivity 100%; specificity 75%		
Bender et al.	1999	11	6	5-FU+FA chemotherapy	72 h	– $\Delta SUV$	Morphologic response on CT scan at 6 wk (WHO criteria)	Responders –22%; nonresponders +13%	<0.01	
Dimitrakopoulou-Strauss et al.	2003	12	28	FOLFOX chemotherapy	2 wk, 3 mo	SUV, FD	Morphologic response on CT scan at 12 wk (WHO criteria)	Correct classification rate: FD baseline PET 90% for PD; FD baseline PET 75% for SD		
Dimitrakopoulou-Strauss et al.	2004	13	25	FOLFOX chemotherapy	2 wk, 8 wk	SUV, $k_1-k_4$ , OS, FD, VB		Correct classification rate: SUV 62% at 2 wk and 69% at 8 wk; $k_1-k_4$ , FD, and VB 78% at 2 and 8 wk	OS–SUV correlation at 2 wk 0.426 OS–SUV correlation at 8 wk 0.517	0.035 0.001
de Geus-Oei et al.	2007	14	50	Various chemotherapy schedules	2 mo, 6 mo	– $\Delta SUV$ , – $\Delta MR_{glu}$	OS  Progression-free survival	– $\Delta SUV$ at 2 mo  – $\Delta MR_{glu}$ at 2 mo – $\Delta SUV$ at 2 mo – $\Delta MR_{glu}$ at 2 mo	0.017  0.049 0.035 0.026	

$\Delta T:L$  = fractional change in tumor-to-liver ratio; FA = folinic acid;  $\Delta SUV$  = fractional change in SUV; FD = fractal dimension; PD = progressive disease; SD = stable disease;  $k_1-k_4$  = rate constants; VB = vascular fraction; OS = overall survival;  $\Delta MR_{glu}$  = fractional change in  $MR_{glu}$ .

tion in  $^{18}F$ -FDG uptake after a single application of chemotherapy and that this effect can be quantified by PET and seems to be indicative of the final therapy outcome after the completion of an anticipated therapy cycle.

Dimitrakopoulou-Strauss et al. (12) studied 28 patients who were treated with second-line 5-FU–folinic acid–oxaliplatin (FOLFOX) therapy. Reference standards for the serial  $^{18}F$ -FDG PET studies were the clinical response

data, according to the World Health Organization (WHO) classification. In contrast to the studies of Findlay et al. (10) and Bender et al. (11), that study investigated absolute standardized uptake values (SUVs) and kinetic parameters and not fractional changes between baseline and follow-up scans. Even the quantitative values from the first PET study (at baseline) were predictive with respect to therapy outcome. The so-called fractal dimension, a kinetic parameter

that describes the heterogeneity of tissue time–activity data in tumors, produced the best results and correctly classified progressive disease and stable disease in 90% and 75% of cases at baseline, respectively.

In a similar study, Dimitrakopoulou-Strauss et al. (13) examined the potential of serial  $^{18}\text{F}$ -FDG PET to predict the response to chemotherapy as reflected by individual survival times. It was shown that a combination of kinetic parameters from the first scan (at baseline) and the third scan (at 8 wk) provided the best results for classification into short- and long-term survival classes (defined as survival of <1 y and >1 y, respectively). The authors suggested that quantitative dynamic  $^{18}\text{F}$ -FDG PET should be used preferentially for chemotherapy response monitoring.

A recent study (14) showed that  $^{18}\text{F}$ -FDG PET could be readily implemented without the need for complex dynamic imaging protocols. It was demonstrated that simplified measures (e.g., SUVs) can replace more complex quantitative measures (e.g., rate of metabolism of glucose [ $\text{MR}_{\text{glu}}$ ]). Dynamic  $^{18}\text{F}$ -FDG PET was performed before and at 2 mo ( $n = 50$ ) and 6 mo ( $n = 19$ ) after the start of treatment. There were increases in the rates of progression and death associated with the worst response as assessed by PET, as determined by Cox proportional regression analysis. The overall and progression-free survival data yielded significant predictive values at broad cutoff levels for changes in  $\text{MR}_{\text{glu}}$  and SUV. It was concluded that the degree of chemotherapy-induced changes in tumor glucose metabolism in advanced colorectal cancer is highly predictive of patient outcome.

A noncomplex approach is advantageous over a full kinetic analysis approach because it will facilitate broad introduction into clinical practice and will improve patient compliance, which is an important feature of successful clinical trials. Another advantage of SUVs is that they can be calculated from static whole-body  $^{18}\text{F}$ -FDG PET studies, which depict all metastases. In dynamic scans, only one axial field of view, typically 15–20 cm, can be studied during dynamic data acquisition. Because metastatic lesions in different parts of the body may respond differently to chemotherapy, this feature represents a principal advantage of SUV analysis over kinetic analysis.

An important observation is that chemotherapy-induced normalization of  $^{18}\text{F}$ -FDG uptake in liver metastases of colorectal cancer does not indicate a complete pathologic response (15–17). Tan et al. (16) found that, despite the absence of detectable metabolic activity above the background, viable tumor cells could still be found in 85% of lesions. In a subgroup of 7 lesions in which neoadjuvant chemotherapy resulted in both a complete metabolic response and a complete CT response (RECIST), histologic analysis revealed viable tumor cells in 6 lesions. A reduction in the number of viable tumor cells below the limit of detection may be an important reason why lesions are not seen by  $^{18}\text{F}$ -FDG PET after treatment. In addition, the relatively high level of  $^{18}\text{F}$ -FDG uptake in normal hepatic

parenchyma makes it more difficult to detect lesions with a partial metabolic response resulting in uptake only slightly greater than that of the liver. Another factor may be the effect of chemotherapy on tumor cell  $^{18}\text{F}$ -FDG uptake. Chemotherapeutic agents may reduce  $^{18}\text{F}$ -FDG uptake by altering the glucose metabolism of tumor cells. Akhurst et al. (18) postulated that this change in  $^{18}\text{F}$ -FDG uptake after cytotoxic therapy is induced by a decrease in the activity of the key glycolytic enzyme hexokinase. Therefore, curative resection of liver metastases that demonstrate a complete metabolic response on  $^{18}\text{F}$ -FDG PET should not be omitted.

#### MONITORING RESPONSES AFTER LOCAL ABLATIVE THERAPY OF LIVER METASTASES

Complete surgical resection offers the best chance for cure in patients with colorectal liver metastases. However, because of the number or localization of liver metastases, complete resection with adequate tumor-free margins and adequate liver function reserve cannot be achieved in all patients. Local ablative techniques that result in intrahepatic tumor destruction have emerged as alternative treatment options, although positive effects on patients' survival remain to be established. These techniques include microwave tumor coagulation (19), laser-induced thermotherapy (20), injection of ethyl alcohol (21), cryosurgical ablation or cryotherapy (CSA) (22), and radiofrequency ablation (RFA) (23).

