

15. Loevinger R, Budinger TF, Watson EE. *MIRD primer for absorbed dose calculations*, Revised. New York: The Society of Nuclear Medicine; 1991:1-22.
16. Goddu SM, Howell RW, Rao DV. Cellular dosimetry: absorbed fractions for monoenergetic electron and alpha-particle sources and S-values for radionuclides uniformly distributed in different cell compartments. *J Nucl Med* 1994;35:303-316.
17. Goddu SM, Howell RW, Buchet LG, Bolch WE, Rao D. MIRD cellular S values: self-absorbed dose per unit cumulated activity for selected radionuclides and monoenergetic electron and alpha particle emitters incorporated into different cell compartments. Reston, VA: The Society of Nuclear Medicine; 1997:2-14.
18. Humm JL. A microdosimetric model of astatine-211 labeled antibodies for radioimmunotherapy. *Int J Radiat Oncol Biol Phys* 1987;13:1767-1773.
19. Humm JL, Chin LM. A model of cell inactivation by alpha-particle internal emitters. *Radiat Res* 1993;134:143-150.
20. Charlton DE, Sephton R. A relationship between microdosimetric spectra and cell survival for high-LET irradiation. *Int J Radiat Biol* 1991;59:447-457.
21. Charlton DE, Kassis AI, Adelstein SJ. A comparison of experimental and calculated survival curves for V79 cells grown as monolayers or in suspension exposed to alpha irradiation from ²¹²Bi distributed in the growth medium. *Radiat Prot Dosim* 1994;52:311-315.
22. Humm JL, Roeske JC, Fisher DR, Chen GTY. Microdosimetric concepts in radioimmunotherapy. *Med Phys* 1993;20:535-541.
23. Roesch WC. Microdosimetry of internal sources. *Radiat Res* 1977;70:494-510.
24. Fisher DR. The microdosimetry of monoclonal antibodies labeled with alpha emitters. In: Schlafke-Stelson AT, Watson EE, eds. *Proceedings of the fourth international radiopharmaceutical dosimetry symposium*. Oak Ridge, TN: Oak Ridge Associated Universities; 1986:26-36.
25. Fisher DR, Harty R. The microdosimetry of lymphocytes irradiated by alpha particles. *Int J Radiat Biol* 1982;41:315-324.
26. Stinchcomb TG, Roeske JC. Analytic microdosimetry for radioimmunotherapeutic alpha emitters. *Med Phys* 1992;19:1385-1393.
27. Stinchcomb TG, Roeske JC. Survival of alpha-particle irradiated cells as a function of the shape and size of the sensitive volume (nucleus). *Radiat Prot Dosim* 1995;62:157-164.
28. Kellerer AM. Analysis of patterns of energy deposition; a survey of theoretical relations in microdosimetry. In: Ebert HG, ed. *Proceedings of the second symposium on microdosimetry*. Brussels: Commission of European Communities; 1970:107-134.
29. Roesch WC. Moments of microdosimetric quantities for particulate sources. *Radiat Res* 1985;102:392-398.
30. Caswell RS. Deposition of energy by neutrons in spherical cavities. *Radiat Res* 1996;27:92-107.
31. Kellerer AM, Chmelevsky D. Criteria for the applicability of LET. *Radiat Res* 1975;63:226-234.
32. Azure MT, Archer RD, Sastry KSR, Rao DV, Howell RW. Biological effects of lead-212 localized in the nucleus of mammalian cells: role of recoil energy in the radiotoxicity of internal alpha-particle emitters. *Radiat Res* 1994;140:276-283.
33. Howell RW, Rao DV, Hou D-Y, Narra VR, Sastry KSR. The question of relative biological effectiveness and quality factor for Auger emitters incorporated into proliferating mammalian cells. *Radiat Res* 1991;128:282-292.
34. ICRU. *Stopping powers, ranges for protons, and alpha particles*, International Commission of Radiation Units and Measurements Report No. 49. Bethesda, MD: ICRU; 1993:256.
35. Eckerman KF, Ryman JC, Taner AC, Kerr GD. Traversals of cells by radiation and absorbed fraction estimates for electrons and alpha particles. In: A. T. Schlafke-Stelson and E. E. Watson, eds. *Proceedings of the fourth international radiopharmaceutical dosimetry symposium*. Oak Ridge, TN: Oak Ridge Associated Universities; 1986:67-86.
36. ICRU. *Microdosimetry*, International Commission of Radiation Units and Measurements Report No. 36. Bethesda, MD: ICRU; 1983:54-55.

