Radiation Dosimetry of β-Amyloid Tracers ¹¹C-PiB and ¹⁸F-BAY94-9172

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β-Amyloid (Aβ) imaging has great potential to aid in the diagnosis of Alzheimer disease and the development of therapeutics. The radiation dosimetry of A_β radioligands may influence their application; therefore, we calculated and compared the effective doses (EDs) of ¹¹C-PiB and a new ¹⁸F-labeled ligand, ¹⁸F-BAY94-9172. Methods: Attenuation-corrected whole-body scans were performed at 0, 15, 30, 45, and 60 min after injection of 350 \pm 28 MBq (mean \pm SD) of $^{11}\text{C-PiB}$ in 6 subjects and at 0, 20, 60, 120, and 180 min after injection of 319 \pm 27 MBq of $^{18}\text{F-}$ BAY94-9172 in 3 subjects. Coregistered CT was used to define volumes of interest (VOIs) on the PET images. The source organs were the brain, lungs, liver, kidneys, spleen, and vertebrae. The VOIs for the contents of the gallbladder, urinary bladder, lower large intestine, upper large intestine, and small intestine were also defined. Total activity in each organ at each time point was calculated by use of reference organ volumes. The resultant time-activity curves were fitted with constrained exponential fits, and cumulated activities were determined. A dynamic bladder voiding model was used. The OLINDA/EXM program was used to calculate the whole-body EDs from the acquired data. Results: For ¹¹C-PiB, the highest absorbed doses were in the gallbladder wall (44.80 \pm 29.30 μ Gy/MBq), urinary bladder wall $(26.30 \pm 8.50 \,\mu\text{Gy/MBq})$, liver (19.88 $\pm 3.58 \,\mu\text{Gy/MBq})$, and kidneys (12.92 \pm 3.37 $\mu\text{Gy/MBq}$). The ED was 5.29 \pm 0.66 $\mu\text{Sv/}$ MBg. For ¹⁸F-BAY94-9172, the highest doses were also in the gallbladder wall (132.40 \pm 43.40 μ Gy/MBg), urinary bladder wall (24.77 \pm 7.36 μ Gy/MBq), and liver (39.07 \pm 8.31 μ Gy/ MBq). The ED was 14.67 \pm 1.39 μ Sv/MBq. Conclusion: The estimated organ doses for ¹¹C-PiB were comparable to those reported in earlier research. With the doses used in published studies (300-700 MBq), the EDs would range from 1.6 to 3.7 mSv. The ED of ¹⁸F-BAY94-9172 was 30% lower than that of ¹⁸F-FDG and, at the published dose of 300 MBq, would yield an ED of 4.4 mSv. The dosimetry of both Aβ radioligands is suitable for clinical and research applications.

Key Words: PET; radiation dosimetry; ¹¹C-PiB; ¹⁸F-BAY94-9172

J Nucl Med 2009; 50:309–315 DOI: 10.2967/jnumed.108.055756 A myloid plaques are one of the pathologic hallmarks of Alzheimer disease (AD). They consist of extracellular aggregates of β -amyloid (A β) (1,2) peptide that have diameters of about 50–100 μ m and that are intimately surrounded by dystrophic axons and dendrites, reactive astrocytes, and activated microglia (3).

Several hypotheses have been postulated to explain the molecular mechanisms leading to AD (4,5), but the A β theory is the dominant etiologic paradigm at this time (2). The hypothesis states that an imbalance between the production and the removal of A β leads to its progressive accumulation and resultant synaptic dysfunction and neuronal loss, clinically manifested as a loss of cognitive functions (6,7). New treatment strategies for AD are aimed at delaying disease onset or slowing disease progression, through either preventing the deposition of A β or increasing the solubilization of A β . Given that these treatments are currently in trials, there is an urgent need for early disease recognition (8–10).