Different morphologic imaging techniques have been used to facilitate intraoperative localization. However, during the process of local ablation, the destruction process cannot easily be ascertained with intraoperative ultrasonography because of the hyperechogenicity that is induced within the treated area (24). Furthermore, evaluation with CT or MRI of residual tumor after the ablation procedure is hampered by a rimlike increase in contrast enhancement that occurs immediately after RFA and that resembles peripheral hyperperfusion. This area of contrast enhancement may interfere with the adequate detection of residual tumor (25). This problem can lead to either a delayed diagnosis of treatment failure or confusion between incomplete local ablative treatment and the occurrence of new metastases in regions adjacent to the treatment site.

Several studies (Table 2) have described the feasibility of  $^{18}\text{F}$ -FDG PET (Fig. 2) for the surveillance of patients with liver metastases (26–30).  $^{18}\text{F}$ -FDG PET appears to have great potential for identifying residual tumor very early after local ablative treatments. Earlier detection offers the opportunity to treat tumors again at an early stage, by either surgery or repeated local ablation in the case of an insufficient initial treatment result. A study performed at our own institute by Langenhoff et al. (26) demonstrated that  $^{18}\text{F}$ -FDG PET performed early after local tumor ablation provided additional information about efficacy by differentiating posttreatment changes from residual or recurrent malignant tumor.  $^{18}\text{F}$ -FDG PET results became negative in 51 lesions within 3 wk after local ablative therapy, meaning

**TABLE 2. Response Monitoring After Local Ablative Therapy for Treatment of Colorectal Cancer Liver Metastases**

Authors	Year	Reference	No. of patients	Therapy	Timing of PET evaluation	PET response criteria	Outcome measures	Results
Langenhoff et al.	2002	26	22	CSA and RFA	<3 wk	Negative results	Recurrence	NPV 100%; PPV 80%
Donckier et al.	2003	28	17	RFA	1 wk, 4 wk	Negative results	Residual tumor	NPV 100%; PPV 100%
Joosten et al.	2005	27	43	CSA and RFA	<3 wk	Negative results	Recurrence	NPV 97%; PPV 88%
Veit et al.	2006	30	11	RFA	<2 d	Negative results	Recurrence	Accuracy 68%
Denecke et al.	2007	20	21	LITT	1–3 d, 1–6 mo, >6 mo	Negative results	Residual tumor or recurrence	NPV 96%; PPV 97%

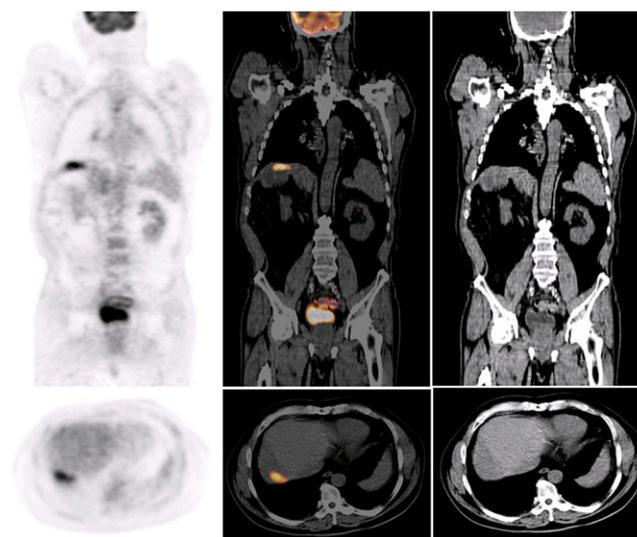
CSA = cryosurgical ablation; RFA = radiofrequency ablation; NPV = negative predictive value; PPV = positive predictive value; LITT = laser-induced thermotherapy.

that  $^{18}\text{F}$ -FDG-accumulating liver metastases became photopenic. The  $^{18}\text{F}$ -FDG PET results for 5 lesions remained positive. A local recurrence was identified on CT during a mean follow-up of 16 mo for 4 of these lesions; the other lesion was found to be an abscess. Conversely, a local recurrence was not identified for lesions without  $^{18}\text{F}$ -FDG uptake on PET (negative predictive value, 100%). In all patients,  $^{18}\text{F}$ -FDG PET detected recurrence considerably earlier than CT (3.8 vs. 8.5 mo).

Donckier et al. (28) performed  $^{18}\text{F}$ -FDG PET at 1 and 4 wk after local ablative treatment. They showed that  $^{18}\text{F}$ -FDG PET was accurate in monitoring the local effect of RFA because it recognized incomplete tumor ablation early, even when incomplete tumor ablation was not detectable by CT. After a median follow-up of 11 mo, none of 24 lesions without  $^{18}\text{F}$ -FDG uptake on PET after ablation showed the development of a local recurrence. In 4 patients,  $^{18}\text{F}$ -FDG PET at 1 wk and 1 mo revealed a peripheral hypermetabolic residue after RFA, whereas CT did not reveal residual tumor. In 3 patients, a local persistence of viable tumor cells was biopsy proven at reintervention. In the fourth patient, follow-up CT revealed the subsequent development of a local recurrence.

Joosten et al. (27) showed that CT after treatment did not predict local treatment failure, whereas  $^{18}\text{F}$ -FDG PET within 3 wk after local ablative treatment predicted 6 of 7 local recurrences in a study population of 43 patients with 104 ablated lesions. One local recurrence was detected on  $^{18}\text{F}$ -FDG PET 3 mo after treatment. The negative predictive value of  $^{18}\text{F}$ -FDG PET at 3 mo was 100%. Because  $^{18}\text{F}$ -FDG PET revealed 1 false-positive result because of focal infection, the positive predictive value was 88%. These results are in line with the findings of Denecke et al. (20). Among 54 lesions, they found 1 false-positive lesion 54 d after treatment, presumably caused by regenerative processes. One false-negative lesion among 6 examined was found within 3 d after treatment, probably because of a small volume of surviving tumor tissue.