D-Lysine Reduction of Indium-111 Octreotide and Yttrium-90 Octreotide Renal Uptake

Bert F. Bernard, Eric P. Krenning, Wout A.P. Breeman, Edgar J. Rolleman, Willem H. Bakker, Theo J. Visser, Helmut Mäcke and Marion de Jong

Departments of Nuclear Medicine and Internal Medicine III, Erasmus Medical University and Academic Hospital Dijkzigt, Rotterdam, The Netherlands; and Department of Nuclear Medicine, Kantonspital Basel, Basel, Switzerland

Indium-111-DTPA-octreotide (¹¹¹In-DTPAOC) is used successfully for imaging somatostatin receptor-positive lesions. A new and promising application is its use in peptide-receptor radionuclide therapy (PRRT). For the latter purpose, [DOTA⁰,D-Phe¹,Tyr³]octreotide (DOTATOC), which is suitable for stable radiolabeling with ⁹⁰Y, is probably even more promising. Significant renal uptake of these octreotide analogs exists, however, reducing the scintigraphic sensitivity for detection of small tumors in the perirenal region and limiting the possibilities for PRRT. We showed that the renal uptake of ¹¹¹In-DTPAOC could be reduced to about 50% of control by L-lysine administration in vivo in rats. This study compares the effects of several doses and different methods of administration of D- and L-lysine, in addition to time-related effects of D-lysine, on kidney uptake of ¹¹¹In-DTPAOC and ⁹⁰Y-DOTATOC. **Methods:** Male Wistar rats (200-250 g) were given ¹¹¹In-DTPAOC (0.2 MBq, 0.5 μg-0.5 mg intravenously, intraperitoneally or orally) in the presence or absence of D- or L-lysine. At 1, 4 or 24 hr, the rats were killed, and the organs were isolated and counted for radioactivity. In different experiments, male Wistar rats (200-250 g) were given ⁹⁰Y-DOTATOC (1 MBq, 0.5 μg intravenously) in the presence or absence of D-lysine. At 24 hr, the rats were killed, and the organs were isolated and counted for radioactivity. **Results:** Administration of D- or L-lysine in a single intravenous dose of 400 mg/kg, resulted in more than 50% inhibition of kidney uptake of ¹¹¹In-DTPAOC at all time points tested, independently of the mass of ¹¹¹In-DTPAOC used. Higher or repeated doses of lysine did not give a significantly higher percentage inhibition. D-lysine, given orally in a dose of 400 mg/kg at 30 or 15 min before ¹¹¹In-DTPAOC injection, resulted in

30% and 20% inhibition of kidney uptake, respectively. L-lysine, given orally 30 min before ¹¹¹In-DTPAOC administration, resulted in 30% inhibition as well. Inhibition of kidney uptake of ¹¹¹In-DTPAOC by L-lysine after intraperitoneal administration was 40%. After L-lysine administration, ¹¹¹In-DTPAOC was decreased in the kidneys and in somatostatin receptor-positive organs such as the pancreas and adrenal glands. In contrast, D-lysine did not have a significant effect on uptake in octreotide receptor-positive organs. Renal uptake of ⁹⁰Y-DOTATOC was reduced by 65% by intravenous D-lysine, whereas radioactivity in blood, pancreas and adrenal glands was not affected. **Conclusion:** D-lysine may be preferred to L-lysine for reduction of renal uptake of radioactivity during scintigraphy and PRRT because of its lower toxicity and because it should not interfere with the natural amino acid metabolic balance.

Key Words: indium-111-octreotide; yttrium-90-octreotide; renal tubular reuptake; D-lysine; L-lysine; peptide-receptor radiotherapy

J Nucl Med 1997; 38:1929-1933

Indium-111 pentetreotide is a radiopharmaceutical that binds to somatostatin receptors present in certain tissues. It is being used for scintigraphic imaging of somatostatin receptor-positive lesions such as gastrointestinal pancreatic tumors, neuroblastoma, pheochromocytoma, breast cancer, Hodgkin's lymphoma and small-cell lung cancer (1-3). A new, interesting area of application is PRRT. Promising results have been reported in humans (4). However, although it emits Auger electrons, ¹¹¹In is not an optimal nuclide for radiotherapy. A beta-particle-emitting isotope such as ⁹⁰Y (maximum beta energy 2.3 MeV, half-life 64 hr) is more suitable for this purpose. However, [⁹⁰Y-DTPA⁰]octreotide is not stable resulting in hematopoietic toxicity in vivo as ⁹⁰Y escapes to the skeleton (5,6). We have

Received Aug. 5, 1996; accepted Dec. 10, 1996.

For correspondence or reprints contact: Marion de Jong, PhD, Department of Nuclear Medicine, Academic Hospital Rotterdam, 3015 GD Rotterdam, The Netherlands.