Amyloid imaging with PET is allowing new insights into A β deposition in the brain. "Pittsburgh Compound-B," or ¹¹C-PiB, is the most widely used PET radioligand for assessing A β in the brain (*11–13*). However, the 20.4-min radioactive half-life of ¹¹C restricts the use of PiB to PET centers with an on-site cyclotron and extensive radiochemistry infrastructure. Each patient dose requires a cyclotron run and radiosynthesis of ¹¹C-PiB immediately before the scan. An ¹⁸F-labeled A β tracer is needed to permit wider application of amyloid imaging. ¹⁸F has a 109.4-min half-life, so that a single large production should permit multiple scans to be performed at multiple PET sites, as is currently the case with ¹⁸F-FDG imaging.

We have evaluated a novel ¹⁸F-labeled tracer developed by Zhang et al. at the University of Pennsylvania (*14*). The tracer, ¹⁸F-BAY94-9172, is a stilbene derivative that has shown high affinity and specificity for A β in vitro and binding to amyloid plaques but not neurofibrillary tangles in postmortem human brain tissue (*14*). In a proof-ofconcept study in 35 subjects, ¹⁸F-BAY94-9172 PET images were able to reliably distinguish subjects with AD from

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healthy elderly subjects and subjects with frontotemporal dementia (11).

The aim of this study was to calculate the dosimetry of ¹¹C-PiB and ¹⁸F-BAY94-9172 in healthy elderly control subjects to assess safety and suitability for clinical application from a radiodosimetric perspective.

MATERIALS AND METHODS

Subjects

Nine healthy control subjects were recruited, 6 for the ¹¹C-PiB protocol and 3 for the ¹⁸F-BAY94-9172 protocol. Details for the subjects recruited and for the 2 protocols are given in Table 1. Subjects were assessed by neuropsychological testing and physical examination. This study was approved by the Austin Health Human Research Ethics Committee, and informed consent was obtained from all subjects before the imaging studies.

Radiosynthesis

¹¹C-PiB and ¹⁸F-BAY94-9172 were synthesized by use of ¹¹C and ¹⁸F produced with an in-house 10-MeV cyclotron (Ion Beam Applications SA).

For ¹¹C-PiB synthesis, ¹¹C-6-OH-benzothiazole (BTA)-1 was produced in a one-step reaction of ¹¹C-methyl triflate (15-17) with the 6-OH-BTA-0 precursor. The product was reformulated with the Sep-Pak (Waters) method and filtered. The radioligand purity averaged 99%, and the specific radioactivity averaged 18 GBq/µmol.

 18 F-BAY94-9172 was produced from the PEGN3-OMs (methylsulfonate) precursor and purified with a semipreparative column by the method of Zhang et al. (*14*). The product was reformulated with the Sep-Pak method. The radioligand purity averaged 96.0%, and the specific radioactivity averaged 140.5 GBq/µmol.

PET Protocol

Serial PET was performed with either a Philips Gemini/GS2 PET/CT scanner or a Philips Allegro dedicated PET scanner. Both systems are germanium oxyorthosilicate 3-dimensional scanners and have identical physical characteristics: an axial field of view of 180 mm, 45 image slices, a slice thickness of 4 mm, and a central spatial resolution of 5 mm full width at half maximum.

All subjects underwent whole-body (WB) low-dose CT (ldCT) with the Gemini PET/CT scanner. PET images were then acquired with the Gemini PET/CT scanner for the ¹¹C-PiB studies and the Allegro dedicated PET scanner for the ¹⁸F-BAY94-9172 studies. In both cases, ldCT was used for anatomic localization to define

volumes of interest (VOIs) for subsequent dosimetric analysis. For the ¹¹C-PiB scans, ldCT was additionally used for attenuation correction of the PET emission scans. Before the ¹⁸F-BAY94-9172 WB scans with the Allegro camera, an attenuation correction scan was acquired by use of a ¹³⁷Cs rotating point source with the Allegro camera.