Similar observations have been reported by Veit et al. (30). Their small study ( $n = 11$ ) was the first to use integrated PET/CT. Four of 6 patients with residual tumor showed residual  $^{18}\text{F}$ -FDG accumulation at the ablative margin as early as 2 d after RFA. The accuracy for the detection of residual tumor directly after RFA was higher for PET/CT than for CT alone (68% vs. 47%). Although the number of patients was too small to draw definite conclusions, the authors pointed out some advantages of a short imaging interval after RFA and of the use of PET/CT for this indication over the use of PET or CT alone. They postulated that dual-modality PET/CT simplifies guidance for reinterventions and that a follow-up scan directly after



**FIGURE 2.**  $^{18}\text{F}$ -FDG PET/CT 3 mo after radiofrequency ablation. Clear  $^{18}\text{F}$ -FDG accumulation revealed residual vital tumor tissue. Abnormalities on CT after 3 and 6 mo were interpreted as posttreatment effects, and tumor recurrence was diagnosed on CT after 9 mo. Upper row: coronal images; lower row: transversal images. From left to right: PET scans, fused PET/CT scans, and CT scans.

RFA is ideal to shorten the time to a possible reintervention. The authors speculated that a follow-up scan should take place within 2 d after RFA, before tissue regeneration takes place (30). Tissue regeneration might cause rimlike tracer distribution at the ablative margin; in contrast, viable tumor residue results in an area of focally increased  $^{18}\text{F}$ -FDG uptake.

### PREOPERATIVE RADIOTHERAPY AND MULTIMODALITY TREATMENT RESPONSE EVALUATION IN PRIMARY RECTAL CANCER

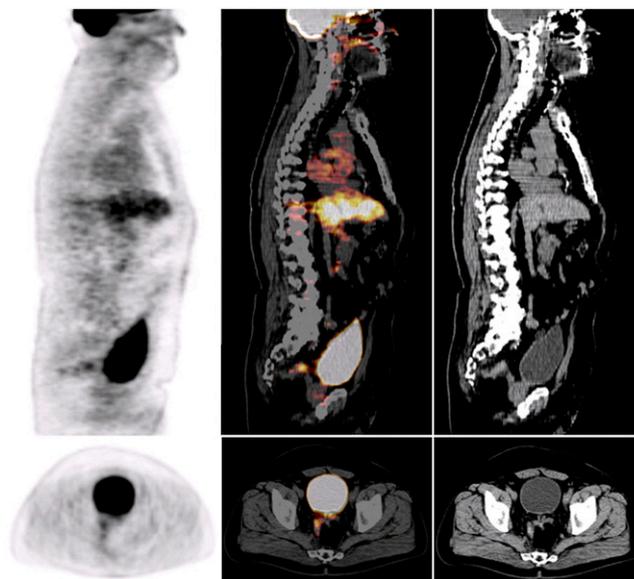
With the exception of very early tumors that can be managed by local excision alone, the mainstay of therapy for rectal cancer is radical surgery. Total excision of the mesorectum has emerged as the surgical technique that can substantially reduce local recurrences. However, the risk of distant and local recurrences continues to threaten patients with rectal cancer. Patients with locally advanced rectal cancer are at the highest risk of failure of local treatment. Therefore, surgery alone is often not curative, and preoperative radiotherapy is required to achieve a radical resection and improve the local control rate (31). For locally advanced rectal cancer, neoadjuvant chemoradiation has been proven successful (32,33). Tumor responses to chemoradiotherapy, however, vary considerably. Some patients experience serious side effects, and not all patients benefit equally (34). Therefore, the surgical approach largely depends on a valid assessment of the preoperative extent of the tumor, particularly for distally located tumors or tumors that have been assessed as being nonresectable during primary staging.

The current standard method for discriminating responders from nonresponders is conventional histopathologic analysis, measuring the extent of the residual tumor. This method, however, is applicable only in a postoperative setting and consequently cannot be used for the preoperative selection of individualized surgery. First, accurate restaging to assess the success of neoadjuvant chemoradiation is critical because it can guide optimization of the surgical approach, such as sphincter-saving surgery in deep-seated tumors, less aggressive resection in minimally advanced tumors, or planning of intraoperative radiation therapy. Second, the correct assessment of responses and the identification of nonresponders may have clinically relevant consequences during the course of chemoradiation. These include the potential for individualized treatment, for instance, the escalation of preoperative treatment (e.g., increasing the radiation dose, adding regional hyperthermia, or using intraoperative radiation) or the addition of chemotherapeutic regimens after tumor resection. Therefore, there is an obvious need for reliable noninvasive methods suitable for the prediction of responses, especially complete pathologic remission.

In the setting of routine clinical practice, mainly anatomically based imaging modalities, such as CT, MRI, and endorectal ultrasound, have been used. Two recent meta-

analyses showed that these 3 imaging modalities are highly accurate in the staging of untreated tumors because of their ability to detect invasion in the perirectal fat or adjacent organ involvement (35,36). The situation is dramatically different when these imaging modalities are considered for the purpose of restaging after chemoradiotherapy, because their accuracies range from only 30% to 60%—accuracies that are clearly too low to support decisions regarding changes in the surgical approach (35–38). Conventional imaging modalities cannot distinguish fibrosis or scar from viable tumor cells in residual masses after chemoradiotherapy; therefore, these methods have a negligible impact on the prediction of pathologic findings (38,39). As a result, great demands are placed on imaging modalities that provide a combination of functional information ( $^{18}\text{F}$ -FDG PET) and morphologic information (CT, MRI, and endorectal ultrasound) (Fig. 3). The introduction of integrated PET/CT scanners has shed new light on this issue.

The first study (Table 3) that reported on metabolic response assessment after irradiation of nonresectable presacral recurrent rectal carcinomas was published in 1992 by Engenhart et al. (40) They noted a small but significant ( $P = 0.002$ ) decrease in  $^{18}\text{F}$ -FDG uptake during treatment but concluded that it was still too early to use  $^{18}\text{F}$ -FDG PET for radiotherapy treatment monitoring because the effects of proliferation, repair, inflammation, and residual viable tumor cells on glycolytic activity had to be determined first. Several years later, Schiepers et al. (41) investigated the



**FIGURE 3.**  $^{18}\text{F}$ -FDG PET/CT of nonresponding patient with locally advanced rectal cancer that was treated with preoperative chemoradiation. In this preoperative study, tumor uptake was still present, and histopathologic analysis confirmed residual vital tumor tissue. Upper row: sagittal images; lower row: transversal images. From left to right: PET scans, fused PET/CT scans, and CT scans.

influence of induction radiotherapy on tumor biology in patients who had primary rectal cancer and were scheduled for elective surgery. A group treated with surgery alone was compared with a group treated with surgery after preoperative radiotherapy. The relationship between glucose use and cell kinetics was investigated. A 65% decrease in glucose use measured 2–3 wk after radiotherapy correlated with a reduction in tumor cell burden and cell death. The authors concluded that it was possible to discriminate successfully from unsuccessfully treated tumors as early as 2 wk after radiotherapy with an accuracy of 80%.