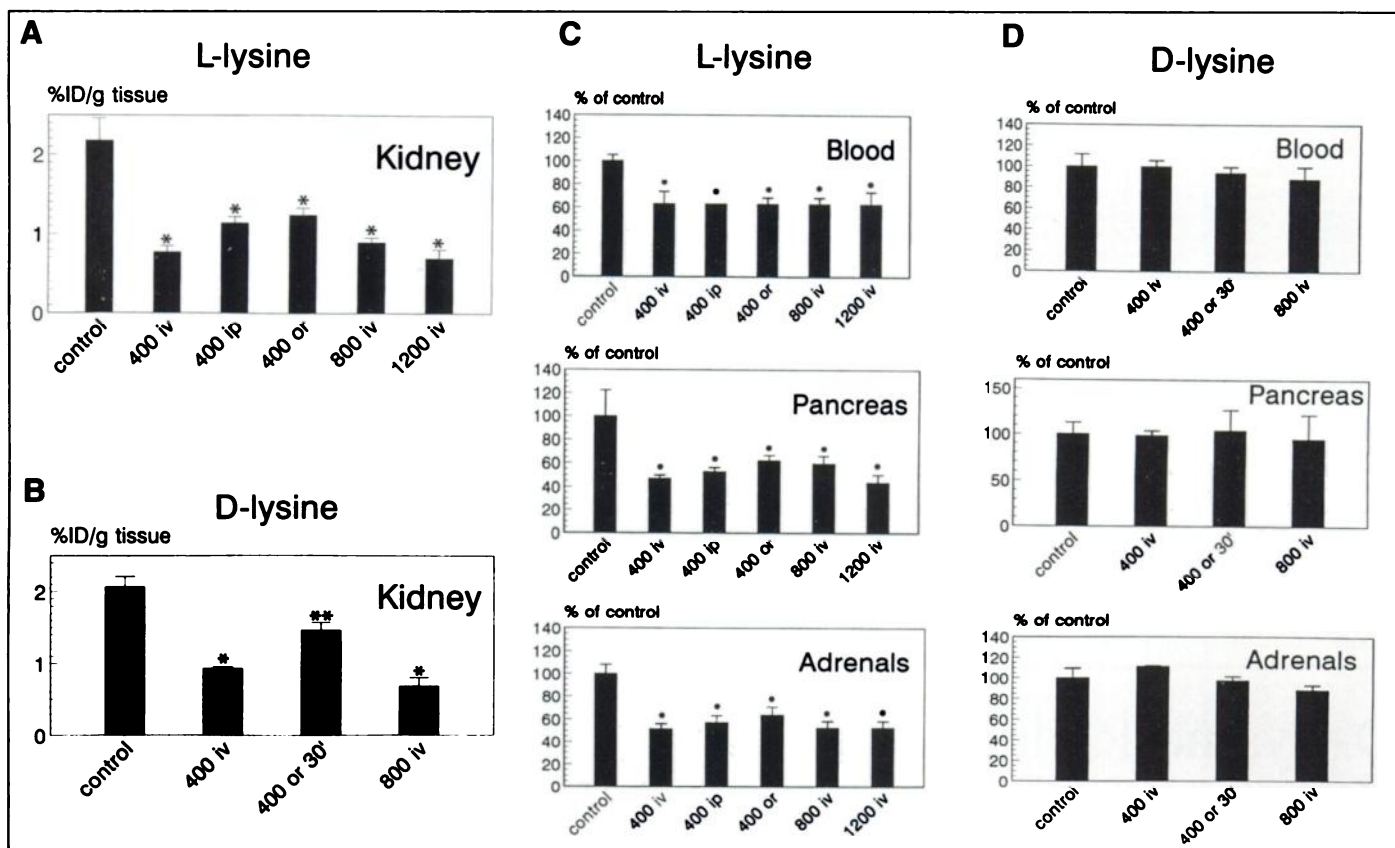


FIGURE 1. (A) Radioactivity in kidneys of control and L-lysine-treated rats, 24 hr after administration of ^{111}In -DTPA-octreotide (0.2 MBq and 0.5 μg). Mean \pm 1 s.d. * $p < 0.001$ versus control. (B) Radioactivity in kidneys of control and D-lysine-treated rats, 24 hr after administration of ^{111}In -DTPA-octreotide (0.2 MBq and 0.5 μg). Mean \pm 1 s.d. * $p < 0.001$ versus control. ** $p < 0.01$ versus control. (C) Radioactivity in several organs of control and L-lysine-treated rats, 24 hr after administration of ^{111}In -DTPA-octreotide (0.2 MBq and 0.5 μg). Mean \pm 1 s.d. * $p < 0.05$ versus control. (D) Radioactivity in several organs of control and D-lysine-treated rats, 24 hr after administration of ^{111}In -DTPA-octreotide (0.2 MBq and 0.5 μg octreotide). Mean \pm 1 s.d. iv = intravenous; ip = intraperitoneal; or = oral.

studied the binding of DOTATOC, suitable for stable radiolabeling with ^{90}Y , to octreotide receptors and compared the biodistribution of ^{111}In - and ^{90}Y -labeled DOTATOC with that of ^{111}In -DTPAOC in rats and found that uptake of radiolabeled DOTATOC in the octreotide receptor-expressing tissues pancreas, pituitary, adrenal glands and tumor was a factor of 3–16 that, after injection of ^{111}In -DTPAOC, makes it a promising pharmaceutical for PRRT of patients with octreotide receptor-positive lesions (7).

