

Synthesis and Biologic Evaluation of Monocationic Asymmetric ^{99m}Tc -Nitride Heterocomplexes Showing High Heart Uptake and Improved Imaging Properties

Alessandra Boschi, PhD¹; Licia Uccelli, PhD¹; Cristina Bolzati, PhD¹; Adriano Duatti, PhD¹; Nicola Sabba, PhD¹; Elena Moretti, BS¹; Giovanni Di Domenico, PhD²; Guido Zavattini, PhD²; Fiorenzo Refosco, PhD³; and Melchiorre Giganti, MD¹

¹Laboratory of Nuclear Medicine, Department of Clinical and Experimental Medicine, University of Ferrara, Ferrara, Italy;

²Department of Physics, University of Ferrara, Ferrara, Italy; and ³Istituto di Chimica Inorganica e delle Superfici, Consiglio Nazionale delle Ricerche, Padua, Italy

The preparation, characterization, and first biologic evaluation in rats of a novel class of monocationic ^{99m}Tc heart imaging agents are reported. The complexes are represented by the general formula $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$, where L is the monoanionic form of a dithiocarbamate ligand of the type $[\text{R}^1(\text{R}^2)\text{-N-C(=S)S}]^-$, PNP is a diphosphine ligand of the type $[(\text{R}^3)_2\text{P-(CH}_2\text{)}_2\text{]}_2\text{-N(R}^4\text{)}$, and $\text{R}^1\text{-R}^4$ are organic functional groups. **Methods:** The new complexes were prepared by use of both liquid and freeze-dried formulations through a 2-step procedure. The first step involved the formation of the $[\text{Tc}\equiv\text{N}]^{2+}$ group through the reaction of $^{99m}\text{TcO}_4^-$ with succinic dihydrazide as a donor of nitride nitrogen atoms (N^{3-}) in the presence of Sn^{2+} ions. This step was followed by the simultaneous addition to the reaction solution of the ligand PNP and of the sodium salt of the dithiocarbamate ligand (NaL) to afford the final products, $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$. The chemical identities of the resulting ^{99m}Tc complexes were determined by comparing their chromatographic properties with those of the corresponding ^{99}Tc analogs prepared by use of the long-lived isotope ^{99}Tc and characterized by spectroscopic and crystallographic techniques. Ex vivo biodistribution studies were conducted in rats. In vivo tomographic images of the rat heart were obtained by use of a small-animal SPECT scanner. **Results:** The $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes are monocationic and possess a distorted square-pyramidal geometry in which the $\text{Tc}\equiv\text{N}$ multiple bond occupies an apical position and the diphosphine and dithiocarbamate ligands span the residual 4 coordination positions on the basal plane through the 2 phosphorus atoms and the 2 sulfur atoms, respectively. Imaging and biodistribution studies demonstrated that these radiopharmaceuticals localize selectively in the myocardium of rats and are retained in this region for a prolonged time. The kinetics of heart uptake and clearance were found to be influenced by variations in the lateral $\text{R}^1\text{-R}^4$ groups. Blood and lung washouts were extremely fast. Elimination occurred mostly through the kidneys

and the liver. Surprisingly, at 1 h after injection, liver activity was almost negligible. Analysis of heart-to-liver and heart-to-lung uptake ratios showed that these values increased exponentially over time and became much higher than those determined for ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin. These findings were confirmed by analysis of high-quality SPECT images collected in rats for the new complexes and compared with images obtained with ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin. **Conclusion:** The high myocardial uptake and the very high heart-to-lung and heart-to-liver uptake ratios indicate that the $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes exhibit very favorable distribution properties and could be used to obtain SPECT cardiac images with improved quality.

Key Words: ^{99m}Tc heart perfusion imaging agents; asymmetric ^{99m}Tc -nitride heterocomplexes; mixed diphosphino-dithiocarbamate ^{99m}Tc -nitride complexes

J Nucl Med 2003; 44:806–814

In the last 2 decades, cationic ^{99m}Tc tracers have been developed as $^{201}\text{Tl}^+$ analogs for the imaging of myocardial perfusion. The most successful cationic, lipophilic ^{99m}Tc perfusion agents are the well-known compounds ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin (1–3). Some years ago, the bis(*N*-ethoxy,*N*-ethyl)dithiocarbamato nitride technetium-99m ($^{99m}\text{Tc-N-NOET}$) complex was introduced as the first example of a neutral heart imaging agent exhibiting prolonged retention in the myocardium (4–8). Surprisingly, the biologic behavior of this compound appeared to be more similar to that of the monocation $^{201}\text{Tl}^+$ than to that of the above-mentioned cationic complexes (4,7).

Despite the great success of current heart imaging agents, their properties are still far from those expected for an ideal perfusion tracer. In particular, the relatively low first-pass extraction of ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin makes these tracers less suitable for measuring blood flow at high

Received Aug. 21, 2002; revision accepted Jan. 20, 2003.

For correspondence or reprints contact: Adriano Duatti, PhD, Laboratory of Nuclear Medicine, Department of Clinical and Experimental Medicine, University of Ferrara, Via L. Borsari 46, 44100 Ferrara, Italy.

E-mail: dta@unife.it

flux rates. Moreover, the persistent liver uptake observed for all cardiac imaging agents completely prevents the assessment of defects located at the inferoapical myocardial wall. As a consequence, there still exists considerable interest in the search for a myocardial perfusion agent having imaging characteristics much closer to those predicted for an ideal perfusion agent (9).

The Tc(V)-nitride complex ^{99m}Tc -N-NOET represents the first reported example of a heart imaging agent characterized by the presence of a terminal $\text{Tc}\equiv\text{N}$ multiple bond. It has a square-pyramidal geometry in which 2 identical bidentate dithiocarbamate ligands are symmetrically bound to a $\text{Tc}\equiv\text{N}$ group in the basal plane of the square-pyramid (5,10). We describe here the preparation and characterization of a new class of Tc(V)-nitride cardiac imaging agents. These complexes are monocationic and are represented by the general formula $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$, where L is the monoanionic form of a dithiocarbamate ligand of the type $[\text{R}^1(\text{R}^2)\text{-N-C(=S)S}]^-$, PNP is a diphosphine ligand of the type $[\text{R}^3_2(\text{P})\text{-}(\text{CH}_2)_2\text{-N}(\text{R}^4)]$, and $\text{R}^1\text{-R}^4$ are organic functional lateral groups (Fig. 1). The unique structural feature of this class of ^{99m}Tc -nitride complexes is attributable to the fact that they are composed of 2 different bidentate ligands bound to the same $\text{Tc}\equiv\text{N}$ group (Fig. 1). This composition gives rise to a new type of 5-coordinated heterocomplexes having an unusual asymmetric structure (11). These compounds were prepared after the development of a somewhat complex chemistry based on the chemical properties of the new metal nitride fragment $[^{99m}\text{Tc}(\text{N})(\text{PNP})]^{2+}$. This fragment is formed by a $\text{Tc}\equiv\text{N}$ group coordinated to a PNP ligand and is easily produced in an aqueous physiologic solution as the dichloride derivative $[^{99m}\text{Tc}(\text{N})(\text{PNP})]\text{Cl}_2$. It was found that this precursor complex exhibits a strong electrophilic character and selectively reacts with electron-rich, soft, π -donor ligands, having S, O, and N as coordinating atoms, without removal of the $\text{Tc}\equiv\text{N}$ group and of the ancillary PNP ligand (12,13). Thus, the formation of

$[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ heterocomplexes can be viewed as resulting from the selective coupling of the metal fragment $[^{99m}\text{Tc}(\text{N})(\text{PNP})]^{2+}$ with the bidentate dithiocarbamate π -donor ligand L.

