

The Radiotoxicity of ^{131}I Therapy of Thyroid Cancer: Assessment by Micronucleus Assay of B Lymphocytes

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We investigated cytologic radiation damage in thyroid cancer after ^{131}I therapy using micronucleus assay (MNA) of B lymphocytes exclusively, as opposed to our previous study in which MNA of all lymphocyte subsets was used. **Methods:** We studied 22 thyroid cancer patients treated with 3.7 GBq of ^{131}I . Peripheral lymphocytes were harvested, and B lymphocytes were isolated by an immunomagnetic method and assayed for the frequency of micronuclei. **Results:** The frequency of micronuclei among B cells after ^{131}I therapy was significantly increased relative to that in untreated control subjects, and the ^{131}I -induced increase in micronuclei frequency among B cells was significantly greater than that among all lymphocytes. **Conclusion:** Compared with the MNA of all lymphocytes, the MNA among specifically B cells may more sensitively detect cytologic radiation damage associated with ^{131}I therapy of thyroid cancer.

Key Words: micronucleus assay; thyroid cancer; radioiodine therapy; radiotoxicity; B lymphocytes

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Dosimetric studies have estimated the radiation dose to blood and bone marrow after treatment with a large amount of radioiodine (1). Chromosomal damage to peripheral lymphocytes in vivo after radioiodine therapy has been reported (2,3). In a previous study using micronucleus assay (MNA), we too demonstrated cytologic radiation damage to lymphocytes in thyroid cancer patients after ^{131}I therapy (4). To our knowledge, however, the cytologic effects of large therapeutic doses of radioiodine on specific subpopulations of lymphocytes have not been examined.

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Because B cells are more sensitive to radiation in vitro than are other lymphocyte subpopulations (5,6), Vral et al. (7) suggested that MNA of B cells might be particularly useful for assessment of cytologic radiation damage in vivo. Therefore, this study used MNA to compare radiation-induced cytologic damage to B lymphocytes with that to peripheral lymphocytes generally in thyroid cancer patients after therapeutic administration of 3.7 GBq of ^{131}I . Chromosomal damage to lymphocytes induced in vivo by ^{131}I should augment the cells with micronuclei (8). A comparison of the increased frequency of micronuclei among B lymphocytes and peripheral lymphocytes irradiated in vivo by ^{131}I may sensitively estimate cytologic radiation damage in thyroid cancer patients.

MATERIALS AND METHODS

Patients

From May 1996 to Feb 2002, 22 patients were admitted for radioiodine treatment of differentiated thyroid carcinoma at the Department of Radiology of Toyama Medical and Pharmaceutical University in Japan. There were 9 men and 13 women, ranging in age from 40 to 71 y (mean, 56.9 y). All patients were diagnosed by biopsy and underwent total thyroidectomy. After giving informed consent, the 22 patients, who received 3.7 GBq of ^{131}I , were studied. The thyroid-stimulating hormone concentration was higher than 30 $\mu\text{U}/\text{mL}$ in these patients on the day before therapy.

Lymphocyte Isolation and Culture

Peripheral blood lymphocytes were collected from each patient before and 1 wk after therapy. The lymphocytes were cultured according to the method of Fenech and Morley (8). Pokeweed mitogen (Gibco BRL) was used to stimulate the lymphocytes to proliferate in culture. A solution of cytochalasin B (Aldrich Chemical Co.) was added 44 h after the commencement of the culture. The cultures were terminated 72 h after initiation.

External Irradiation

In an in vitro study, lymphocytes were collected from 4 young, healthy volunteers and were irradiated with electron beams (3

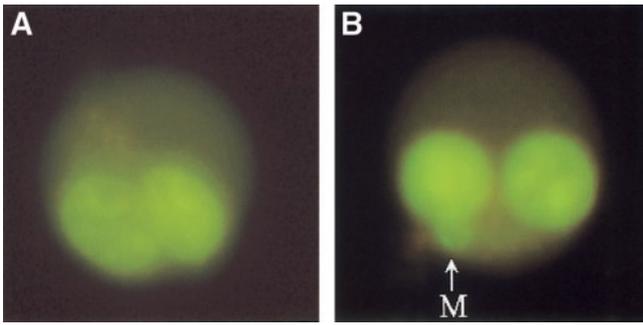


FIGURE 1. Photomicrographs of binucleated cells, including one without (A) and one with (B) a micronucleus (M).

Gy/min, 5 MeV, Metatron, MD 2/40; Toshiba) at doses ranging from 0.5 to 2 Gy and were assessed for cytologic damage by MNA.

MNA

Anti-CD 19 (anti-Pan B) antibody-conjugated Dynabeads M-450 magnetic beads (DynaL AS) were incubated with lymphocytes at 4°C for 15 min. The rosetted CD 19-positive B cells were isolated using a magnet. Dynabeads were detached from B cells by incubation with polyclonal antibody DETACHaBEAD (DynaL AS) at 36°C for 60 min. After incubation, B cells were isolated by a magnet (9). The acridine orange fluorescent staining procedure was applied to the MNA in this study according to the method of Hayashi et al. (10). Micronuclei were stained yellowish green with green fluorescence (Fig. 1). The number of micronuclei per 500 binucleated cells was scored visually.