Indium-111-DTPAOC and ^{90}Y -DOTATOC are mostly cleared from the body by the kidneys, 50% within the first 4 hr after injection. However, a significant amount of the dose accumulates in the renal parenchyma reducing the scintigraphic sensitivity for detection of small tumors in the perirenal region in the abdomen. The possibilities of PRRT are reduced as well, as rapid excretion of nontumor-bound radioactivity is necessary to realize this potential.

It has been reported repeatedly that renal accumulation of peptides or proteins can be reduced. Infusion of certain amino acids, particularly lysine and arginine, has been shown to block renal tubular peptide reabsorption in general (8). An infusion of synthetic amino acids, containing among others lysine and arginine, significantly reduced kidney uptake of ^{111}In -DTPAOC in eight patients (9). Also, in mice, reduction of renal tubular reabsorption of ^{111}In -labeled Fab-fragment was affected by systemic administration of lysine (10). We reported on the inhibiting effects of intravenous administration of 400 mg/kg L-lysine on the kidney uptake of ^{111}In - or ^{161}Tb -labeled DTPAOC in-vivo in rats (11,12). Behr et al. recently showed that a variety of basic compounds was capable of inhibiting the

tubular reabsorption, thus lowering the kidney uptake of antibody fragments significantly (13). In patients, renal uptake of monoclonal antibody fragments could be reduced significantly by amino acid infusion (14).

Building on the latter studies with antibody fragments, we compared the effects of different doses of D- and L-lysine on kidney uptake of ^{111}In -DTPAOC and ^{90}Y -DOTATOC and the influence on uptake in somatostatin receptor-positive organs. We also examined the effect of different methods of administration (oral, intravenous and intraperitoneal administration) of lysine and the time dependence of its effects on renal uptake of different masses of ^{111}In -DTPAOC.

MATERIALS AND METHODS

Radiolabeling and Quality Control of the Radiopharmaceuticals

The radiolabeling procedures were performed as described by De Jong et al. (7) and Bakker et al. (15).

Tissue Distribution of Indium-111-DTPAOC

Male Wistar rats (200–250 g) were placed in metabolic cages 24 hr before the start of the experiment. In experiments with fasting rats, food was withdrawn for 24 hr before the start of the experiment. Drinking water was always available ad libitum.

Effect of Peptide Mass

At time $t = 0$, rats were anesthetized with ether and injected intravenously with 0.2 MBq and 0.5 μg or 0.5 mg ^{111}In -DTPAOC, with or without co-injection of D- or L-lysine (see below).

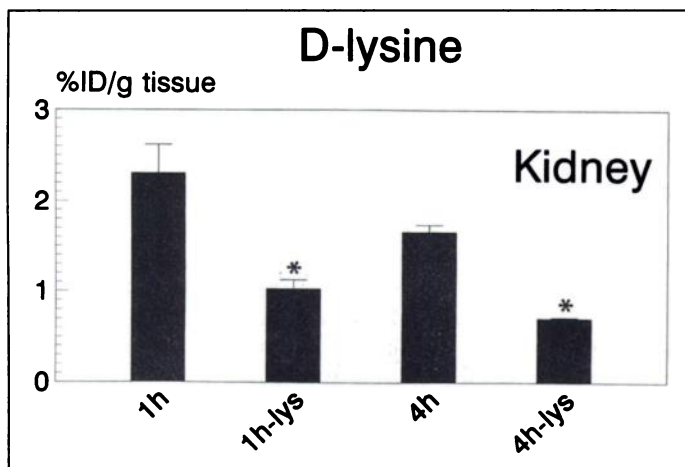


FIGURE 2. Radioactivity in kidneys of control and D-lysine-treated rats, 1 or 4 hr after administration of ^{111}In -DTPA-octreotide (0.2 MBq and 0.5 μg). Mean \pm 1 s.d. * $p < 0.001$ versus control.

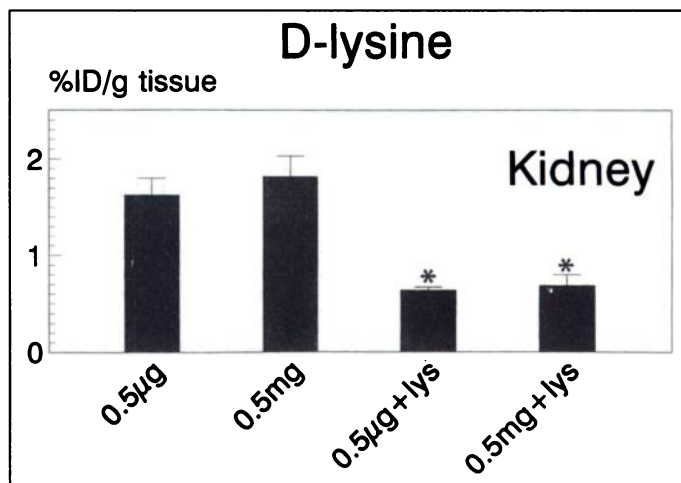


FIGURE 3. Radioactivity in kidneys of control and D-lysine-treated rats, 24 hr after administration of 0.5 μg or 0.5 mg ^{111}In -DTPA-octreotide (0.2 MBq). Mean \pm 1 s.d. * $p < 0.001$ versus control.

