lism of radiolabeled putrescine in gliomas. Implications for positron emission tomography of brain tumors. *Neurosurg* 1988;23:464–469.

- Shin WW, Fong WF, Pang SF, Wong P C-L. Limited blood-brain barrier transport of polyamines. J Neurochem 1985;44:1056-1059.
- Tabor CW, Tabor H. 1,4-Diaminobutane (putrescine), spermidine, and spermine. Ann Rev Biochem 1976;915:285-306.
- Ogawa T, Uemura K, Shishido F, et al. Changes of cerebral blood flow, and oxygen and glucose metabolism following radiochemotherapy of gliomas: a PET study. J Comp Assist Tomog 1988;12:290-297.
- Russell DH, Durie BGM. Polyamines as biochemical markers of malignancy. New York: Raven Press; 1978:15-41.
- Cavanaugh PF, Zlatko PP, Porter CW. Enhancement of 1,3-bis [2-chloroethyl]-1-nitrosourea-induced cytotoxicity and DNA damage by alpha-difluoromethylornithine in L1210 leukemia cells. *Cancer Res* 1984;44:3856– 3861.
- Pegg AE, McCann PP. Polyamine metabolism and function. Am J Physiol 1982;243:C212–C221.
- Hopewell JW. Late radiation damage to the central nervous system: a radiobiological interpretation. *Neuropath Applied Neurobiol* 1979;5:329– 349.

- Conomy JP, Kellermeyer RW. Delayed cerebrovascular consequences of therapeutic radiation. *Cancer* 1975;36:1702-1708.
- De Ruiter J, Van Putten LM. Measurement of blood flow in the mouse tail after irradiation. *Radiation Res* 1975;61:427-438.
- Butler AR, Horii SC, Kricheff II, Shannon MB, Budzilovich GN. Computed tomography in astrocytomas. A statistical analysis of the parameters of malignancy and the positive contrast-enhanced CT scan. *Radiology* 1978;129:433–439.
- Andreou J, George AE, Wise A, et al. Prognostic criteria of survival after malignant glioma surgery. AJNR 1983;4:488-490.
- Di Chiro G, Oldfield E, Wright DC, et al. Cerebral necrosis after radiotherapy and/or intraarterial chemotherapy for brain tumors: PET and neuropathologic studies. AJNR 1988;150:189-197.
- Valk PE, Budinger TF, Levin VA, Silver P, Gutin PH, Doyle WK. PET of malignant cerebral tumors after interstitial brachytherapy. Demonstration of metabolic activity and correlation with clinical outcome. J Neurosurg 1988;69:830-838.
- 40. Burger PC, Vogel FS, Green SB, Strike TA. Glioblastoma multiforme and anaplastic astrocytoma: pathologic criteria and prognostic implications. *Cancer* 1985;56:1106-1111.

EDITORIAL Carbon-11-Putrescine: Back to the Drawing Board

he oft-stated aim of PET neuro-**1** oncologists is to exploit metabolic differences between tumor tissue and surrounding normal brain in order to improve tumor localization and permit non-invasive determinations of tumor histology and growth rate that can be used to assess histological grade and response to therapy (1). As the article by Hiesiger et al. in the current issue of The Journal of Nuclear Medicine demonstrates, this laudable aim remains elusive: [¹¹C]putrescine, the high-profile PET brain-tumor tracer of the 1980s (2,3) has proved to be a disappointment in the 1990s. What lessons can be learned from the putrescine experience?

At first glance, the rationale for synthesizing ¹¹C-labeled putrescine seems unusually attractive. Endogenous putrescine, the immediate precursor of spermidine and spermine, is synthesized from ornithine by ornithine decarboxylase (ODC), the rate-limiting enzyme in polyamine synthesis (4,5). A second decarboxylase, S-adenosyl-L-methionine decarboxylase (SAMDC) catalyzes the formation of S-adenosyl-S-methylhomocysteamine, from which an aminopropyl moiety is transferred to putrescine to form spermidine, and to spermidine to form spermine (4). Whereas ODC activity and putrescine concentration are low in normal brain (1,4,6), elevated concentrations of di- and polyamines and their biosynthetic and catabolic enzymes have been reported in a wide variety of rapidly growing tissues, including primary and metastatic brain tumors (4-8). Finally, and perhaps most to the point, ODC activity, putrescine concentration and SAMDC activity in biopsy specimens of rat and human tumors, including gliomas, have been correlated with histopathological criteria of malignancy (5,6,9-11).