Subjects were scanned in the supine position with the arms down. For the purposes of this study, a WB scan included the area from the top of the subject's head to the groin and ranged from 880 to 1,020 mm in axial extent. After a planning CT scan, a single WB ldCT scan was acquired with 30 mAs per slice and a 0.5-s rotation time. Subjects then underwent 5 consecutive WB scans. Each WB scan consisted of 10 or 11 bed positions with 1 min of acquisition time per bed position. Subjects were permitted to leave the scanner between the third and fourth and between the fourth and fifth ¹⁸F-BAY94-9172 WB scans. Additional attenuation correction scans were acquired with a ¹³⁷Cs rotating point source before the fourth and fifth WB scans. The imaging sequences for ¹¹C-PiB and ¹⁸F-BAY94-9172 are described in Table 2. For both ¹¹C-PiB scanning and ¹⁸F-BAY94-9172 scanning, a standard cylindric source of activity (with an activity of 5 MBq and a volume of 5 mL) was placed opposite the subject's head for calibration confirmation purposes.

PET Data Analysis

The Gemini and Allegro scans were reconstructed with the RAMLA-3D algorithm after being corrected for attenuation, dead time, half-life decay, and scatter with the single-scatter simulation (SSS) correction (18). PET images were generated with an image size of 144×144 per slice and a voxel size of $4 \times 4 \times 4$ mm³. For scans acquired with the Allegro scanner, the reconstructed images were reconstructed with standardized uptake value (SUV) units that were defined as

$$SUV = \frac{C(t) \times 2^{t/\tau_{1/2}}}{D/M_{body}},$$
 Eq. 1

where C(t) is the voxel radioactivity concentration (kBq/mL) at time t after injection, $\tau_{1/2}$ is the isotope decay half-life, D is the injected dose (MBq), and M_{body} (kg) is the mass of the subject.

The camera sensitivity is calibrated on a quarterly basis with a uniform cylindric phantom (dimensions: diameter, 20 cm; length, 30 cm; and volume, 9,420 mL) filled with a well counter–calibrated

TABLE 1. Subject Details and Injected Radioactivities of ¹¹ C-PiB and ¹⁸ F-BAY94-9172					
Tracer or parameter	Subject	Sex	Age (y)	Mass (kg)	Injected dose (MBq)
¹¹ C-PiB	1	F	63	68.5	370.1
	2	F	56	66.0	377.7
	3	F	70	61.0	363.9
	4	М	65	93.5	344.3
	5	Μ	64	79.0	300.2
	6	М	61	88.0	352.1
Mean ± SD			63.2 ± 4.2	76.0 ± 11.8	351.4 ± 25.4
¹⁸ F-BAY94-9172	1	М	63.5	101	299.3
	2	Μ	73.5	103	307.2
	3	F	63.1	66.9	349.5
Mean ± SD			66.7 ± 4.8	90.3 ± 16.6	318.7 ± 22.0

1 ¹⁸ F-BAY94-9172	WB Scanning
Time (min)	Scan
-10	IdCT
0	WB1
15	WB2
30	WB3
45	WB4
60	WB5
-20	IdCT
-5	¹³⁷ Cs AC
0	WB1
20	WB2
60	WB3
110	¹³⁷ Cs AC
120	WB4
170	¹³⁷ Cs AC
180	WB5
	Time (min) -10 0 15 30 45 60 -20 -5 0 20 60 110 120 170 180

Five WB scans (WB1–WB5) were done for each tracer. ¹³⁷Cs transmission scan was repeated when subject was repositioned on scanner after being removed from scanner to allow for bladder voiding. AC = attenuation correction.

concentration of ¹⁸F-FDG. A weekly validation of the calibration is performed with a uniform ⁶⁸Ge phantom (dimensions: diameter, 20 cm; length, 20 cm; and volume, 6,283 mL) that is cross-calibrated with respect to a calibrated ¹⁸F-FDG source.