Time Dependence of Lysine Effects

Rats were killed and organs were isolated 1, 4 or 24 hr after injection of the radiolabeled product and lysine.

D- and L-lysine Administration Methods and Doses

D- and L-lysine were given in doses of 400, 800 or 1200 mg/kg, intravenously or intraperitoneally. Administration consisted of single injections of these doses or repeated injections of 400 mg/kg at the time points indicated in the Results section. Oral administration of 400 mg/kg D- or L-lysine was performed through an oral intubation catheter at time points indicated in the Results section.

Tissue Distribution of Yttrium-90-DOTATOC

Experiments were performed essentially the same as described for ^{111}In -DTPAOC: Wistar rats were injected under ether anesthesia with 1 MBq (0.5 μg) ^{90}Y -DOTATOC intravenously. For reduction of kidney uptake, 400 mg/kg D-lysine was co-injected with the radiopharmaceutical.

Tissue distribution of both radiopharmaceuticals was studied by measurement of radioactivity (in the case of ^{90}Y as bremsstrahlung) in isolated organs, as well as in blood samples, using an LKB-1282-Compu-gamma system.

Statistical evaluation was performed using one-way analysis of variance followed by comparison among class means and Student's t-test corrected for multiple pairwise comparisons between means. Results are expressed as mean \pm 1 s.d.; in each group $n \geq 6$.

RESULTS

The inhibition of renal uptake of ^{111}In -DTPAOC by L-lysine, 24 hr after administration of both, is shown in Figure 1A. The effects of different doses and administration pathways are visualized also. Single doses of 400, 800 or 1200 mg/kg L-lysine, intravenously or intraperitoneally, were co-injected with ^{111}In -DTPAOC; 2×400 mg/kg L-lysine intravenously was given at 30 min before and 15 min after administration of ^{111}In -DTPAOC; 3×400 mg/kg L-lysine intravenously was given at 30 min before and 15 and 60 min after administration of ^{111}In -DTPAOC. Oral administration of 400 mg/kg L-lysine was 30 min before ^{111}In -DTPAOC. Repeated or higher doses than a single intravenous injection of 400 mg/kg L-lysine did not result in significantly greater inhibition of ^{111}In -DTPAOC kidney uptake. Oral and intraperitoneal administration of L-lysine also had a significant inhibitory effect on kidney uptake. These effects were smaller than those after intravenous administration, albeit not significantly.

In Figure 1B, the inhibition exerted by D-lysine on renal uptake of ^{111}In -DTPAOC, 24 hr after administration, is shown.

Doses and timing of D-lysine administration were mostly the same as for L-lysine. Oral doses were given at 30 min prior to ^{111}In -DTPAOC. Again, in accordance with the findings for L-lysine, repeated or higher doses than a single intravenous injection of 400 mg/kg D-lysine did not result in significantly greater inhibition of kidney uptake of ^{111}In -DTPAOC. Oral administration of D-lysine also reduced kidney uptake; however, this effect was significantly smaller than after intravenous administration ($p < 0.05$). The effects of 400 mg/kg D- and L-lysine administered intravenously on kidney uptake of ^{111}In -DTPAOC appeared to be similar.

In Figure 1C, the distribution of radioactivity, expressed as percent of control, 24 hr after administration of ^{111}In -DTPAOC, is given for blood, pancreas and adrenal glands of control and L-lysine-treated rats. Radioactivity in control animals was $0.0017 \pm 0.0003\%$ of the injected dose (ID)/g, $1.12\% \pm 0.35\%$ ID/g and $1.54\% \pm 0.22\%$ ID/g for blood, pancreas and adrenal glands, respectively. Indium-111-DTPAOC clearance from the blood was faster in L-lysine-treated rats, and uptake in the somatostatin receptor-positive organs pancreas and adrenal glands was less than that in control animals.