The Previous Study

We reevaluated the increased number of peripheral lymphocyte micronuclei found after therapy in the 25 thyroid cancer patients of our previous study (4).

Statistical Analysis

The number of B-cell micronuclei before and after therapy was measured. The 22 patients represented a new patient group, whose data were compared with those of another group of 25 similar thyroid cancer patients analyzed in our previous study. Data are expressed as mean \pm SD. The Student paired or unpaired *t* test was used to analyze differences between means. Probability values of <0.01 were considered significant.

RESULTS

Radiation Damage to Lymphocytes

The number of micronuclei per 500 binucleated B lymphocytes was 6.8 ± 1.9 before and 22.5 ± 5.4 after ^{131}I therapy, a statistically significant increase ($P < 0.001$). The number of micronuclei per 500 binucleated peripheral lymphocytes was 5.4 ± 1.4 before and 15.7 ± 2.7 after ^{131}I therapy in our previous study (Fig. 2). The increased number of B lymphocyte and peripheral lymphocyte micronuclei after therapy was 15.6 ± 4.8 and 10.0 ± 3.1 , and these differences were statistically significant ($P < 0.001$) (Fig. 3). The percentage increases in the frequencies of micronuclei were $247\% \pm 105\%$ for B cells and $212\% \pm 132\%$ for all lymphocytes. Although these differences were not statistically significant, a trend toward higher values was noted in the former.

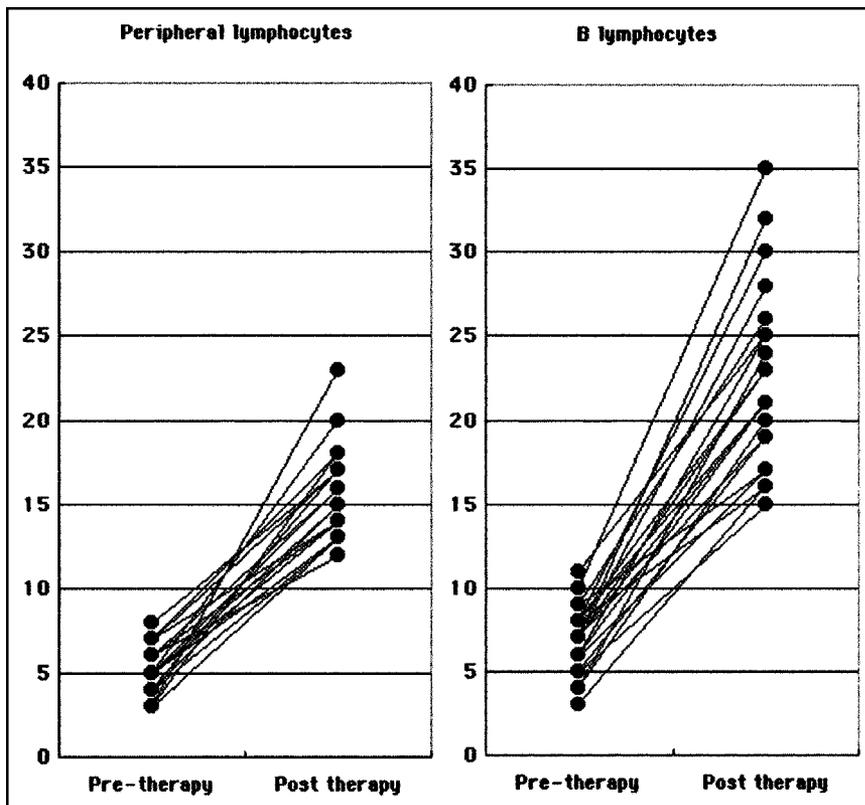


FIGURE 2. The frequency of micronuclei among peripheral lymphocytes and B lymphocytes before and after ^{131}I therapy.

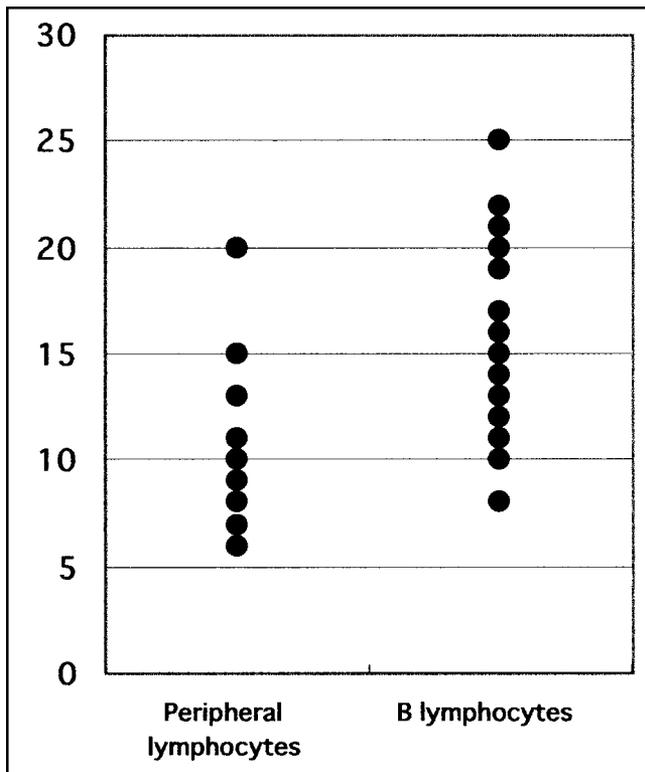


FIGURE 3. The increased number of micronuclei among peripheral lymphocytes and B lymphocytes.