¹¹C SUV Validation

In addition to the quarterly ¹⁸F-FDG calibration and weekly ⁶⁸Ge validation, a ¹¹C-phantom validation was undertaken. The 9,420-mL phantom was filled with 370 MBq of ¹¹C-PiB, and images were acquired under conditions identical to those used for subject imaging. Three bed positions were used to encompass the entire phantom, and ldCT was used to allow CT-based attenuation correction. Five scans were acquired with a 1-min duration per bed position and starting acquisition times identical to those used for ¹¹C-PiB WB scanning (Table 2). After reconstruction, representative circular regions of interest were defined, and mean SUVs were determined. The resultant mean SUVs were in agreement with the ¹⁸F-FDG–based routine SUV calibration results.

Dosimetry

For both ¹¹C-PiB and ¹⁸F-BAY94-9172, after reconstruction, VOIs were defined on the basis of ldCT. The organs defined by the ldCT VOI process were the brain, lungs, liver, kidneys, spleen, and vertebrae. For these organs, representative regions were defined on the basis of ldCT and, when applied to emission images, allowed the determination of the average SUV for an organ. Because representative regions of these organs were used, quantitation issues arising from effects, such as respiratory motion effects, on the dome of the liver and the partial volume at the boundary of organs were minimized. By use of the known mass of an organ or its contents, the mean SUV could be converted to percentage injected radioactivity, as expressed in Equation 2. For organs in which the radioactivity of the contents was considered, such as the gallbladder, urinary bladder, lower large intestine, upper large intestine, and small intestine, VOIs were defined on the basis of dynamic PET emission images.

The VOIs for organs and organ contents were then applied to the coregistered PET images from which the mean SUV was determined. The activity in each organ was then determined by half-life decaying the SUV followed by scaling by the ratio of the reference man (19) organ mass or organ content volume to that of the study subject body mass as is represented in Equation 2. This procedure provided an estimate of the activity present in a reference man organ at time t for the appropriate tracer. By repeating this analysis for each time point, we obtained a time– activity curve for a standardized organ; the data were expressed as percentage injected radioactive dose (%IRD) (as distinct from %ID, which generally refers to percentage injected pharmacologic dose), as follows:

$$\% IRD_{organ}(t) = \overline{SUV}_{organ}(t) \times 2^{-t/\tau_{1/2}} \times \frac{M_{organ}}{M_{body}} \times 100, \quad \text{Eq. 2}$$

where $\% IRD_{organ}(t)$ and $\overline{SUV}_{organ}(t)$ are, respectively, the %IRD and the mean SUV of the organ of interest determined at time *t* after injection and M_{organ} is the mass of the organ of interest.

The cumulated activity of the urinary bladder contents was determined by fitting the time–activity curve for the bladder contents with an exponential in-growth function of the form

$$U(t) = U(0) \times (1 - e^{-t/\tau}),$$
 Eq. 3

where U(t) is the fraction of activity in the bladder at time *t* after injection and τ is the isotope half-time (where $\tau = \tau_{1/2}/\ln 2$). These parameters were entered into the bladder voiding model module of OLINDA/EXM (20) with a voiding interval of 2.4 h.

The OLINDA/EXM application developed by Stabin et al. (20) was used to determine the effective doses (EDs) for individual organs on the basis of the set of source organs defined in the organ markup stage. Stabin et al. introduced the concept of cumulated activity, as opposed to the MIRDOSE concept of residence time (21). In the framework of Stabin et al., the dose absorbed by a target organ, denoted as r_k , is expressed as

$$D(r_k) = \sum_h N_h \times S(r_k \leftarrow r_h), \qquad \text{Eq. 4}$$

where $S(r_k \leftarrow r_h)$ (21) is the S factor (which Stabin et al. (20) referred to as the dose fraction), which is the fraction of the dose absorbed by the target organ (r_k) from radiation emitted by the source organ (r_h) . The number of disintegrations occurring in the source organ (N_h) was referred to by Stabin et al. as the cumulated activity in that organ.

For both ¹¹C-PiB and ¹⁸F-BAY94-9172, constrained monoexponential fits were applied to the standardized organ %IRD time–activity curves to allow parametric calculation of cumulated activities.