Other studies, however, warned about confounding radiotherapy-induced effects on  $^{18}\text{F}$ -FDG uptake. On the one hand, they can be caused by the interference of inflammatory cells because approximately 25% of  $^{18}\text{F}$ -FDG uptake can occur in nontumor tissues, such as macrophages, neutrophils, fibroblasts, and granulation tissue (42,43). On the other hand, they can be caused by a short-lived reversible decrease in glucose metabolism attributable to the so-called stunning of tumor cells (44). The second phenomenon can mimic actual cytotoxic therapy effects, although only temporarily. These effects led Haberkorn et al. (45) to recommend that  $^{18}\text{F}$ -FDG PET studies should be postponed for 60 d after radiotherapy. Such a long interval, however, is not clinically feasible in a neoadjuvant setting. After the study of Haberkorn et al. (45), only a few studies of radiotherapy response monitoring were performed (40,41,46–48). Siegel et al. (47) studied metabolic responses to a short course of radiotherapy (Table 3). A significant reduction (40%) in SUVs was found as early as days 7–8 after the start of the short course of radiotherapy. At this short interval, however, no correlation was seen between the reduction in SUVs and the tumor regression grade. The results of most of the studies (41,46–48) implied a true radiation-induced reduction in glucose use because of tumor cell loss. These results call for systematic investigations of the required interval for radiotherapy response evaluation with  $^{18}\text{F}$ -FDG PET.

As shown in Table 3, 15 studies of multimodality treatment (chemoradiation with or without regional hyperthermia) response monitoring were performed (44,47,49–61). Several of these studies (44,50–52,58) compared metabolic and morphologic response evaluations. It was demonstrated that the reduction in SUVs was significantly greater in (histopathologically confirmed) responders than in nonresponders.  $^{18}\text{F}$ -FDG PET predicted therapy outcomes significantly better than endorectal ultrasound, CT, and MRI (44,50–52). In the study of Amthauer et al. (44),  $^{18}\text{F}$ -FDG PET had a sensitivity of 100% and a specificity of 86% when a minimum posttherapeutic SUV reduction of 36% was used to define a response. The positive and negative predictive values were 93% and 100%, respectively. Calvo et al. (50) showed that T downstaging was significantly correlated with absolute changes in the maximum SUV ( $\text{SUV}_{\text{max}}$ ) of  $-3.3$  and  $-1.9$  for responders and nonresponders, respectively ( $P = 0.03$ ). Guillem et al. (49)

introduced the metabolic response parameter range in total lesion glycolysis ( $\delta\text{TLG}$ ), an important variable that subsequently was also analyzed by Melton et al. (58). This parameter not only incorporates the difference in tumor activity between baseline (pre) and posttherapy (post) scans but also takes into account changes in tumor size (volume), as follows:  $\delta\text{TLG} = \{[(\text{SUV}_{\text{post}} \times \text{volume}_{\text{post}}) - (\text{SUV}_{\text{pre}} \times \text{volume}_{\text{pre}})] / (\text{SUV}_{\text{pre}} \times \text{volume}_{\text{pre}})\} \times 100\%$ .

In a subsequent study, Guillem et al. (51) demonstrated (probably in the same patient population) that  $^{18}\text{F}$ -FDG PET was able to predict long-term clinical outcomes (51). The fractional change in tumor glucose use at 4–5 wk after the completion of chemoradiation was found to be the best predictor of recurrence-free survival. The mean percentage decreases in the  $\text{SUV}_{\text{max}}$  after a median follow-up of 42 mo were 69% for patients without recurrence and 37% for patients with recurrence ( $P = 0.004$ ). Patients with a decrease in the  $\text{SUV}_{\text{max}}$  of  $\geq 62.5\%$  and a  $\delta\text{TLG}$  of  $\geq 69.5\%$  had significantly improved recurrence-free survival ( $P = 0.02$  and  $P = 0.01$ , respectively), and patients with a  $\delta\text{TLG}$  of  $\geq 69.5\%$  also had significantly longer overall survival ( $P = 0.03$ ). The pathologic response in this group of patients surprisingly was not a significant predictor of overall survival or recurrence-free survival. In that study, a minimal metabolic response of the primary tumor was shown to reflect unfavorable tumor biology, as evidenced by a predilection for distant metastatic disease, which resulted in poor overall survival. The metabolic response of a primary tumor probably reflects overall tumor behavior rather than a local response only (51).

Kalff et al. (56) graded response visually as complete, partial, or absent. After a median follow-up of 3.1 y, all 17 patients with a complete metabolic response were free of disease. Among the 10 patients with a partial metabolic response, only 6 were free of disease. All 3 metabolic nonresponders had died. Another study in which overall survival and disease-free survival were applied as outcome measures was performed by Capirci et al. (55). This (by far the largest) study showed that evaluation of the pathologic stage combined with  $^{18}\text{F}$ -FDG PET at restaging identified a subgroup of patients characterized by a good response to chemoradiotherapy and a more favorable prognosis. Cascini et al. (54) performed  $^{18}\text{F}$ -FDG PET at baseline, 12 d after the start of chemoradiotherapy ( $n = 33$ ), and after the completion of chemoradiotherapy ( $n = 17$ ). That study was one of 2 studies (47,54) in which  $^{18}\text{F}$ -FDG PET was evaluated at an earlier and perhaps more clinically relevant stage of treatment. As early as 12 d after the start of chemoradiation, responders were identified correctly by decreases in the mean SUV ( $\text{SUV}_{\text{mean}}$ ) (with a decrease of  $\geq 52\%$ , the accuracy was 100%) and  $\text{SUV}_{\text{max}}$  (with a decrease of  $\geq 42\%$ , the accuracy was 94%). Measurement of the  $\text{SUV}_{\text{mean}}$  most likely better reflects the behavior of the entire tumor mass, in which the heterogeneity and the architecture of tumor cells (viable cells mixed with fibrosis or necrosis) must be considered.