In Figure 1D, the distribution of radioactivity expressed as percent of control, 24 hr after administration of ^{111}In -DTPAOC, is given for blood, pancreas and adrenal glands of control and D-lysine-treated rats. No effect of D-lysine was found on both ^{111}In -DTPAOC clearance from the blood and uptake in the receptor-positive organs. The ratios pancreas/kidney (both expressed as %ID/g) and adrenals/kidney of ^{111}In -DTPAOC uptake appeared to be 0.8 and 1.2, respectively, after L-lysine treatment, whereas after D-lysine administration these ratios were 1.4 and 1.9, respectively. For the control rats, the figures

TABLE 1
Radioactivity in Organs and Blood of Rats 24 Hr after Administration of [^{90}Y -DOTA 0 , Tyr 3]octreotide without or with Co-injection of 400 mg/kg D-lysine

Organ	Radioactivity (% of control)*
Blood	100.0 \pm 0.0
Kidneys	34.8 \pm 5.2 [†]
Pancreas	111.0 \pm 5.4
Adrenal glands	97.0 \pm 6.5

*Data from D-lysine-treated rats are expressed as % of those in controls.

[†] $p < 0.01$ vs. control.

were 0.5 and 0.7 for pancreas/kidney and adrenals/kidney, respectively.

Figure 2 shows the reduction of ^{111}In -DTPAOC kidney uptake by D-lysine 1 or 4 hr after administration. It shows that the reducing effect of lysine on kidney uptake of ^{111}In -DTPAOC is comparable at the indicated time points. Furthermore, no significant effect was found on the ^{111}In -DTPAOC clearance from blood and uptake in the receptor-positive organs (not shown).

Figure 3 shows that 400 mg/kg D-lysine reduced kidney uptake of 0.5 mg ^{111}In -DTPAOC, which is very high, even blocking the dose for uptake of radioactivity in octreotide receptor-positive organs.

Table 1 shows the distribution of radioactivity in rats 24 hr after administration of ^{90}Y -DOTATOC without or with co-injection of 400 mg/kg D-lysine. Renal uptake was reduced to 35% of control by D-lysine, whereas radioactivity in blood, pancreas and adrenal glands was not affected.

DISCUSSION

Peptides (and proteins less than 60 kD) in plasma are filtered through the glomerular capillaries in the kidneys and subsequently reabsorbed almost completely ($\geq 99\%$) by the proximal tubular cells through saturable receptor-mediated endocytosis. The peptide then may be intracellularly routed to the lysosomes where degradation takes place. Lysosomal degradation also has been described for ^{111}In -DTPA-labeled peptides, its radiolabeled degradation products are retained in the lysosomes and likely transferred to intracellular metalloproteins (16).

It has been shown repeatedly that renal accumulation of peptides or proteins can be reduced. Administration of amino acids, among others lysine and arginine, has been shown to block renal tubular peptide or protein reabsorption (8–15). It was concluded that the mechanism of the reduction of the renal uptake of radiolabeled peptides seemed to rely on an inhibition of the tubular reabsorption so that they appear directly in the urine without prior lysosomal degradation to low-molecular weight compounds (14). Membranes of renal tubular cells contain negatively charged sites, to which positively charged amine or guanidine groups of peptides can bind (8). So, decreased binding of ^{111}In -DTPAOC after administration of the positively charged amino acids lysine and arginine can be explained by this phenomenon (9).

An important finding of the experiments described here is that L- and D-lysine were equally potent in inhibiting kidney uptake of ^{111}In -DTPAOC, but that after L-lysine administration the uptake in the somatostatin receptor-positive organs was reduced as well. This reduction of uptake of ^{111}In -DTPAOC in the receptor-positive organs was not found after administration of D-lysine. These findings in rats may have implications for the clinical use of lysine as an inhibitor of kidney uptake of ^{111}In -DTPAOC. For both diagnostic and radiotherapeutical use of ^{111}In -DTPAOC, it is important that a maximal uptake of radioactivity in the tumor is achieved. Therefore, D-lysine is the preferred agent for inhibition of kidney uptake of ^{111}In -DTPAOC. Another advantage of D-lysine is that toxicity of lysine at high doses seems to be restricted to the L-isomer (LD_{50} in fasted rats 4000 mg/kg (17)) and that D-lysine should not interfere with the natural amino acid metabolic balance as D-lysine is not used as a source for L-lysine in humans (18).

An increase in glomerular filtration rate (GFR) and renal plasma flow (RPF) after administration of L-lysine, but not after D-lysine, could at least partially explain our findings. The measured radioactivity in the blood, 24 hr after administration of ^{111}In -DTPAOC, was significantly lower after L-lysine ad-

ministration than in control rats ($p < 0.05$). It may be hypothesized from these experiments that a lower blood concentration, because of a higher renal clearance, results in a lower uptake of ^{111}In -DTPAOC in the organs, except for the kidneys. This is in accordance with our opposite findings with regard to increased uptake of ^{111}In -DTPAOC in all organs, except for the kidneys, after sodium maleate administration, explained by the inhibitory effect of this compound on the GFR (11).