For determination of the cumulated activity of the gallbladder contents, the area under the curve (AUC) was calculated from the available time–activity curve via the trapezoidal rule method. After the final imaging time point, when a single exponential decay function with a half-life equal to the physical decay constant is applied, it is assumed that there is no further excretion and that clearance occurs only through the physical decay of the isotope. The resultant reference man-normalized cumulated activities were entered into OLINDA/EXM as source organs from which the EDs were calculated for a range of sources or target organs by use of MIRD methods (22,23). The radiation transport phantom selected from OLINDA/EXM was the hermaphroditic phantom, which is based on the measurements of Cristy and Eckerman (24) for a 73.7-kg adult phantom but nominally refers to the 70-kg adult phantom from ICRP Publication 23 (19). As a result, the radiation dose estimates were considered to apply to both male and female subjects with a body weight of 73.7 kg.

RESULTS

WB coronal projections of a subject infused with ¹¹C-PiB are shown in Figure 1. It is visually evident that there were high levels of accumulation of ¹¹C-PiB in the liver, gallbladder, and urinary bladder and significant levels of accumulation in the kidneys, spleen, and upper large intestine. Similar patterns of uptake in the liver, urinary bladder, and upper large intestine were observed for ¹⁸F-BAY94-9172, as shown in Figure 2. In addition, compared with ¹¹C-PiB, for ¹⁸F-BAY94-9172, there was evidence of higher levels of uptake in the spleen as well as in the lower large intestine and small intestine, as shown in Figure 3.

Cumulated activities were determined for individual subjects to allow individualized dose estimates to be determined. Measures of the reproducibility of cumulated activities after reference man organ normalization for both ¹¹C-PiB and ¹⁸F-BAY94-9172 are shown in Figures 4 and 5, respectively; these figures show the population-averaged %IRD time–activity curves. The data for the subset of organs shown demonstrated good reproducibility, with the bladder contents showing the greatest variation, particularly for ¹⁸F-BAY94-9172; in that study, subjects had the opportunity to void the urinary bladder. The large variability in bladder content measurements shown for ¹⁸F-BAY94-9172 in Figure 5 could be addressed with a more rigorous bladder voiding regimen and a sample size larger than the sample size of 3 used in the present study.

The OLINDA/EXM results for ¹¹C-PiB and ¹⁸F-BAY94-9172 are shown in Table 3, along with the EDs reported by



FIGURE 2. Coronal projections (100 mm thick) of 3 sequential WB images (0, 60, and 180 min after injection of ¹⁸F-BAY94-9172) of subject 1. Images are displayed on SUV scale, giving indication of differences in %IRDs.

the ICRP Publication 60 (25). Table 3 shows the ¹¹C-PiB organ dosimetry for the 6 subjects for whom the ED was determined 5.29 \pm 0.66 µSv/MBq (mean \pm SD). The organs receiving the highest absorbed doses were the gallbladder wall (44.80 \pm 29.30 µGy/MBq), urinary bladder wall (26.30 \pm 8.50 µGy/MBq), liver (19.88 \pm 3.58 µGy/MBq), and kidneys (12.92 \pm 3.37 µGy/MBq).

Table 3 shows the ¹⁸F-BAY94-9172 organ dosimetry for the 3 subjects for whom the ED was determined to be 14.67 \pm 1.39 μ Sv/MBq. As with ¹¹C-PiB, the organs receiving the highest doses were the gallbladder wall (132.40 \pm 43.40 μ Gy/MBq), urinary bladder wall (24.77 \pm 7.36 μ Gy/ MBq), and liver (39.07 \pm 8.31 μ Gy/MBq).