A biologic evaluation of the new asymmetric heterocomplexes was performed in rats. The results showed that these radiopharmaceuticals localized selectively in the rat myocardium and were retained in this region for a prolonged time. Liver washout was extremely fast and quantitative. Tomographic images of the rat heart were obtained by use of a small-animal SPECT scanner and compared with those collected with ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin. These data, along with the unprecedented high heart-to-lung and heart-to-liver uptake ratios, suggest that this class of perfusion agents could be conveniently used to obtain high-quality cardiac images. A brief account of the preliminary data on the preparation and biodistribution in rats of only 2 members of the large class of complexes described here has been communicated elsewhere (14).

MATERIALS AND METHODS

Reagents and Physical Measurements

The diphosphine ligands bis(dimethoxypropylphosphinoethyl)-methoxyethylamine [PNP3; $(\text{CH}_3\text{OC}_3\text{H}_6)_2\text{P}(\text{CH}_2)_2\text{-N}(\text{C}_2\text{H}_4\text{OCH}_3)$ - $(\text{CH}_2)_2\text{P}(\text{C}_3\text{H}_6\text{OCH}_3)_2$], bis(dimethoxypropylphosphinoethyl)-ethoxyethylamine [PNP5; $(\text{CH}_3\text{OC}_3\text{H}_6)_2\text{P}(\text{CH}_2)_2\text{-N}(\text{C}_2\text{H}_4\text{-OCH}_2\text{CH}_3)$ - $(\text{CH}_2)_2\text{P}(\text{C}_3\text{H}_6\text{OCH}_3)_2$], and bis(diethoxypropylphosphinoethyl)ethoxyethylamine [PNP6; $(\text{CH}_3\text{CH}_2\text{OC}_3\text{H}_6)_2\text{P}(\text{CH}_2)_2\text{-N}(\text{C}_2\text{H}_4\text{OCH}_2\text{CH}_3)$ - $(\text{CH}_2)_2\text{P}(\text{C}_3\text{H}_6\text{OCH}_2\text{CH}_3)_2$] (Fig. 2) were prepared by following published procedures (13,15) or were obtained from Argus Chemical. The sodium salts of dithiocarbamate ligands, $[\text{R}^1(\text{R}^2)\text{-N-C(=S)S}]^-\text{Na}$ (Fig. 2), were prepared by reacting the corresponding secondary amines with equivalent amounts of carbon disulfide in NaOH solutions as reported previously (5) or were obtained from Alchemy. Sodium diethyldithiocarbamate and sodium dimethyldithiocarbamate were commercially available (Aldrich Chimica). Succinic dihydrazide [SDH; $\text{H}_2\text{NNH}(\text{OC})\text{-}(\text{CH}_2)_2(\text{CO})\text{NHNH}_2$], ethylenediaminetetraacetic acid (EDTA), $\text{SnCl}_2\cdot 2\text{H}_2\text{O}$, tetrabutylammonium bromide, triphenylphosphine (PPh_3), and γ -cyclodextrin were purchased from Aldrich. All common solvents were reagent grade and were used without further purification. The radiopharmaceuticals ^{99m}Tc -sestamibi (Cardiolite; Dupont Radiopharmaceuticals) and ^{99m}Tc -tetrofosmin (Myoview; Amersham Healthcare) were obtained from commercial kit formulations.

Electrospray ionization (ESI) mass (16) spectra of technetium-99 compounds ($\sim 10^{-6}$ mol dm^{-3} solutions) were recorded on an LCQ instrument (Finnigan). The complexes were injected by use of a syringe pump at a flow rate of 5 $\mu\text{L}/\text{min}$.

Preparation of $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ Complexes

All preparations were made by excluding oxygen from vials and solutions by nitrogen filling and purging, respectively. The PNP and L ligands are illustrated in Figure 2.

Liquid Formulation. $\text{Na}^{99m}\text{TcO}_4$ (0.250 mL, 50.0 MBq-4.0 GBq) was mixed in a vial containing 5.0 mg of SDH, 5.0 mg of EDTA, 0.10 mg of $\text{SnCl}_2\cdot 2\text{H}_2\text{O}$ (suspended in 0.10 mL of saline), and 1.0 mL of ethanol. The mixture was kept at room temperature for 30 min. To the resulting solution, 1.0 mg of PNP (dissolved in

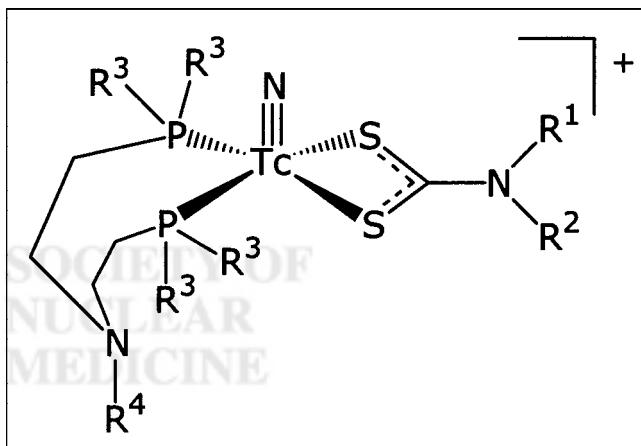


FIGURE 1. Chemical structure of $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes. PNP = diphosphine ligand; L = monoanionic dithiocarbamate ligand; $\text{R}^1\text{-R}^4$ = lateral organic functional groups.

L = [R ¹ (R ²)-N-C(=S)S] ⁻			L = [R ¹ (R ²)-N-C(=S)S] ⁻		
R ¹	R ²	Acronym	R ¹	R ²	Acronym
CH ₃	CH ₃	DMDC	CH ₃ (CH ₂) ₂ OCH ₂ CH ₂	CH ₃ (CH ₂) ₂ OCH ₂ CH ₂	DPOE
CH ₃ CH ₂	CH ₃ CH ₂	DEDC	CH ₃ CH ₂ OCH ₂ CH ₂	CH ₃ CH ₂ CH ₂ CH ₂	DBOET
CH ₃	CH ₃ O	NOME	CH ₃ OCH ₂ CH ₂	CH ₃ CH ₂ CH ₂ CH ₂	DBOME
CH ₃ CH ₂	CH ₃ CH ₂ O	NOET	CH ₃ CH ₂ OCH ₂ CH ₂	(CH ₃) ₂ CH	ISOET
CH ₃ CH ₂	CH ₃ OCH ₂ CH ₂	POET	CH ₃ OCH ₂ CH ₂	(CH ₃) ₂ CH	ISOME
CH ₃ CH ₂	CH ₃ CH ₂ OCH ₂ CH ₂	BOET	PNP = [(R ³) ₂ P-(CH ₂) ₂] ₂ -N(R ⁴)		
CH ₃ CH ₂ CH ₂	CH ₃ CH ₂ CH ₂	DPDC	R ³		
CH ₃ CH ₂	CH ₃ OCH ₂ CH ₂ CH ₂	PROME	CH ₃ OCH ₂ CH ₂ CH ₂	CH ₃ OCH ₂ CH ₂	PNP3
CH ₃ OCH ₂ CH ₂	CH ₃ OCH ₂ CH ₂	DPODC	CH ₃ OCH ₂ CH ₂ CH ₂	CH ₃ CH ₂ OCH ₂ CH ₂	PNP5
CH ₃ CH ₂ OCH ₂ CH ₂	CH ₃ (CH ₂) ₃ OCH ₂ CH ₂	DBODC	CH ₃ CH ₂ OCH ₂ CH ₂ CH ₂	CH ₃ CH ₂ OCH ₂ CH ₂	PNP6
CH ₃ (CH ₂) ₃ OCH ₂ CH ₂	CH ₃ (CH ₂) ₃ OCH ₂ CH ₂	DBOE	R ⁴		

FIGURE 2. Dithiocarbamate (L) and diphosphine (PNP) ligands and acronyms used in this study.