**TABLE 3. (Preoperative) Multimodality and Radiotherapy Treatment Response Evaluation in Primary Rectal Cancer**

Stage	Authors	Year	Reference	No. of patients	Therapy	Timing after therapy of PET evaluation	PET response criteria	Outcome measures	Results	P
Recurrent nonresectable tumor	Engenhart et al.	1992	40	21	Radiotherapy	8–9 wk	SUV normalization to background	Local control	PPV 20%; NPV 67%	
cT3NXM0	Schiepers et al.	1999	41	9	Radiotherapy	2–3 wk	$-\Delta MR_{glu}$	Histopathology, cell kinetics	Accuracy 80%	
cT3 or N1	Guillem et al.	2000	49	15	Chemoradiation	4–5 wk	$-\Delta SUV$ , visual response score, $-\delta TLG$	Histopathology	PPV 100%	
cT2, cT3, cT4, N1, N2, or N3	Oku et al.	2002	46	40	Radiotherapy	3–5 wk	SUV cutoff level after radiotherapy 3.2; SUV ratio 3.2; SUV ratio	Recurrence, CT shrinkage rate	Responders <3.2; nonresponders >3.2	0.046
cT3 or cT4	Amthauer et al.	2004	44	20	Chemoradiation + hyperthermia	2–4 wk	$-\Delta SUV_{max,BSA} > 36\%$	Histopathology	Correlation $-0.383$ PPV 93%; NPV 100%	0.014 <0.001
cT2NXM0, cT3NXM0, or cT4NXM0	Calvo et al.	2004	50	25	Chemoradiation	4–5 wk	$-\Delta SUV_{max}$	Histopathology (T downstaging)	Responders $-3.3$ ; nonresponders $-1.9$	0.03
cT3, cT4, or N1	Guillem et al.	2004	51	15	Chemoradiation	4–5 wk	$-\Delta SUV_{max} \geq 62.5\%$ , visual response score, $-\delta TLG \geq 69.5\%$	Recurrence, overall survival, recurrence-free survival	Responders $-69\%$ ; nonresponders 37%	0.004
cT3 or cT4	Denecke et al.	2005	52	23	Chemoradiation + hyperthermia	2–4 wk	$-\Delta SUV_{max} > 36\%$	Histopathology (T downstaging)	Responders $-60\%$ ; nonresponders $-37\%$ PPV 77%; NPV 100%	0.03 0.002
uT3, uT4, or N1	Konski et al.	2005	53	20	Chemoradiation	3–4 wk	$-\Delta SUV_{max}$	Histopathology	Responders $-75\%$ ; nonresponders $-52\%$	0.24
cT3, cT4, or N1	Cascini et al.	2006	54	33	Chemoradiation	Day 12 during and after	$-\Delta SUV_{mean} > 52\%$ (after 12 d)	Histopathology (tumor regression grade)	Responders $-63\%$ ; nonresponders $-22\%$ ; accuracy 100%	<0.0001
cT3, cT4, N1M0, N2M0, or N3M0	Capirci et al.	2006	55	88	Chemoradiation	5–6 wk	PET-positive  PET-negative (SUV <sub>max</sub> < 0.9)	5-y overall survival  5-y disease-free survival	PET-negative: 5-y OS 91%; PET-positive: 5-y DFS 62% PET-negative: 5-y DFS 81%; PET-positive: 5-y OS 72%	0.024 0.003
cT3NXM0 or cT4NXM0	Kalif et al.	2006	56	34	Chemoradiation	7–43 d	Visual response score	Overall survival	CMR: 3-y PFS 100%; PMR: 3-y OS 79% CMR: 3-y OS 100%; PMR: 3-y PFS 47%	<0.0001 <0.0001

cT3 or cT4	Capirci et al.	2007	57	45	Chemoradiation	5–6 wk	$-\Delta\text{SUV}_{\text{max}} > 66.2\%$	Histopathology (tumor regression grade, T downstaging)	Responders –76%; nonresponders –47%; PPV 77%; NPV 89%; accuracy 80%	0.0015
cT3, cT4, or N1	Melton et al.	2007	58	21	Chemoradiation	4–5 wk	$-\Delta\text{SUV}_{\text{max}} > 70\%$ , visual response score, $-\delta\text{TLG}$ , $-\Delta\text{CT}/-\Delta\text{PET}$ volume	Histopathology (T downstaging)	Responders –72%; nonresponders –44%; PPV 58%; NPV 100%	<0.001
cT3 or cT4	Kristiansen et al.	2008	59	30	Chemoradiation	7 wk	Visual response on PET/CT	Histopathology (tumor regression grade)	PPV 50%; NPV 58%	
uT2N+ or uT3N0+	Siegel et al.	2008	47	32	Short course of radiotherapy	Days 7–8 after start	$-\Delta\text{SUV}_{\text{max}} > 40\%$	Histopathology (tumor regression grade)	No correlation between PET responses and outcome measures	NS
uT3, uT4, or N1	Nakagawa et al.	2008	48	59	Radiotherapy	2–3 wk	SUV cutoff level after radiotherapy 5 SUV ratio < 100%	Median survival 5-y overall survival Histopathology	Responders 95 mo; nonresponders 42 mo Responders 70%; nonresponders 44% Relative risk 0.239 Significant correlation	0.042 0.028 0.047
cT3 or cT4	Vliegen et al.	2008	60	20	Chemoradiation	4–6 wk	$-\Delta\text{SUV}_{\text{max}}$	Histopathology (tumor regression grade)	Responders –83%; nonresponders –59%	0.025
cT3NX, cT4NX, or N1	Konski et al.	2008	61	53	Chemoradiation	3–4 wk	$-\Delta\text{SUV}_{\text{max}}$	Histopathology, disease-free survival	Responders –67%; nonresponders –55%	0.08

PPV = positive predictive value; NPV = negative predictive value;  $\Delta\text{MR}_{\text{glu}}$  = fractional change in  $\text{MR}_{\text{glu}}$ ;  $\Delta\text{SUV}$  = fractional change in SUV; SUV ratio = ratio of SUV between follow-up and baseline scans;  $\Delta\text{SUV}_{\text{max,BSA}}$  =  $\Delta\text{SUV}_{\text{max}}$  corrected for body surface area;  $\Delta\text{SUV}_{\text{max}}$  = fractional change in  $\text{SUV}_{\text{max}}$ ;  $\Delta\text{SUV}_{\text{mean}}$  = fractional change in  $\text{SUV}_{\text{mean}}$ ; OS = overall survival; DFS = disease-free survival; CMR = complete metabolic response; PFS = progression-free survival; PMR = partial metabolic response;  $\Delta\text{CT}/\Delta\text{PET}$  volume = change in lesion volume based on CT or PET measurements; NS = not significant.