DISCUSSION

As anticipated, the organ and WB EDs for ¹¹C-PiB were substantially lower than those for ¹⁸F-BAY94-9172 because of the much shorter decay half-life of ¹¹C (20.4 min) than of ¹⁸F (109.8 min). Both tracers showed substantial clearance through the liver and excretion of radioactivity into the bowel. Both tracers also showed considerable renal excretion, so that the data for the critical organs (i.e., gallbladder wall, liver, and urinary bladder wall) were the



FIGURE 1. Coronal projections (100 mm thick) of 3 sequential WB images (0, 30, and 60 min after injection of ¹¹C-PiB) of subject 1. Images are displayed on SUV scale (range, 0–10), giving indication of differences in %IRDs.



FIGURE 3. Coronal projections (100 mm thick) of WB images of 3 subjects 3 h after injection with ¹⁸F-BAY94-9172, illustrating variable levels of radioactivity in contents of large intestine.



FIGURE 4. ¹¹C-PiB time-activity curves for liver, brain, kidneys, and bladder contents, expressed as %IRD normalized to reference man (*19*). Variability represented by error bars resulted from differences in individual subjects, placement of regions of interest, and emission image statistics.

same. However, a relatively high radioactive dose of ¹¹C-PiB (typically 555 MBq) is recommended to yield reasonable image quality at the time of acquisition, between 40 and 70 min after injection (2–3.5 half-lives after injection for ¹¹C). Thus, the WB ED for the recommended dose of ¹¹C-PiB is 2.9 mSv. The optimal imaging time for ¹⁸F-BAY94-9172 is between 90 and 120 min after injection (0.8–1.1 half-lives for ¹⁸F), and excellent results have been achieved with 300 MBq (*11*) at this time point. This dose of ¹⁸F-BAY94-9172 yields a WB ED of 4.4 mSv.

Scheinin et al. (26) previously studied ¹¹C-PiB radiation dosimetry in humans by using a different methodology. In the present study, time–activity curves were determined for each individual across the entire body and then used to calculate the dosimetry for each individual. In contrast, Scheinin et al. (26) determined averaged time-activity curves by scanning different sets of subjects for different anatomic regions with 2 different PET scanners. Results were then scaled to the reference man, and an ED was determined. There was generally good agreement between the data of Scheinin et al. (26) and those in the present study, as summarized in Table 4.

Table 5 summarizes the EDs of other common ¹⁸Fradiopharmaceuticals, ¹⁸F-BAY94-9172, and ¹¹C-PiB. The ED of ¹⁸F-BAY94-9172 was generally somewhat lower than the EDs of the other ¹⁸F-labeled radiopharmaceuticals, as the organs with the highest ¹⁸F-BAY94-9172 uptake had relatively low radiosensitivity.

The radiation doses from diagnostic levels of infused activities of ¹¹C-PiB make it well suited to serial studies, but



FIGURE 5. ¹⁸F-BAY94-9172 timeactivity curves for liver, brain, kidneys, and bladder contents, expressed as %IRD normalized to reference man (*19*). Variability represented by error bars resulted from differences in individual subjects, placement of regions of interest, and emission image statistics. High variability in bladder contents illustrated different patterns of individual bladder voiding.

TABLE 3. ¹¹C-PiB and ¹⁸F-BAY94-9172 Radiation Doses Determined With 73.7-kg Hermaphroditic Adult Phantom