0.30 mL of ethanol) and 2.0 mg of the sodium salt of the appropriate dithiocarbamate ligand (dissolved in 0.10 mL of saline) were simultaneously added. The vial was heated at 100°C for 15 min. The yield was >95%.

Lyophilized Formulation. A lyophilized kit formulation was developed for the preparation of the [^{99m}Tc(N)(PNP3)(DBODC)]⁺ (where DBODC is bis-N-ethoxyethyl-dithiocarbamate) and [^{99m}Tc(N)(PNP5)(DBODC)]⁺ complexes as described below.

One milliliter of generator-eluted Na^{99m}TcO₄ (50.0 MBq—4.5 GBq) was added to a vial containing 5.0 mg of SDH, 5.0 mg of EDTA, 0.1 mg of SnCl₂·2H₂O, and 1.0 mL of phosphate buffer (0.1 mol dm⁻³) in a freeze-dried form. The resulting solution was kept at room temperature for 30 min. The contents of a second lyophilized vial—3.5 mg of PNP3 (or PNP5), 3.5 mg of DBODC, and 3.5 mg of γ-cyclodextrin—were reconstituted with 1.75 mL of saline. Then, 1.0 mL of the resulting solution was withdrawn from the second vial and added to the first vial, which was heated at 100°C for 15 min. The yield was >95%.

Chromatography

Radiochemical purity (RCP) was determined by thin-layer chromatography (TLC) and high-performance liquid chromatography (HPLC). TLC was performed on silica gel plates (Merck) with ethanol:chloroform:toluene:NH₄Ac (0.5 mol dm⁻³) (5:3:3:0.5) as the mobile phase and on reversed-phase C₁₈ plates (Merck) with saline:methanol:tetrahydrofuran:glacial acetic acid (2:8:1:1) as the mobile phase. Thin-layer chromatograms were analyzed with a Cyclone instrument equipped with a phosphorImaging screen and OptiQuant image analysis software (Packard Instruments). HPLC was performed on a System Gold Instrument (Beckman Instruments) equipped with programmable solvent module 126, scanning detector module 166, and radioisotope detector module 170. Chromatographic runs were performed on a reversed-phase C₁₈ column (Ultrasphere; 4.6 × 250 mm; Beckman) with a reversed-phase C₁₈ precolumn (Ultrasphere; 4.6 × 45 mm; Beckman). A methanol:phosphate buffer mixture (0.02 mol dm⁻³) was used as the mobile phase at a flow rate of 1.0 mL min⁻¹. R_f values and HPLC retention times (*t*_r) for some selected complexes are reported in Table 1.

Purification

To avoid the interference of free reagents with evaluation of the stability and the biodistribution of ^{99m}Tc-nitrile heterocomplexes,

a purification procedure was carried out as follows. A reversed-phase SepPak C₁₈ cartridge (Waters Corp.) was activated by passage of 5.0 mL of ethanol and 5.0 mL of deionized water. After preparation, the reaction solution containing the final [^{99m}Tc(N)(PNP(L)]⁺ heterocomplex was diluted by the addition of 8.0 mL of deionized water. The resulting solution was passed through the activated SepPak cartridge. Approximately 100% of the initial activity was retained on the cartridge. The cartridge was washed with 20.0 mL of deionized water and 3.0 mL of an ethanol:water mixture (80:20). The complex was recovered by passage through the cartridge of 1.0 mL of a mixture composed of a water solution of tetrabutylammonium bromide {[N(C₄H₉)₄]Br; 0.1 mol dm⁻³} and ethanol at a volume ratio of 10:90. Approximately 80% of the initial activity was finally collected.

TABLE 1
R_f Values, Retention Times (*t*_r), and log *k'* Values
for [^{99m}Tc(N)(PNP(L)]⁺ Complexes

Complex	TLC		HPLC	
	R _f , SiO ₂	reversed phase	<i>t</i> _r (min)	log <i>k'</i>
[^{99m} Tc(N)(PNP3)(DEDC)] ⁺	0.30	0.50	14.2	2.91
[^{99m} Tc(N)(PNP3)(NOET)] ⁺	0.40	0.49	15.8	2.79
[^{99m} Tc(N)(PNP3)(DMDC)] ⁺	0.24		10.1	2.83
[^{99m} Tc(N)(PNP3)(NOME)] ⁺	0.28		10.3	2.84
[^{99m} Tc(N)(PNP3)(DPDC)] ⁺	0.57		22.8	3.51
[^{99m} Tc(N)(PNP3)(DPODC)] ⁺	0.47		13.1	3.18
[^{99m} Tc(N)(PNP3)(DBODC)] ⁺	0.60	0.41	21.5	3.82
[^{99m} Tc(N)(PNP3)(BOET)] ⁺	0.45		17.4	3.24
[^{99m} Tc(N)(PNP3)(POET)] ⁺	0.38		13.6	2.88
[^{99m} Tc(N)(PNP3)(PROME)] ⁺	0.50		14.0	3.28
[^{99m} Tc(N)(PNP3)(DBOET)] ⁺	0.40		33.4	4.00
[^{99m} Tc(N)(PNP3)(DBOME)] ⁺	0.50		34.2	3.99
[^{99m} Tc(N)(PNP3)(DPOE)] ⁺	0.57			4.10
[^{99m} Tc(N)(PNP3)(DBOE)] ⁺	0.50			4.35
[^{99m} Tc(N)(PNP5)(DBODC)] ⁺	0.65	0.44	17.83	3.69
[^{99m} Tc(N)(PNP5)(NOME)] ⁺	0.50	0.17	8.05	2.48
[^{99m} Tc(N)(PNP5)(BOET)] ⁺	0.50	0.44		
[^{99m} Tc(N)(PNP3)(ISOET)] ⁺	0.60		25.08	3.46
[^{99m} Tc(N)(PNP6)(DBODC)] ⁺	0.84			

Determination of $\log k_0'$ Values

After purification, $\log k_0'$ values for some representative complexes were determined by HPLC as described elsewhere (17). The results are reported in Table 1. It was shown previously that $\log k_0'$ values are well correlated with $\log P$ values (P is the water:octanol partition coefficient) for lipophilic compounds (17). $\log k_0'$ is obtained from $\log k'$, where k' is the capacity factor measuring the partitioning of the compound between a nonpolar stationary phase and a polar mobile phase. The $\log k'$ values were determined here at various compositions of the mobile phase. An analysis of the relationship between $\log k'$ and mobile phase composition gave extrapolated $\log k'$ values at a 0% organic solvent composition ($\log k_0'$) as a measure of the activity partitioning between the hydrophobic stationary phase and water. The $\log k_0'$ values were extrapolated from the linear part of the curve $\log k' = a + bC$, where C is the methanol concentration and $\log k' = \log(t_r - t_o)/t_o$. The elution time (t_r) for a nonretained component was regarded as being equal to the elution time for $\text{Na}^{99\text{m}}\text{TcO}_4$. For each compound, retention data were measured by use of a minimum of 3 different methanol concentrations in the mobile phase.