In evaluations of the pelvic region (as in patients with rectal cancer), the fusion of metabolic imaging and morphologic imaging is especially advantageous for better lesion localization and thus for reducing interpretation pitfalls (such as those associated with nonspecific  $^{18}\text{F}$ -FDG uptake in the bowel lumen, cavity of the uterus, inflammatory processes, and muscles). Only a few studies were performed with an integrated PET/CT system (57–60). Capirci et al. (57) studied 44 patients at baseline and at 5–6 wk after the completion of chemoradiation and used the pathologic response as an outcome measure. When they used a decrease in the  $\text{SUV}_{\text{max}}$  of 66.2% as the cutoff value (identified by receiver operating characteristic curve analysis) for identifying a response to therapy, Capirci et al. (57) found 79.2% specificity, 81.2% sensitivity, 77% positive predictive value, 89% negative predictive value, and 80% overall accuracy. Kristiansen et al. (59) evaluated the predictive value of integrated PET/CT in primary rectal cancer. Because they administered contrast medium before performing both PET and CT, their results may have been influenced by attenuation correction artifacts on the PET scan, resulting in a low predictive value of this multimodality imaging approach (62–65).

Because the influence of contrast enhancement (in CT) on the calculation of SUVs has not yet been totally clarified, it is preferable to use non-contrast-enhanced CT data for attenuation correction to avoid small errors in attenuation correction factors when one is monitoring changes in SUVs with PET/CT (66). However, on this topic the current literature is controversial, and some authors believe that contrast-enhanced CT can be used for attenuation correction without any difficulty (67). Although it is not clear yet whether contrast-enhanced CT should be used for attenuation correction, it can still be used for image fusion to aid in the differentiation of anatomic structures, improve lesion localization, and support lesion characterization. It is remarkable that the confounding radiotherapy-induced effects discussed earlier have less impact on the results of  $^{18}\text{F}$ -FDG PET when it is combined with chemotherapy or regional hyperthermia. This finding implies that the nature of the combination of treatment modalities for neoadjuvant therapy is important in the timing of  $^{18}\text{F}$ -FDG PET evaluations. Further studies are required to ascertain the exact sequence of time-dependent (radio)biologic effects during neoadjuvant multimodality treatment.

Despite the fact that these 19 studies (Table 3) were very heterogeneous with respect to the methods applied for PET quantification, the evaluation interval, the metabolic response criteria, and the clinical endpoints (histology or survival), most of the studies showed that  $^{18}\text{F}$ -FDG PET is a significant predictor of therapy outcome.

## CONCLUSION

Published data indicate that  $^{18}\text{F}$ -FDG PET has a high predictive value in the therapeutic management of colo-

rectal cancer. This technique could be an asset for improving patient care by reducing the effort, costs, and morbidity associated with ineffective treatment in nonresponders. The available studies on chemotherapy response monitoring in advanced colorectal cancer and on preoperative radiotherapy and multimodality treatment response evaluation in primary rectal cancer indicate that  $^{18}\text{F}$ -FDG PET is a significant predictor of therapy outcome in both situations. In primary rectal cancer,  $^{18}\text{F}$ -FDG PET is applicable after neoadjuvant treatment in a preoperative setting (important for the preoperative selection for an individually tailored surgical approach) and correlates better with pathology than morphologic imaging modalities. Interestingly, when  $^{18}\text{F}$ -FDG PET is able to predict the final outcome, it may be used to guide adjuvant chemotherapy for rectal cancer after optimal neoadjuvant and local treatments.  $^{18}\text{F}$ -FDG PET could play a central role in optimizing the use of local ablative treatment of liver metastases because it recognizes, at early times, incomplete tumor ablation that is not detectable by CT.  $^{18}\text{F}$ -FDG PET could play a pivotal role in determining the need for further investigations and in guiding the reading of CT scans; the interpretation of the latter alone at early times after local ablative therapy appears to be difficult. Furthermore,  $^{18}\text{F}$ -FDG PET may be helpful in shortening the duration of early clinical trials assessing new antineoplastic agents. Therefore, therapy response assessment with  $^{18}\text{F}$ -FDG PET remains a very worthwhile research topic. The reported findings call for systematic implementation in randomized trials comparing PET-controlled strategies to adequately position  $^{18}\text{F}$ -FDG PET in treatment time lines.

## REFERENCES

1. Jemal A, Siegel R, Ward E, et al. Cancer statistics, 2008. *CA Cancer J Clin*. 2008;58:71–96.
2. Chong G, Cunningham D. Gastrointestinal cancer: recent developments in medical oncology. *Eur J Surg Oncol*. 2005;31:453–460.
3. Punt CJ. New options and old dilemmas in the treatment of patients with advanced colorectal cancer. *Ann Oncol*. 2004;15:1453–1459.
4. Bennett JJ, Cao D, Posner MC. Determinants of unresectability and outcome of patients with occult colorectal hepatic metastases. *J Surg Oncol*. 2005;92:64–69.
5. de Geus-Oei LF, Wiering B, Krabbe PF, Ruers TJ, Punt CJ, Oyen WJ. FDG-PET for prediction of survival of patients with metastatic colorectal carcinoma. *Ann Oncol*. 2006;17:1650–1655.
6. de Geus-Oei LF, Ruers TJ, Punt CJ, Leer JW, Corstens FH, Oyen WJ. FDG-PET in colorectal cancer. *Cancer Imaging*. 2006;6(suppl):S71–S81.
7. Mijnhout GS, Riphagen II, Hoekstra OS. Update of the FDG PET search strategy. *Nucl Med Commun*. 2004;25:1187–1189.
8. Therasse P, Arbutnot SG, Eisenhauer EA, et al. New guidelines to evaluate the response to treatment in solid tumors. European Organization for Research and Treatment of Cancer, National Cancer Institute of the United States, National Cancer Institute of Canada. *J Natl Cancer Inst*. 2000;92:205–216.
9. Avril NE, Weber WA. Monitoring response to treatment in patients utilizing PET. *Radiol Clin North Am*. 2005;43:189–204.
10. Findlay M, Young H, Cunningham D, et al. Noninvasive monitoring of tumor metabolism using fluorodeoxyglucose and positron emission tomography in colorectal cancer liver metastases: correlation with tumor response to fluorouracil. *J Clin Oncol*. 1996;14:700–708.
11. Bender H, Bangard N, Metten N, et al. Possible role of FDG-PET in the early prediction of therapy outcome in liver metastases of colorectal cancer. *Hybridoma*. 1999;18:87–91.