Our findings of reduced radioactivity in receptor-positive organs after L-lysine administration are in contrast with those of Behr et al. (13) and Pimm et al. (10) who did not find a significant effect of L-lysine on the uptake and retention of radiolabeled antibody fragments in tumor or normal organs in mice. This discrepancy may be due to species differences (mice versus rats), or by the fact that their experiments were performed with radiolabeled monoclonal antibody fragments, whereas ours were done with radiolabeled peptides. Peptides are cleared faster from the circulation and receptor binding and uptake processes occur at higher rates than those of the bigger monoclonal antibody fragments. In the period just after injection of ^{111}In -DTPAOC, lysine concentration in the circulation is relatively high and may hamper ^{111}In -DTPAOC binding to its receptors by the same mechanism as described for the kidneys (see above). When binding of monoclonal antibody fragments to their receptors occurs, lysine concentration in the blood decreases and may not influence the binding process. Furthermore, an increased GFR by L-lysine will affect the rapid ^{111}In -DTPAOC clearance more than that of monoclonal antibody fragments.

From our data, we conclude that the maximal inhibitory effect of both L- and D-lysine is reached after intravenous administration of 400 mg/kg as higher doses or repeated doses of 400 mg/kg do not result in an increased reduction of ^{111}In -DTPAOC uptake in the kidneys. After intraperitoneal administration, lysine was less effective than after intravenous injection. The same holds for a single oral administration. Overnight fasting, thereby emptying the stomachs of the rats, did not improve the effect of orally administered D-lysine on kidney uptake (results not shown). It may be useful to further investigate the effects of oral administration of higher doses of D-lysine as this means of administration avoids long intravenous infusion of lysine in humans. The different time point measurements showed that D-lysine had the same reducing effect on kidney dose measured at 1, 4 and 24 hr after administration of ^{111}In -DTPAOC.

Currently, PRRT of octreotide receptor-positive lesions is explored by repeated administration of high doses of radiolabeled octreotide. Promising results have been reported both in human and in rat studies (4,19). Stolz et al. (19) found up to 100% longer survival of tumor-bearing rats after radionuclide therapy with [^{90}Y -DTPA-benzyl-acetamido]-octreotide. We investigated in vivo in rats the effect of PRRT, using ^{111}In -DTPAOC, on the development and growth of somatostatin receptor-positive tumors (CA20948) inoculated in the liver. Treatment was given at Day 1 and/or Day 8 after inoculation with: (a) 0.5 μg unlabeled DTPA-octreotide, (b) 10 mCi ^{111}In -DTPAOC (0.5 μg) or (c) 10 mCi ^{111}In -DTPAOC after saturation of the receptors with 1 mg unlabeled octreotide. Significantly fewer tumors were found in animals of Group B compared to the other groups (manuscript submitted for publication). As for PRRT in humans, we treated 11 end-stage patients with neuroendocrine tumors with ^{111}In -DTPAOC, up to a cumulative dose of 53 GBq (for dose estimates (20)) per patient, in a Phase I trial (4). There were no major side effects after up to 2 yr of treatment and positive effects were found on

clinical symptoms, hormone production and tumor proliferation (4). However, ^{111}In is not the most appropriate radionuclide for PRRT; it lacks the preferable higher energies of beta particles. Yttrium-90 is a good candidate with its maximum beta energy of 2.3 MeV and high affinity for the DOTA chelator. Radiotherapeutic use of ^{90}Y -DOTATOC will lead to a higher and more evenly distributed radiation dose to the tumor because of its larger particle range and tissue penetration. With respect to in vivo tissue distribution in the rat, we showed that specific uptake of both ^{90}Y - and ^{111}In -labeled DOTATOC in octreotide receptor-expressing tissues was significantly higher than that of ^{111}In -DTPAOC, making radiolabeled DOTATOC favorable for both scintigraphy and radiotherapy of receptor-positive lesions (7).

D-lysine administration resulted in a significant reduction of labeled DOTATOC uptake in the kidneys without affecting uptake in receptor-positive tissues, thus bringing the application of the labeled compound for radionuclide therapy further within reach. In this respect, it is also interesting to note that the D-lysine dose (400 mg/kg) used in rats was able to reduce kidney uptake of up to 0.5 mg ^{111}In -DTPAOC, which is a very high and blocking dose of octreotide for uptake in the octreotide receptor-positive organs.

More knowledge of ways to decrease renal uptake of radiometals is of great value for tumor scintigraphy and PRRT as this may lead to better protection of the kidneys and, thus, less radiotoxicity in this organ, especially after PRRT with high-energy beta-emitters, such as ^{90}Y , coupled to octreotide analogs.

CONCLUSION

While this article was in press, further experiments in CA20948 tumor-bearing rats showed that the tumor uptake after ^{111}In -DTPAOC injection was not affected by 400 mg/kg L-lysine. We still conclude, however, that D-lysine may be preferred to L-lysine for reduction of renal uptake of radioactivity during scintigraphy and PRRT because of its lower toxicity and because it should not interfere with the natural amino acid metabolic balance.