Stability

The in vitro stability of the $[^{99\text{m}}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes was evaluated by monitoring RCP at different times (15, 30, 60, 120, and 240 min) with the following procedure. After preparation, the selected compound was purified by the reversed-phase SepPak procedure described above. Then, 100 μL of the collected radioactive eluate were incubated at 37°C with 900 μL of saline, phosphate buffer, rat serum, or human serum. RCP was measured by TLC analysis.

Cysteine and glutathione (GSH) are ligands containing sulfur donor atoms that are potentially capable of coordinating the metal fragment $[^{99\text{m}}\text{Tc}(\text{N})(\text{PNP})]^{2+}$ by substitution of a dithiocarbamate ligand (L) in $[^{99\text{m}}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes. Challenge experiments were performed with the following procedure. A 250- μL volume of phosphate buffer (0.2 mol dm^{-3} ; pH 7.4), 50 μL of a freshly prepared aqueous solution of L-cysteine (0.01 mol dm^{-3}) or, alternatively, of GSH (0.01 mol dm^{-3}), and 50 μL of the appropriate complex were placed in a test tube and incubated in a water bath at 37°C. A blank experiment was performed with an equal volume of distilled water in place of the cysteine (or GSH) solution. Aliquots of the resulting solutions were withdrawn at 15, 30, 60, and 120 min and analyzed by TLC.

Preparation of $[^{99\text{g}}\text{Tc}(\text{N})(\text{PNP})(\text{L})]\text{Cl}$ Complexes

$[^{99\text{g}}\text{Tc}(\text{N})(\text{PNP})(\text{L})]\text{Cl}$ complexes were prepared by reacting the ligands PNP3 or PNP5 and L (Fig. 2) with the precursor Tc(V)-nitride complex $[^{99\text{g}}\text{Tc}(\text{N})\text{Cl}_2(\text{PPh}_3)_2]$ (18,19) as briefly described below.

$[^{99\text{g}}\text{Tc}(\text{N})\text{Cl}_2(\text{PPh}_3)_2$ (0.04 mmol) was reacted with an equimolar amount of PNP in dichloromethane at room temperature for 30 min. To the resulting bright yellow solution, the sodium salt of the appropriate dithiocarbamate ligand (NaL) (0.04 mmol dissolved in ethanol) was added, and the mixture was stirred at room temperature for 1 h. After filtration, the solution was taken to dryness, and the resulting oily residue was treated with diethyl ether (5 mL). The yellow ethereal solution was recovered and evaporated to give a yellow compound that was further purified by overnight high-vacuum suction. The yield was 89%.

All $[^{99\text{g}}\text{Tc}(\text{N})(\text{PNP})(\text{L})]\text{Cl}$ complexes were characterized by elemental analysis and mass spectroscopy. The ESI mass spectrum

of the $[^{99\text{g}}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^+$ complex is shown in Figure 3. The ESI mass spectrum (m/z , percent abundance) was as follows: 847 [M] $^+$, 100; and 640 $\{\text{M}-(\text{CH}_2)_3\text{P}[(\text{CH}_2)_2\text{OCH}_3]\}_2^+$, 5.

Animal Studies

Biodistribution Studies. Animal experiments were performed in compliance with the relevant national laws relating to the conduct of animal experimentation and were approved by the Ferrara University hospital ethical committee. Before injection, the complexes were purified by the procedures outlined above. After purification, the collected activity was further diluted with phosphate buffer (0.1 mol dm^{-3} ; pH 7.4) to give a final solution with an ethanol content of 10%. When the dithiocarbamate ligand was nonsymmetric ($\text{R}^1 \neq \text{R}^2$), a racemic mixture of 2 enantiomeric complexes was obtained. These components were not separated, and the whole mixture was used for biologic experiments.

Female Sprague-Dawley rats weighing 200–250 g were anesthetized with an intramuscular injection of a mixture of ketamine (80 mg kg^{-1}) and xylazine (19 mg kg^{-1}). A jugular vein was surgically exposed, and 100 μL (300–370 kBq) of the solution containing the radioactive complex were injected. The animals ($n = 3$) were sacrificed by cervical dislocation at different times after injection. The blood was withdrawn from the heart through a syringe immediately after the sacrifice, and blood counts were determined. Organs were excised, rinsed in saline, and weighed, and counts were determined with an NaI well counter. Measured

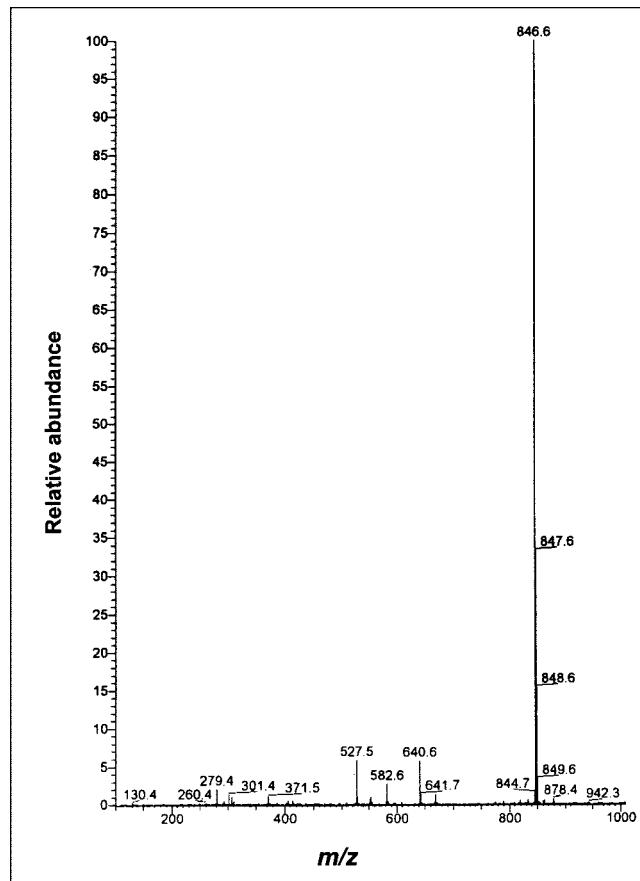


FIGURE 3. ESI mass spectrum of $[^{99\text{g}}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]\text{Cl}$ complex.