12. Dimitrakopoulou-Strauss A, Strauss LG, Rudi J. PET-FDG as predictor of therapy response in patients with colorectal carcinoma. *Q J Nucl Med.* 2003;47:8–13.
13. Dimitrakopoulou-Strauss A, Strauss LG, Burger C, et al. Prognostic aspects of <sup>18</sup>F-FDG PET kinetics in patients with metastatic colorectal carcinoma receiving FOLFOX chemotherapy. *J Nucl Med.* 2004;45:1480–1487.
14. de Geus-Oei LF, van Laarhoven HW, Visser EP, et al. Chemotherapy response evaluation with FDG PET in patients with colorectal cancer. *Ann Oncol.* 2007;19:348–352.
15. Lubezky N, Metser U, Geva R, et al. The role and limitations of 18-fluoro-2-deoxy-D-glucose positron emission tomography (FDG-PET) scan and computerized tomography (CT) in restaging patients with hepatic colorectal metastases following neoadjuvant chemotherapy: comparison with operative and pathological findings. *J Gastrointest Surg.* 2007;11:472–478.
16. Tan MC, Linehan DC, Hawkins WG, Siegel BA, Strasberg SM. Chemotherapy-induced normalization of FDG uptake by colorectal liver metastases does not usually indicate complete pathologic response. *J Gastrointest Surg.* 2007;11:1112–1119.
17. Goshen E, Davidson T, Zwas ST, Aderka D. PET/CT in the evaluation of response to treatment of liver metastases from colorectal cancer with bevacizumab and irinotecan. *Technol Cancer Res Treat.* 2006;5:37–43.
18. Akhurst T, Kates TJ, Mazumdar M, et al. Recent chemotherapy reduces the sensitivity of [<sup>18</sup>F]fluorodeoxyglucose positron emission tomography in the detection of colorectal metastases. *J Clin Oncol.* 2005;23:8713–8716.
19. Seki T, Wakabayashi M, Nakagawa T, et al. Percutaneous microwave coagulation therapy for solitary metastatic liver tumors from colorectal cancer: a pilot clinical study. *Am J Gastroenterol.* 1999;94:322–327.
20. Denecke T, Steffen I, Hildebrandt B, et al. Assessment of local control after laser-induced thermotherapy of liver metastases from colorectal cancer: contribution of FDG-PET in patients with clinical suspicion of progressive disease. *Acta Radiol.* 2007;48:821–830.
21. Ishii H, Okada S, Nose H, et al. Local recurrence of hepatocellular carcinoma after percutaneous ethanol injection. *Cancer.* 1996;77:1792–1796.
22. Hinshaw JL, Lee FT Jr. Cryoablation for liver cancer. *Tech Vasc Interv Radiol.* 2007;10:47–57.
23. Wood TF, Rose DM, Chung M, Allegra DP, Foshag LJ, Bilchik AJ. Radiofrequency ablation of 231 unresectable hepatic tumors: indications, limitations, and complications. *Ann Surg Oncol.* 2000;7:593–600.
24. Rossi S, Buscarini E, Garbagnati F, et al. Percutaneous treatment of small hepatic tumors by an expandable RF needle electrode. *AJR.* 1998;170:1015–1022.
25. Antoch G, Vogt FM, Veit P, et al. Assessment of liver tissue after radiofrequency ablation: findings with different imaging procedures. *J Nucl Med.* 2005;46:520–525.
26. Langenhoff BS, Oyen WJ, Jager GJ, et al. Efficacy of fluorine-18-deoxyglucose positron emission tomography in detecting tumor recurrence after local ablative therapy for liver metastases: a prospective study. *J Clin Oncol.* 2002;20:4453–4458.
27. Joosten J, Jager G, Oyen W, Wobbes T, Ruers T. Cryosurgery and radiofrequency ablation for unresectable colorectal liver metastases. *Eur J Surg Oncol.* 2005;31:1152–1159.
28. Donckier V, Van Laethem JL, Goldman S, et al. [<sup>18</sup>F] fluorodeoxyglucose positron emission tomography as a tool for early recognition of incomplete tumor destruction after radiofrequency ablation for liver metastases. *J Surg Oncol.* 2003;84:215–223.
29. Blokhuis TJ, van der Schaaf MC, van den Tol MP, Comans EF, Manoliu RA, van der Sijp JR. Results of radio frequency ablation of primary and secondary liver tumors: long-term follow-up with computed tomography and positron emission tomography-<sup>18</sup>F-deoxyfluoroglucose scanning. *Scand J Gastroenterol Suppl.* 2004;241:93–97.
30. Veit P, Antoch G, Stergar H, Bockisch A, Forsting M, Kuehl H. Detection of residual tumor after radiofrequency ablation of liver metastasis with dual-modality PET/CT: initial results. *Eur Radiol.* 2006;16:80–87.
31. Wong RK, Tandan V, De Silva S, Figueredo A. Pre-operative radiotherapy and curative surgery for the management of localized rectal carcinoma. *Cochrane Database Syst Rev.* 2007;(2):CD002102.
32. Bosset JF, Collette L, Calais G, et al. Chemotherapy with preoperative radiotherapy in rectal cancer. *N Engl J Med.* 2006;355:1114–1123.
33. Hospers GA, Punt CJ, Tesselar ME, et al. Preoperative chemoradiotherapy with capecitabine and oxaliplatin in locally advanced rectal cancer: a phase I-II multicenter study of the Dutch Colorectal Cancer Group. *Ann Surg Oncol.* 2007;14:2773–2779.
34. Gosens MJ, Dresen RC, Rutten HJ, et al. Preoperative radiochemotherapy is successful also in patients with locally advanced rectal cancer who have intrinsically high apoptotic tumours. *Ann Oncol.* 2008;19:2026–2032.
35. Bipat S, Glas AS, Slors FJ, Zwinderman AH, Bossuyt PM, Stoker J. Rectal cancer: local staging and assessment of lymph node involvement with endoluminal US, CT, and MR imaging—a meta-analysis. *Radiology.* 2004;232:773–783.
36. Kwok H, Bissett IP, Hill GL. Preoperative staging of rectal cancer. *Int J Colorectal Dis.* 2000;15:9–20.
37. Hoffmann KT, Rau B, Wust P, et al. Restaging of locally advanced carcinoma of the rectum with MR imaging after preoperative radio-chemotherapy plus regional hyperthermia. *Strahlenther Onkol.* 2002;178:386–392.
38. Rau B, Hunerbein M, Barth C, et al. Accuracy of endorectal ultrasound after preoperative radiochemotherapy in locally advanced rectal cancer. *Surg Endosc.* 1999;13:980–984.
39. Chen CC, Lee RC, Lin JK, Wang LW, Yang SH. How accurate is magnetic resonance imaging in restaging rectal cancer in patients receiving preoperative combined chemoradiotherapy? *Dis Colon Rectum.* 2005;48:722–728.
40. Engenhardt R, Kimmig BN, Strauss LG, et al. Therapy monitoring of presacral recurrences after high-dose irradiation: value of PET, CT, CEA and pain score. *Strahlenther Onkol.* 1992;168:203–212.
41. Schiepers C, Hausermans K, Geboes K, Filez L, Bormans G, Penninckx F. The effect of preoperative radiation therapy on glucose utilization and cell kinetics in patients with primary rectal carcinoma. *Cancer.* 1999;85:803–811.
42. Kubota R, Yamada S, Kubota K, Ishiwata K, Tamahashi N, Ido T. Intratumoral distribution of fluorine-18-fluorodeoxyglucose in vivo: high accumulation in macrophages and granulation tissues studied by microautoradiography. *J Nucl Med.* 1992;33:1972–1980.
43. Larson SM. Cancer or inflammation? A Holy Grail for nuclear medicine. *J Nucl Med.* 1994;35:1653–1655.
44. Amthauer H, Denecke T, Rau B, et al. Response prediction by FDG-PET after neoadjuvant radiochemotherapy and combined regional hyperthermia of rectal cancer: correlation with endorectal ultrasound and histopathology. *Eur J Nucl Med Mol Imaging.* 2004;31:811–819.
45. Haberkorn U, Strauss LG, Dimitrakopoulou A, et al. PET studies of fluorodeoxyglucose metabolism in patients with recurrent colorectal tumors receiving radiotherapy. *J Nucl Med.* 1991;32:1485–1490.
46. Oku S, Nakagawa K, Momose T, et al. FDG-PET after radiotherapy is a good prognostic indicator of rectal cancer. *Ann Nucl Med.* 2002;16:409–416.
47. Siegel R, Dresel S, Koswig S, et al. Response to preoperative short-course radiotherapy in locally advanced rectal cancer: value of f-fluorodeoxyglucose positron emission tomography. *Onkologie.* 2008;31:166–172.
48. Nakagawa K, Yamashita H, Nakamura N, et al. Preoperative radiation response evaluated by 18-fluorodeoxyglucose positron emission tomography predicts survival in locally advanced rectal cancer. *Dis Colon Rectum.* 2008;51:1055–1060.
49. Guillem JG, Puig-La CJ Jr, Akhurst T, et al. Prospective assessment of primary rectal cancer response to preoperative radiation and chemotherapy using 18-fluorodeoxyglucose positron emission tomography. *Dis Colon Rectum.* 2000;43:18–24.
50. Calvo FA, Domper M, Matute R, et al. <sup>18</sup>F-FDG positron emission tomography staging and restaging in rectal cancer treated with preoperative chemoradiation. *Int J Radiat Oncol Biol Phys.* 2004;58:528–535.
51. Guillem JG, Moore HG, Akhurst T, et al. Sequential preoperative fluorodeoxyglucose-positron emission tomography assessment of response to preoperative chemoradiation: a means for determining long-term outcomes of rectal cancer. *J Am Coll Surg.* 2004;199:1–7.
52. Denecke T, Rau B, Hoffmann KT, et al. Comparison of CT, MRI and FDG-PET in response prediction of patients with locally advanced rectal cancer after multimodal preoperative therapy: is there a benefit in using functional imaging? *Eur Radiol.* 2005;15:1658–1666.
53. Konski A, Hoffman J, Sigurdson E, et al. Can molecular imaging predict response to preoperative chemoradiation in patients with rectal cancer? A Fox Chase Cancer Center prospective experience. *Semin Oncol.* 2005;32(suppl):S63–S67.
54. Cascini GL, Avallone A, Delrio P, et al. <sup>18</sup>F-FDG PET is an early predictor of pathologic tumor response to preoperative radiochemotherapy in locally advanced rectal cancer. *J Nucl Med.* 2006;47:1241–1248.
55. Capirci C, Rubello D, Chierichetti F, et al. Long-term prognostic value of <sup>18</sup>F-FDG PET in patients with locally advanced rectal cancer previously treated with neoadjuvant radiochemotherapy. *AJR.* 2006;187:W202–W208.
56. Kalf J, Duong C, Drummond EG, Matthews JP, Hicks RJ. Findings on <sup>18</sup>F-FDG PET scans after neoadjuvant chemoradiation provides prognostic stratification in patients with locally advanced rectal carcinoma subsequently treated by radical surgery. *J Nucl Med.* 2006;47:14–22.
57. Capirci C, Rampin L, Erba PA, et al. Sequential FDG-PET/CT reliably predicts response of locally advanced rectal cancer to neo-adjuvant chemo-radiation therapy. *Eur J Nucl Med Mol Imaging.* 2007;34:1583–1593.