Dose Estimation

The number of B-cell micronuclei was measured after electron irradiation *in vitro* at absorbed doses of 0.5–2 Gy, and the resulting data fit a straight line, $y = 35.7x + 6.3$ ($r = 0.95$). Based on this dose–response relationship and the observed frequency of B-cell micronuclei *in vivo*, the absorbed dose among the 22 thyroid cancer patients treated with ^{131}I was 0.45 ± 0.14 Gy.

DISCUSSION

MNA was used to assess radiation-induced cytologic damage to B lymphocytes in thyroid cancer patients after therapeutic administration of 3.7 GBq of ^{131}I . In this study, B lymphocytes (CD19) were collected by an immunomagnetic technique (Dynabeads) after being cultured from peripheral blood lymphocytes. This Dynabeads method for separation of cultured lymphocytes is simple and fast (9). Our findings showed that B lymphocytes exposed to ^{131}I have statistically higher frequencies of micronuclei than do controls. We also revealed that the degree of micronucleus induction among B lymphocytes after ^{131}I therapy is higher than that among peripheral lymphocytes. Thus, radiation-induced damage to B lymphocytes after ^{131}I therapy may be greater than that to peripheral lymphocytes *in vivo*.

In an *in vitro* study, Wuttke et al. confirmed that B cells are more radiosensitive than T cells (11). Vral et al. also demonstrated the highly radiosensitive behavior of B cells

with respect to micronucleus production *in vitro* (7). On the basis of immune function assessment (12) and apoptosis (13), B cells appear to be more radiosensitive than other lymphocyte subpopulations. However, the radiosensitivity of B lymphocytes to radiotherapy *in vivo* has not been clarified until now. In a previous study (4), we used phytohemagglutinin to stimulate peripheral lymphocytes, which are a mixture of mainly 3 groups of cells: B lymphocytes, T lymphocytes, and natural killer cells. Generally, phytohemagglutinin stimulates T cells, and phytohemagglutinin-stimulated lymphocytes can be considered predominantly T cells. Our finding of a higher degree of micronucleus induction *in vivo* for B lymphocytes than for peripheral lymphocytes appears to be consistent with the *in vitro* results noted above. These results suggest that B lymphocytes may express high radiosensitivity *in vivo* after ^{131}I therapy.

We evaluated an external-irradiation study to compare the micronucleus incidence in B lymphocytes irradiated *in vivo* with that in B lymphocytes irradiated *in vitro* and to estimate the biologically equivalent absorbed dose to B lymphocytes *in vivo*. We determined that cytologic damage to B lymphocytes *in vivo* after ^{131}I therapy may be equivalent to the damage observed after a mean external-irradiation dose of 0.45 Gy *in vitro*. In our previous study, using similar methods, we estimated that peripheral lymphocytes receive a dose of 0.33 Gy (4). Our estimated radiation dose was somewhat higher than the previous estimation, possibly because of the limited number of control subjects available for *in vitro* studies; B lymphocytes were collected from only 4 young, healthy volunteers, and peripheral lymphocytes from 3 patients. One may argue that the total dose from low-dose-rate radiation exposure *in vivo* with ^{131}I is considerably underestimated when compared with high-dose-rate external-radiation exposure *in vitro* (14). Therefore, both doses may be underestimated.

The estimated radiation dose with ^{131}I therapy in thyroid cancer may vary. Monsieurs et al., using lymphocyte MNA, showed that the mean equivalent dose was 0.32 Gy (15). M'Kacher et al., using a similar method, estimated the biologic dose to be 0.54 Gy (3). For the patients in our study, using B-lymphocyte MNA, the mean estimated internal radiation exposure appeared to be in a range similar to these results.

Internal radiation damage after ^{131}I therapy in thyroid cancer was slightly greater to B lymphocytes than to peripheral lymphocytes, based on the comparative cytologic studies using MNA. These results suggest that B-lymphocyte MNA could be more sensitive for assessing cytologic radiation-induced chromosomal damage. It is true that the small number of subjects limits our results and that we believe the dose to have been underestimated. Nevertheless, this approach remains the most sensitive and practical method presently available to evaluate cytologic radiation damage in thyroid cancer after ^{131}I therapy.

CONCLUSION

Compared with MNA of all lymphocytes, MNA of specifically B cells may more sensitively detect cytologic radiation damage associated with ^{131}I therapy of thyroid cancer and may facilitate estimation of the radiation doses absorbed with this therapy.

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