TABLE 2
Biodistribution in Rats for Selected $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ Complexes

Organ	% Dose/g for 3 rats at 60 min (mean \pm SD)						
	$[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{NOME})]^+$	$[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{BOET})]^+$	$[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{ISOET})]^+$	$[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{DPODC})]^+$	$[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{NOME})]^+$	$[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{BOET})]^+$	$[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{ISOET})]^+$
Heart	1.31 \pm 0.05	2.84 \pm 0.04	2.98 \pm 0.19	2.26 \pm 0.60	2.05 \pm 0.31	2.69 \pm 0.19	2.74 \pm 0.19
Blood	0.01 \pm 0.00	0.02 \pm 0.00	0.01 \pm 0.00	0.02 \pm 0.01	0.02 \pm 0.00	0.01 \pm 0.00	0.02 \pm 0.00
Lungs	0.21 \pm 0.03	0.40 \pm 0.07	0.42 \pm 0.01	0.27 \pm 0.09	0.29 \pm 0.07	0.40 \pm 0.06	0.35 \pm 0.03
Liver	0.12 \pm 0.02	0.26 \pm 0.03	0.29 \pm 0.07	0.42 \pm 0.23	0.33 \pm 0.05	0.47 \pm 0.09	0.43 \pm 0.15
Kidneys	1.93 \pm 0.27	3.66 \pm 0.50	4.52 \pm 0.51	3.44 \pm 0.76	3.02 \pm 0.40	5.28 \pm 0.58	5.12 \pm 0.35
Heart-to-lung uptake ratio	6.24	7.10	7.10	8.37	7.07	6.73	7.83
Heart-to-liver uptake ratio	10.92	10.92	10.28	5.38	6.21	5.72	6.37

activity was corrected for the decay factor. Results, expressed as percent dose per gram of tissue, are reported in Tables 2–5.

A similar procedure was applied for determination of the biodistribution in rats of the cardiac agents ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin prepared with the corresponding commercial kit formulations and injected without further purification.

Small-Animal Scanner Studies. Tomographic images of the rat heart were obtained by use of a small-animal, hybrid PET-SPECT scanner assembled in our laboratory and described in detail elsewhere (20). Briefly, the YAP-(S)PET scanner has 4 detection heads, each composed of a YAP:Ce planar crystal matrix coupled to a position-sensitive photomultiplier (where YAP is yttrium aluminium Perovskite doped with cerium). The application of high-resolution parallel-hole collimators on 2 opposite detector heads allows this PET device to be used also as a dedicated, small-animal SPECT scanner. The field of view in both PET and SPECT modes is a cylinder with a diameter of 4 cm and a depth of 4 cm. Imaging with ^{99m}Tc complexes was performed 1 h after injection of Sprague-Dawley rats (200–250 g) anesthetized as described above by collection of data with 1 detector head working in the SPECT mode. $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^+$ complexes, ^{99m}Tc -sestamibi, and ^{99m}Tc -tetrofosmin were used in imaging studies, and the injected activities were 174, 184, and 159 MBq, respectively. Acquisition and reconstruction parameters were as follows: angular views, 64 over 180°; recorded events, 192×10^4 ; reconstructed events, $\sim 64 \times 10^4$. Images with dimensions of 40 pixels (radial) \times 40 pixels (axial) \times 20 pixels (transaxial) were reconstructed by means of an expectation maximization iterative algorithm and by excluding events below 140 keV.

RESULTS

Chemistry

The preparation of a class of monocationic, square-pyramidal, ^{99m}Tc -nitride radiopharmaceuticals of the type $[^{99m}\text{Tc}(\text{N})(\text{L})(\text{PNP})]^+$, where L is the monoanionic form of a dithiocarbamate ligand and PNP is a diphosphine ligand (Fig. 2), was performed with a liquid formulation involving a 2-step reaction. The first step produced the $[^{99m}\text{Tc}\equiv\text{N}]^+$ core through the initial reaction of $^{99m}\text{TcO}_4^-$ with SDH as a donor of nitride nitrogen atoms (N^{3-}) and in the presence of Sn^{2+} ions as a reductant (5,21). The preparation was completed by adding simultaneously to the same reaction vial the 2 ligands PNP and NaL to afford the final products, $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$. Notably, the asymmetric $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ heterocomplexes were always obtained without the concomitant formation of the symmetric neutral $[^{99m}\text{Tc}(\text{N})(\text{L})_2]$ complexes, comprising 2 identical bidentate ligands.

A lyophilized, 2-vial kit formulation was specifically developed for the preparation of the compounds $[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{DBODC})]^+$ and $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^+$. The first vial was filled with the reagents SDH and SnCl_2 , required to form the $\text{Tc}\equiv\text{N}$ group, while the second vial contained the sodium salt of DBODC, free PNP3 (or PNP5), and γ -cyclodextrin, used as a solubilizing agent for the diphosphine ligand. The presence of EDTA in

TABLE 3
Biodistribution in Rats of $[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{DBODC})]^+$ Complex

Organ	% Dose/g for 3 rats (mean \pm SD)				
	2 min	10 min	30 min	60 min	120 min
Blood	0.11 \pm 0.01	0.03 \pm 0.00	0.02 \pm 0.00	0.01 \pm 0.00	0.01 \pm 0.00
Heart	3.42 \pm 0.19	3.65 \pm 0.56	3.27 \pm 0.36	3.27 \pm 0.62	3.00 \pm 0.42
Lungs	1.01 \pm 0.49	0.77 \pm 0.10	0.69 \pm 0.12	0.34 \pm 0.11	0.27 \pm 0.05
Liver	1.72 \pm 0.26	1.43 \pm 0.47	0.42 \pm 0.01	0.16 \pm 0.05	0.12 \pm 0.03
Kidneys	11.57 \pm 2.37	6.74 \pm 0.63	5.67 \pm 0.39	4.24 \pm 0.53	3.48 \pm 0.61
Intestine	7.42 \pm 1.03	12.11 \pm 2.92	13.41 \pm 4.62	4.39 \pm 2.86	7.03 \pm 2.61
Muscle	0.23 \pm 0.01	0.23 \pm 0.07	0.24 \pm 0.08	0.17 \pm 0.01	0.36 \pm 0.15

TABLE 4
Biodistribution in Rats of $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^+$ Complex

Organ	% Dose/g for 3 rats (mean \pm SD)				
	2 min	10 min	30 min	60 min	120 min
Blood	0.11 \pm 0.02	0.03 \pm 0.01	0.02 \pm 0.00	0.01 \pm 0.00	0.01 \pm 0.00
Heart	3.94 \pm 0.32	3.69 \pm 0.29	3.73 \pm 0.48	3.76 \pm 0.39	3.31 \pm 0.06
Lungs	0.99 \pm 0.36	0.88 \pm 0.03	0.64 \pm 0.13	0.46 \pm 0.07	0.25 \pm 0.01
Liver	2.66 \pm 0.88	1.61 \pm 0.21	0.72 \pm 0.06	0.20 \pm 0.05	0.10 \pm 0.03
Kidneys	14.69 \pm 2.30	9.16 \pm 1.08	6.70 \pm 0.98	5.73 \pm 0.55	3.48 \pm 0.14
Intestine	7.97 \pm 0.94	9.04 \pm 1.71	6.70 \pm 0.71	6.52 \pm 7.65	6.57 \pm 5.38
Muscle	0.20 \pm 0.04	0.21 \pm 0.05	0.19 \pm 0.04	0.21 \pm 0.03	0.23 \pm 0.06

the first vial of the freeze-dried formulation was required to prevent the precipitation of the neutral disubstituted $\text{Sn}(\text{DBODC})_2$ complexes upon the addition of the dithiocarbamate ligand. With the kit formulation, the yields of $[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{DBODC})]^+$ and $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^+$ achieved were similar to those obtained with the 2-step liquid formulation (95%–98%).