58. Melton GB, Lavelly WC, Jacene HA, et al. Efficacy of preoperative combined 18-fluorodeoxyglucose positron emission tomography and computed tomography for assessing primary rectal cancer response to neoadjuvant therapy. *J Gastrointest Surg*. 2007;11:961–969.
59. Kristiansen C, Loft A, Berthelsen AK, et al. PET/CT and histopathologic response to preoperative chemoradiation therapy in locally advanced rectal cancer. *Dis Colon Rectum*. 2008;51:21–25.
60. Vliegen RF, Beets-Tan RG, Vanhauten B, et al. Can an FDG-PET/CT predict tumor clearance of the mesorectal fascia after preoperative chemoradiation of locally advanced rectal cancer? *Strahlenther Onkol*. 2008;184:457–464.
61. Konski A, Li T, Sigurdson E, et al. Use of molecular imaging to predict clinical outcome in patients with rectal cancer after preoperative chemotherapy and radiation. *Int J Radiat Oncol Biol Phys*. November 10, 2008 [Epub ahead of print].
62. Nakamoto Y, Chin BB, Kraitchman DL, Lawler LP, Marshall LT, Wahl RL. Effects of nonionic intravenous contrast agents at PET/CT imaging: phantom and canine studies. *Radiology*. 2003;227:817–824.
63. Antoch G, Freudenberg LS, Egelhof T, et al. Focal tracer uptake: a potential artifact in contrast-enhanced dual-modality PET/CT scans. *J Nucl Med*. 2002;43:1339–1342.
64. Beyer T, Antoch G, Muller S, et al. Acquisition protocol considerations for combined PET/CT imaging. *J Nucl Med*. 2004;45(suppl 1):25S–35S.
65. Blodgett TM, McCook BM, Federle MP. Positron emission tomography/computed tomography: protocol issues and options. *Semin Nucl Med*. 2006;36:157–168.
66. Weber WA, Figlin R. Monitoring cancer treatment with PET/CT: does it make a difference? *J Nucl Med*. 2007;48(suppl 1):36S–44S.
67. Yau YY, Chan WS, Tam YM, et al. Application of intravenous contrast in PET/CT: does it really introduce significant attenuation correction error? *J Nucl Med*. 2005;46:283–291.