REFERENCES

- Krenning EP, Kwekkeboom DJ, Bakker WH, et al. Somatostatin receptor scintigraphy with [^{111}In -DTPA-D-Phe 1]- and [^{123}I -Tyr 3]-octreotide: the Rotterdam experience with more than 1000 patients. *Eur J Nucl Med* 1993;20:716-731.
- Krenning EP, Kwekkeboom DJ, Pauwels S, Kvols LK, Reubi JC. Somatostatin receptor scintigraphy. In: Freeman LM, ed. *Nuclear medicine annual*. New York: Raven Press; 1995:1-50.
- Krenning EP, Bakker WH, Kooij PP, et al. Somatostatin receptor scintigraphy with indium-111-DTPA-D-Phe-1-octreotide in man: metabolism, dosimetry and comparison with iodine-123-Tyr-3-octreotide. *J Nucl Med* 1992;33:652-658.
- Krenning EP, Kooij PPM, Pauwels S, et al. Somatostatin receptor: scintigraphy and radionuclide therapy. *Digestion* 1996;57:57-61.
- Rowlinson G, Snook D, Stewart S, Epenetos AA. Intravenous EDTA to reduce bone uptake of Y-90 following Y-90 labeled antibody administration. *Br J Cancer* 1989;59:322.
- Jowsey J, Rowland RE, Marshall JH. The deposition of the rare earths in bone. *Radiation Res* 1958;8:490-501.
- De Jong M, Bakker WH, Krenning EP, et al. ^{90}Y and ^{111}In labeling, receptor binding and biodistribution of [DOTA 0 ,D-Phe 1 ,Tyr 3]octreotide, a promising somatostatin analog for radionuclide therapy. *Eur J Nucl Med* 1997;24:368-371.
- Mogensen CE, Solling K. Studies on renal tubular protein reabsorption: partial and near complete inhibition by certain amino acids. *Scan J Clin Lab Invest* 1977;37:477-486.
- Hammond PJ, Wade AF, Gwilliam ME, et al. Amino acid infusion blocks renal tubular uptake of an indium-labeled somatostatin analog. *Br J Cancer* 1993;67:1437-1439.
- Pimm MV, Gribben SJ. Prevention of renal tubule reabsorption of radiometal (indium-111) labeled Fab fragment of a monoclonal antibody in mice by systemic administration of lysine. *Eur J Nucl Med* 1994;21:663-665.
- De Jong M, Bernard BF, Rolleman EJ, et al. Inhibition of renal re-uptake of [^{111}In -DTPA-D-Phe 1]-octreotide in vivo. *J Nucl Med* 1996;23:1361-1366.
- De Jong M, Breeman WAP, Bernard BF, et al. [^{111}In -DTPA-D-Phe 1]octreotide preparation, in vitro receptor binding and biological activity, metabolism in isolated perfused rat livers and distribution in vivo in normal and tumor-bearing rats in comparison with [^{111}In -DTPA-D-Phe 1]octreotide. *Eur J Nucl Med* 1995;22:608-616.
- Behr TM, Sharkey RM, Juweid ME, et al. Reduction of the renal uptake of radiolabeled monoclonal antibody fragments by cationic amino acids and their derivatives. *Cancer Res* 1995;55:3825-3834.
- Behr TM, Becker WS, Sharkey RM, et al. Reduction of renal uptake of monoclonal antibody fragments by amino acid infusion. *J Nucl Med* 1996;37:829-833.
- Bakker WH, Albert R, Bruns C, et al. [^{111}In -DTPA-D-Phe 1]octreotide, a potential radiopharmaceutical for imaging of somatostatin receptor-positive tumors: Synthesis, radiolabeling and in vitro validation. *Life Sci* 1991;49:1583-1591.
- Duncan JR, Welch MJ. Intracellular metabolism of indium-111-DTPA-labeled receptor-targeted proteins. *J Nucl Med* 1993;34:1728-1738.
- Gullino P, Winitz M, Birnbaum SM, Clyde Otey M, Cornfield J, Greenstein JP. The toxicity of individual essential amino acids and their diastereomers in rats and the effect on blood-sugar levels. *Arch Biochem Biophys* 1955;58:253-255.
- Friedman M. Formation, nutritional value and safety of D-amino acids. In: Friedman M, ed. *Nutritional and toxicological consequences of food processing*. New York: Plenum Press; 1991:447-481.
- Stolz B, Smith-Jones PM, Albert R, et al. Somatostatin analogs for somatostatin-receptor-mediated radiotherapy of cancer. *Digestion* 1996;57:17-21.
- Krenning EP, Bakker WH, Kooij PPM, et al. Somatostatin receptor scintigraphy with [^{111}In -DTPA-D-Phe 1]-octreotide in man: metabolism, dosimetry and comparison with [^{123}I -Tyr 3]-octreotide. *J Nucl Med* 1992;33:652-658.