The $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes were characterized by chromatographic methods. Their chemical structures were investigated by comparison with the corresponding analogs prepared with the long-lived isotope ^{99g}Tc . The ESI mass spectrum of the representative $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^+$ complex is shown in Figure 3. The crystal structure of the analogous rhenium complex $[\text{Re}(\text{N})(\text{PNP})(\text{DEDCC})]^+$, where PNP is $\text{Ph}_2\text{P}(\text{CH}_2)_2\text{N}(\text{CH}_2\text{CH}_2\text{OCH}_3)\text{-}(\text{CH}_2)_2\text{PPh}_2$ and DEDC is bis-*N*-ethyl-dithiocarbamato, was reported elsewhere (22).

The $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes were found to be highly stable in solution and in blood, even after removal of excess free ligands. In particular, all of the complexes were strongly resistant to transchelation by cysteine and GSH. HPLC measurement of $\log k_o'$ values showed that these complexes were lipophilic, with $\log k_o'$ values ranging from 2.48 to 4.35 (Table 1).

Animal Studies

Biodistribution data for the $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ heterocomplexes in rats are reported in Tables 2–5. Time–activity curves for heart uptake for some selected heterocomplexes

and for ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin are shown in Figures 4 and 5. Variations in heart-to-lung and heart-to-liver uptake ratios for the $[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{DBODC})]^+$ and $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^+$ complexes, ^{99m}Tc -sestamibi, and ^{99m}Tc -tetrofosmin are shown in Figure 6. Tomographic gamma camera images of the rat myocardium that were collected with the YAP-(S)PET small-animal scanner after injection of the $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^+$ complex and of the commercial radiopharmaceuticals ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin are shown in Figure 7.

All of the $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes accumulated in the rat myocardium, and activity in this region remained almost constant over time. Blood and lung washouts were rapid, elimination occurring mainly through the kidneys and liver. Interestingly, liver uptake was transient, and washout from this organ was fast and quantitative, activity being almost completely eliminated into the intestine at 1 h after injection. Differences in the biologic behaviors of $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes were found to be dependent on modification of the pendant functional groups attached to both the nitrogen atom of the dithiocarbamate ligand and the phosphorus and nitrogen atoms of the diphosphine ligand (Fig. 2). The most important effects of these variations were changes in accumulation in the heart and elimination from nontarget tissues, such as the lungs, liver, and blood. Replacement of the diphosphine ligands PNP3 and PNP5 with PNP6 caused a significant decrease in accumulation in the heart and a concomitant increase in uptake in nontarget organs.

TABLE 5
Biodistribution in Rats of $[^{99m}\text{Tc}(\text{N})(\text{PNP6})(\text{DBODC})]^+$ Complex

Organ	% Dose/g for 3 rats (mean \pm SD)				
	2 min	10 min	30 min	60 min	120 min
Blood	0.17 \pm 0.01	0.06 \pm 0.02	0.05 \pm 0.03	0.02 \pm 0.00	0.02 \pm 0.01
Heart	1.24 \pm 0.14	1.39 \pm 0.30	1.05 \pm 0.19	1.10 \pm 0.11	1.29 \pm 0.16
Lungs	1.25 \pm 0.42	1.10 \pm 0.21	0.60 \pm 0.08	0.48 \pm 0.02	0.33 \pm 0.05
Liver	3.59 \pm 1.82	7.71 \pm 1.05	4.66 \pm 0.56	3.55 \pm 0.64	2.55 \pm 0.69
Kidneys	9.57 \pm 1.93	9.80 \pm 2.06	7.52 \pm 1.16	6.60 \pm 1.29	8.83 \pm 1.19
Intestine	3.54 \pm 0.75	6.21 \pm 0.16	7.75 \pm 2.71	8.37 \pm 3.09	9.70 \pm 2.51
Muscle	0.17 \pm 0.07	0.16 \pm 0.05	0.09 \pm 0.02	0.12 \pm 0.00	0.14 \pm 0.01

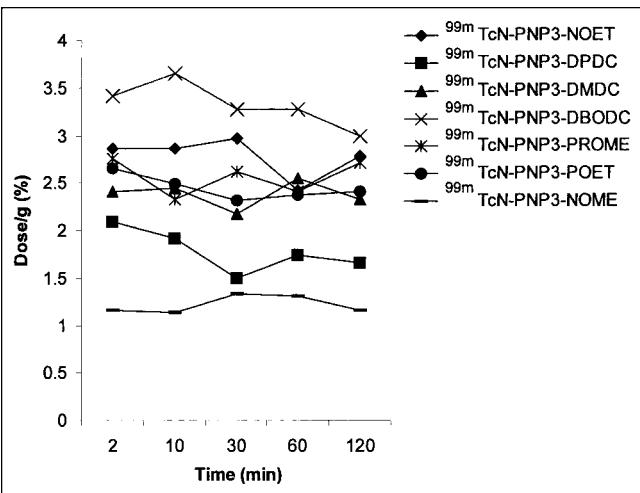


FIGURE 4. Time-activity curves for heart uptake for some selected $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes in rats. SDs are omitted for clarity.

DISCUSSION

The combination of π -donor and π -acceptor properties associated with 2 different bidentate ligands was the key factor in determining the selective formation of asymmetric ^{99m}Tc -nitride heterocomplexes without the concomitant production of the corresponding symmetric compounds containing 2 identical bidentate ligands (12). On the basis of the previous characterization of the molecular structure of the analogous rhenium complex $[\text{Re}(\text{N})(\text{PNP})(\text{DED})]^+$ (22), the distorted square-pyramidal geometry was attributed to the $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^+$ complexes. In this arrangement, the $\text{Tc}\equiv\text{N}$ multiple bond occupies an apical position and

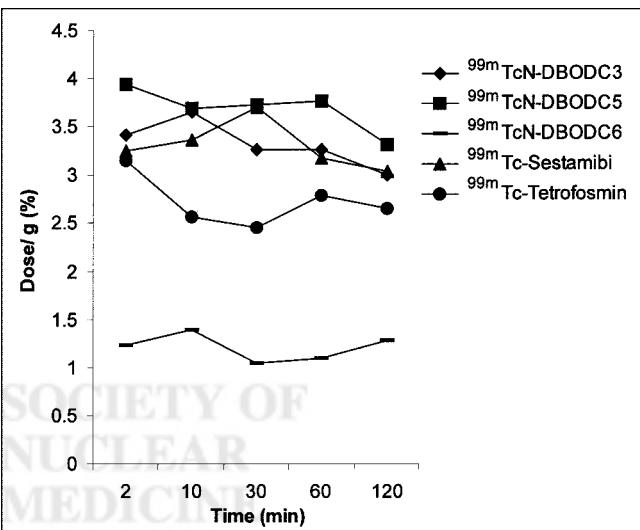


FIGURE 5. Time-activity curves for heart uptake for $[^{99m}\text{Tc}(\text{N})(\text{PNP}3)(\text{DBODC})]^+$, $(^{99m}\text{TcN-DBODC}3)$, $[^{99m}\text{Tc}(\text{N})(\text{PNP}5)(\text{DBODC})]^+$, $(^{99m}\text{TcN-DBODC}5)$, and $[^{99m}\text{Tc}(\text{N})(\text{PNP}6)(\text{DBODC})]^+$, $(^{99m}\text{TcN-DBODC}6)$ complexes, ^{99m}Tc -sestamibi, and ^{99m}Tc -tetrofosmin in rats. SDs are omitted for clarity.