	Mean ± SD dose of:		
Organ or	¹¹ C-PiB	¹⁸ F-BAY94-9172	
parameter	(n = 6)	(n = 3)	
Adrenal glands	4.00 ± 0.32	15.30 ± 1.56	
Brain	3.92 ± 1.44	10.28 ± 1.55	
Breasts	2.21 ± 0.23	9.05 ± 0.56	
Gallbladder wall	44.80 ± 29.30	132.40 ± 43.40	
Lower large intestine wall	3.10 ± 0.35	15.73 ± 0.61	
Small intestine	4.65 ± 0.74	14.03 ± 0.42	
Stomach wall	3.11 ± 0.28	13.20 ± 1.04	
Upper large intestine wall	3.87 ± 0.43	17.83 ± 2.18	
Heart wall	3.24 ± 0.34	12.90 ± 0.96	
Kidneys	12.92 ± 3.37	20.03 ± 5.59	
Liver	19.88 ± 3.58	39.07 ± 8.31	
Lungs	6.59 ± 3.31	8.26 ± 0.56	
Muscle	2.57 ± 0.24	10.90 ± 0.66	
Ovaries	3.21 ± 0.33	13.70 ± 0.80	
Pancreas	4.02 ± 0.33	15.93 ± 1.63	
Red marrow	2.43 ± 0.20	18.93 ± 4.28	
Osteogenic cells	3.60 ± 0.48	22.03 ± 4.16	
Skin	1.98 ± 0.20	8.38 ± 0.43	
Spleen	4.03 ± 0.57	15.13 ± 7.02	
Testes	2.12 ± 0.10	10.06 ± 5.82	
Thymus	2.54 ± 0.31	11.07 ± 0.80	
Thyroid	2.24 ± 0.21	10.47 ± 0.35	
Urinary bladder wall	26.30 ± 8.50	24.77 ± 7.36	
Uterus	3.73 ± 0.42	13.83 ± 0.90	
Total body	3.23 ± 0.30	12.23 ± 0.95	
ED equivalent (μSv/MBq)	8.93 ± 1.80	23.17 ± 4.29	
ED (μSv/MBq)	5.29 ± 0.66	14.67 ± 1.39	
	0.445		
Data are reported in	ι μGy/MBq unless	otherwise indicated.	

the short physical half-life of ¹¹C, which requires individual dose production immediately before use, is a disadvantage. The ED of ¹⁸F-BAY94-9172 is somewhat lower than that of ¹⁸F-FDG; with judicious selection of infused activities, ¹⁸F-BAY94-9172 may be suitable for serial studies. Some restrictions on the number of ¹⁸F-BAY94-9172 studies performed per year on healthy subjects will be needed to comply with radiation protection guidelines for research subjects. Although the ED of ¹⁸F-BAY94-9172 is almost 3 times higher than that

TABLE 4. Comparison of ¹¹ C-PiB Dosimetry from Scheinin et al. (<i>26</i>) and Present Study				
	μGy/MBq from:			
Organ	Scheinin et al.	Present study		
Gallbladder wall	41.5	44.80 ± 29.30		
Urinary bladder wall	16.6	26.30 ± 8.50		
Liver	19.0	19.88 ± 3.58		
Kidney	12.6	12.92 ± 3.37		
Upper large intestine wall	9.0	3.87 ± 0.43		

TABLE 5. Comparison of ED (Adult Phantom Model) Estimates for ¹¹C-PiB, ¹⁸F-BAY94-9172, and Other ¹⁸F-Labeled Pharmaceuticals

Tracer (study)	ED (μSv/MBq)
¹¹ C-PiB	5.29 ± 0.66
¹¹ C-PiB (26)	4.74
¹⁸ F-BAY94-9172	14.67 ± 1.39
¹⁸ F-FDG (22,23)	19.0
¹⁸ F-A85380 (27)	19.4
^{99m} Tc-phosphonate* (23)	5.7
*Common SPECT agent	
Common of Eor agent.	

of ¹¹C-PiB, the longer physical half-life of ¹⁸F-BAY94-9172 than of ¹¹C-PiB is an advantage for multicenter studies and widespread clinical use.

CONCLUSION

The radiation dosimetry for amyloid imaging agents ¹¹C-PiB and ¹⁸F-BAY94-9172 has been calculated; the EDs are $5.29 \pm 0.66 \,\mu$ Sv/MBq and $14.67 \pm 1.39 \,\mu$ Sv/MBq, respectively. The calculated ED of ¹¹C-PiB is in good agreement with a previous measurement made by Scheinin et al. (26). With diagnostic imaging dose levels of 550 MBq for ¹¹C-PiB and 350 MBq for ¹⁸F-BAY94-9172, the radiation doses of ¹¹C-PiB and ¹⁸F-BAY94-9172 will be 1.85 and 5.13 mSv, respectively.

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