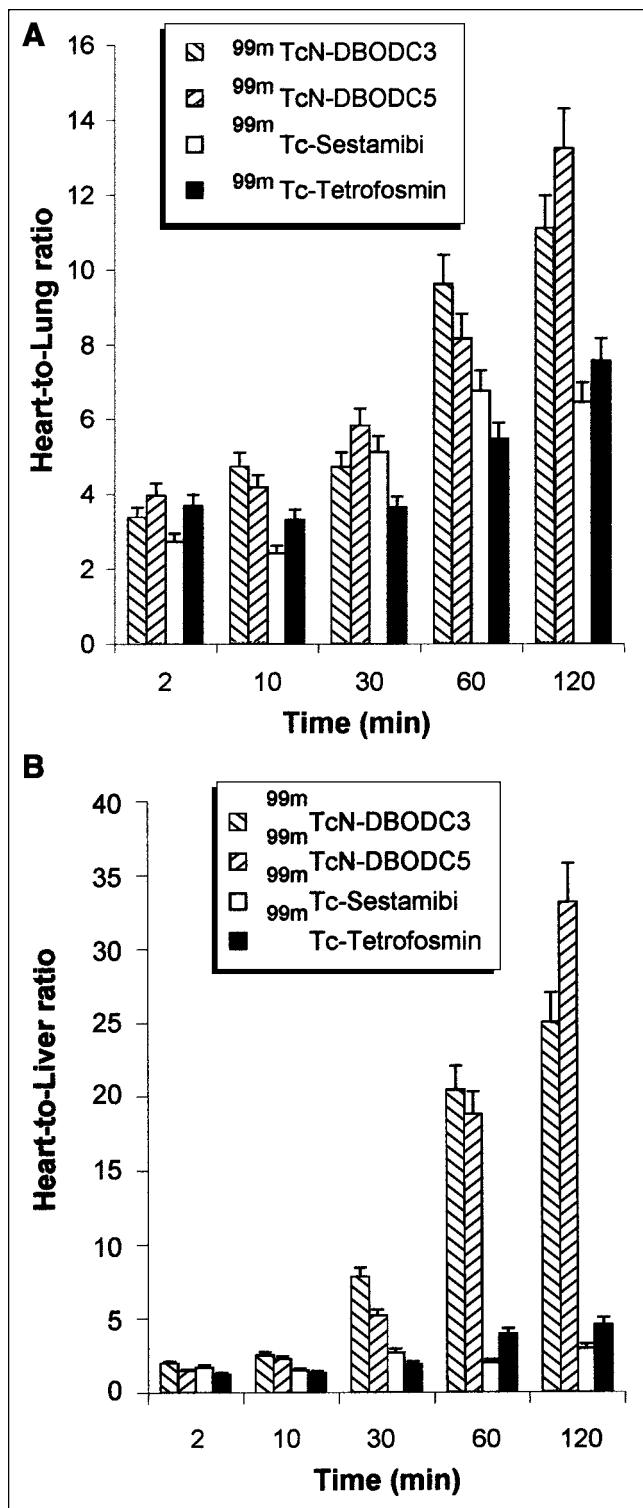


FIGURE 6. Heart-to-lung (A) and heart-to-liver (B) uptake ratios for $[^{99m}\text{Tc}(\text{N})(\text{PNP}3)(\text{DBODC})]^+$, $(^{99m}\text{TcN-DBODC}3)$ and $[^{99m}\text{Tc}(\text{N})(\text{PNP}5)(\text{DBODC})]^+$, $(^{99m}\text{TcN-DBODC}5)$ complexes, ^{99m}Tc -sestamibi, and ^{99m}Tc -tetrofosmin.

the diphosphine and dithiocarbamate ligands span the residual 4 coordination positions on the basal plane through the 2 phosphorus atoms and the 2 sulfur atoms, respectively (Fig. 1).

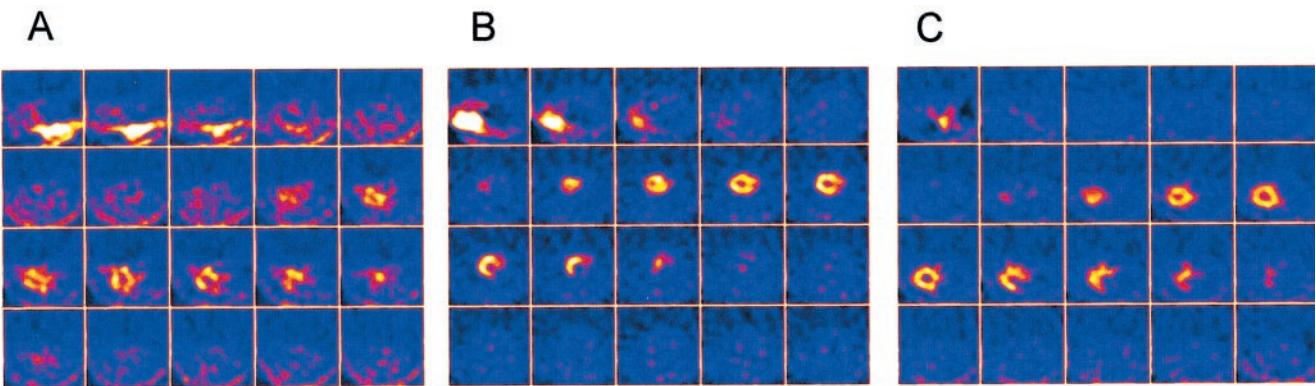


FIGURE 7. Tomographic transaxial images of rat heart obtained with small-animal SPECT scanner (10) 60 min after injection of ^{99m}Tc -sestamibi (A), ^{99m}Tc -tetrofosmin (B), and $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^{+}$ complex (C). Slices are ordered from rat tail (top left) to rat head (bottom right). Color intensity is proportional to pixel activity (counts per second).

The monocationic $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^{+}$ heterocomplexes (where PNP is PNP3 or PNP5) localize selectively in the myocardium of rats and exhibit interesting new properties when viewed in terms of the search for an ideal heart perfusion agent. In fact, although the myocardial uptake of these heterocomplexes was comparable to that of ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin, the $[^{99m}\text{Tc}(\text{N})(\text{PNP})(\text{L})]^{+}$ complexes showed much more favorable heart-to-lung and heart-to-liver uptake ratios. Generally, the heart-to-lung and heart-to-liver uptake ratios for the asymmetric complexes increased exponentially over time and became dramatically high in the interval between 60 and 120 min after injection (Fig. 6). In the series of compounds with PNP3 and PNP5, the most promising biodistribution properties were observed for the $[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{DBODC})]^{+}$ and $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^{+}$ complexes. These findings were interpreted by assuming that the chemical and physical characteristics of these 2 complexes approached a suitable combination of parameters such as size, lipophilicity, and molecular weight, thus resulting in the most favorable biologic behavior. However, substitution of PNP3 (or PNP5) with PNP6, to yield the $[^{99m}\text{Tc}(\text{N})(\text{PNP6})(\text{DBODC})]^{+}$ complex, caused a remarkable decrease in heart uptake and a concomitant increase in liver accumulation, results that might be attributable to the excessive growth of the lipophilic character as a result of the presence of longer alkoxy groups on P atoms of PNP6. These characteristics would cause the values to fall outside the most effective range of biologic parameters.

High-quality images for rats were recorded with a small-animal SPECT scanner assembled in our laboratory (20). Specifically, we attempted a comparison among the ^{99m}Tc -sestamibi, ^{99m}Tc -tetrofosmin, and $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^{+}$ agents. Tomographic images were collected 1 h after injection of the appropriate radiopharmaceutical (Fig. 7). Images were scaled with respect to their own maximum found in the heart region. With the compounds ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin, peak activity was localized in the abdominal region and was associated mainly with higher liver uptake. In contrast, peak activity

with the $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^{+}$ complex was positioned exactly in the myocardial area, and liver activity was found to be almost negligible. As a consequence, with the $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^{+}$ agent, it was possible to obtain a clear visualization of the cardiac region, in close agreement with biodistribution results showing rapid liver clearance. These results support the conclusion that the $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^{+}$ complex appears to possess highly favorable distribution properties in comparison with other cardiac tracers.

CONCLUSION

In this work, a new class of monocationic perfusion heart imaging agents having an unusual asymmetric structure was obtained through the application of improved procedures for the preparation of ^{99m}Tc complexes containing a terminal $\text{Tc}\equiv\text{N}$ multiple bond. These unprecedented structural properties resulted in more favorable biologic behavior. In particular, the $[^{99m}\text{Tc}(\text{N})(\text{PNP3})(\text{DBODC})]^{+}$ and $[^{99m}\text{Tc}(\text{N})(\text{PNP5})(\text{DBODC})]^{+}$ complexes displayed the most appropriate molecular properties for application to heart imaging. Heart-to-lung and heart-to-liver uptake ratios for these compounds were observed to be dramatically higher than those for ^{99m}Tc -sestamibi and ^{99m}Tc -tetrofosmin. These results, in combination with the high heart uptake and fast blood clearance, indicate that the novel complexes possess favorable biologic properties and could be used as improved heart perfusion tracers for collecting high-quality scintigraphic images. Evaluation of the biologic behavior of these new agents in other animal models is currently under way.

ACKNOWLEDGMENTS

The authors thank Prof. Alberto Del Guerra and Prof. Adriano Piffanelli for helpful discussions. Financial support provided by Nihon Medi-Physics Co., Ltd., Tokyo, Japan, is gratefully acknowledged.

REFERENCES

1. Schwaiger M, Melin J. Cardiological applications of nuclear medicine. *Lancet*. 1999;354:661–666.
2. Kailasnath P, Sinusas AJ. Comparison of Tl-201 with Tc-99m-labeled myocardial perfusion agents: technical, physiologic, and clinical issues. *J Nucl Cardiol*. 2001;8:482–498.
3. Banerjee S, Pillai MR, Ramamoorthy N. Evolution of Tc-99m diagnostic radiopharmaceuticals. *Semin Nucl Med*. 2001;3:260–277.
4. Fagret D, Ghezzi C, Vanzetto G. ^{99m}Tc -N-NOET imaging for myocardial perfusion: can it offer more than we already have? *J Nucl Med*. 2001;42:1395–1396.
5. Pasqualini R, Duatti A, Bellande E, et al. Bis(dithiocarbamato) nitrido technetium-99m radiopharmaceuticals: a class of neutral myocardial imaging agents. *J Nucl Med*. 1994;35:334–341.
6. Takehana K, Beller GA, Ruiz M, et al. Assessment of residual coronary stenoses using ^{99m}Tc -N-NOET vasodilator stress imaging to evaluate coronary flow reserve early after coronary reperfusion in a canine model of subendocardial infarction. *J Nucl Med*. 2001;42:1388–1394.
7. Sinusas AJ. Technetium-99m-N-NOET: although not equivalent to thallium-201, it still offers new opportunities. *J Nucl Cardiol*. 2000;7:185–188.
8. Uccelli L, Giganti M, Duatti A, et al. Subcellular distribution of technetium-99m-N-NOEt in rat myocardium. *J Nucl Med*. 1995;36:2075–2079.
9. Llaurado JG. The quest for the perfect myocardial perfusion indicator... still a long way to go. *J Nucl Med*. 2001;42:282–284.
10. Duatti A, Uccelli L. Technetium complexes and radiopharmaceuticals containing the $\text{Tc}\equiv\text{N}$ multiple bond. *Trends Inorg Chem*. 1996;4:27–41.
11. Boschi A, Bolzati C, Benini E, et al. A novel approach to the high-specific-activity labeling of small peptides with the technetium-99m fragment [^{99m}Tc (N)(PXP)] $^{2+}$ (PXP = diphosphine ligand). *Bioconjug Chem*. 2001;12:1035–1042.
12. Bolzati C, Boschi A, Duatti A, et al. Geometrically controlled selective formation of nitrido technetium(V) asymmetrical heterocomplexes with bidentate ligands. *J Am Chem Soc*. 2000;122:4510–4511.
13. Bolzati C, Boschi A, Uccelli L, et al. Chemistry of the strong electrophilic metal fragment [$^{99}\text{Tc}(\text{N})(\text{PXP})]^{2+}$ (PXP = diphosphine ligand): a novel tool for the selective labelling of small molecules. *J Am Chem Soc*. 2002;124:11468–11479.
14. Boschi A, Bolzati C, Uccelli L, et al. A class of asymmetrical nitrido technetium-99m heterocomplexes as heart imaging agents with improved biological properties. *Nucl Med Commun*. 2002;23:689–693.
15. Bianchini C, Casares JA, Peruzzini M, Romerosa A, Zanobini F. The mechanism of Ru-assisted C—C bond cleavage of terminal alkynes by water. *J Am Chem Soc*. 1996;118:4585–4595.
16. Marotta E, Gioacchini AM, Tisato F, Cagnolini A, Uccelli L, Traldi P. Electrospray ionization in the characterization of *mer* and *fac* isomeric forms of [Re(N)Cl₂(POP)] (POP = bis[(2-diphenylphosphino)ethyl]ether). *Rapid Commun Mass Spectrom*. 2001;15:2046–2049.
17. Pietrogrande MC, Bighi C, Borea PA, Barbaro AM, Guerra MC, Biagi GL. Relationship between log K' values of benzodiazepines and composition of the mobile phase. *J Liq Chromatogr*. 1985;8:1711–1729.
18. Baldas J, Bonnyman J, Williams GA. Studies of technetium complexes. 9. Use of the tetrachloronitridotechnetate(VI) anion for the preparation of nitride complexes of technetium: crystal structure of bis(8-quinolinethiolato)nitridotechnetium(V). *Inorg Chem*. 1986;25:150–153.
19. Abram U, Lorenz B, Kaden L, Scheller D. Nitrido complexes of technetium with tertiary phosphines and arsines. *Polyhedron*. 1988;7:285–289.
20. Damiani C, Del Guerra A, Di Domenico G, et al. An integrated PET-SPECT imager for small animals. *Nucl Instrum Methods Phys Res A*. 2001;461:416–419.
21. Bolzati C, Uccelli L, Boschi A, et al. Synthesis of a novel class of nitrido Tc-99m radiopharmaceuticals with phosphino-thiol ligands showing transient heart uptake. *Nucl Med Biol*. 2000;27:369–374.
22. Refosco F, Bolzati C, Duatti A, Tisato F, Uccelli A. Mixed ligand Tc- and Re-nitrido complexes for radiolabeling bioactive molecules. *Recent Results Dev Inorg Chem*. 2000;2:89–98.



SOCIETY OF
NUCLEAR
MEDICINE