Although exogenously administered putrescine does not readily cross the intact blood-brain barrier (BBB), it rapidly traverses the more permeable blood-tumor barrier (2,3,12). Preliminary [¹⁴C]putrescine autoradiographic studies in T9-gliosarcomabearing rats indicated that target-tobackground (i.e., tumor-to-contralateral brain) concentration ratios as high as 35:1 were achievable and suggested that ¹¹C-labeled putrescine might serve as a "near ideal" PET tracer for the metabolic imaging of human brain tumors and, within the context of an appropriate pharmacodynamic model, as a marker for tumor growth rate (2). These high hopes were bolstered in 1987 by Hiesiger et al. (3), who reported in this journal that [¹¹C]putrescine PET studies of primary and metastatic brain tumors provided a better signal-to-noise ratio than glucose metabolic rate measurements obtained with [11C]2-deoxyglucose (¹¹CDG); Hiesiger et al. also anticipated that [¹¹C]putrescine would prove useful for locating small glycolytically hypometabolic lesions and would provide a quantitative index of degree of malignancy.

But doubts began to emerge, even as new claims for [¹¹C]putrescine were being made. In their 1987 Journal of Nuclear Medicine article, Hiesiger et al. grappled with the possibility that some or all of the observed tumor uptake of plasma ¹¹C radioactivity ([¹¹C]putrescine, ¹¹CO₂ and nonvolatile ¹¹C-labeled putrescine metabolites) was due to deficiency of the BBB, and that uptake of exogenous putrescine did not necessarily reflect the rate of tumor polyamine biosynthesis. In 1988, Warnick et al. reported surprisingly low in vivo rates of [exogenous] putrescine conversion to spermidine and spermine in T9 rat gliosarcoma

Received October 10, 1991; accepted October 10, 1991.

For reprints contact: D. A. Rottenberg, PET Imaging Service (11P), VA Medical Center, Minneapolis, Minnesota 55417.

and U-87 MG human glioblastoma (1). In order to explain these results, which seemed to contradict the findings of previous studies describing enhanced polyamine metabolism in rat and human gliomas, the authors conjectured that polyamine metabolism might vary with tumor growth fraction, and they discussed the importance of alternative pathways of putrescine metabolism (1,4,13) and the existence of multiple endogenous polyamine pools. In their current paper, Hiesiger et al. conclude that [¹¹C]putrescine uptake relates primarily to BBB breakdown rather than to tumor polyamine metabolism or mitotic activity and, therefore, "appears to have more limited diagnostic utility than ¹⁸FDG or ¹¹CDG."

In retrospect, it is not difficult to see where things went wrong. First, the lack of a comprehensive biophysical/biochemical model for [¹¹C]putrescine uptake, based on accepted in vitro and in vivo biochemical data and taking into account BBB transport, extra- and intracellular compartmentalization and tracer metabolism to polyamines, nonpolyamine metabolites and CO₂, led to ambiguous or uninterpretable PET results. Second, the graphical evidence of irreversible tumor uptake of [11C]putrescine, presented by Hiesiger et al. in their 1987 paper, was ultimately based on a single 50-min patient study in which, at 20 min after tracer injection, only 9% of the plasma ¹¹C radioactivity was identified by HPLC as putrescinethe remaining 91% was characterized as "[¹¹C]O₂ and nonvolatile metabolites." Third, the difficulties in interpretation occasioned by tumor heterogeneity (both histological and metabolic) and volume averaging, and the

limitations of data analytic strategies relying on average regional (region of interest) or peak values were not fully appreciated (14). Finally, insufficient attention was paid to the mechanism of putrescine uptake, the limitations of experimental brain-tumor models, and the relative merits of other nonglucose ¹¹C and ¹⁸F PET tracers employed for brain-tumor imaging (15-17).

In conclusion, the lessons learned from the [¹¹C]putrescine experience the importance of tracer kinetic modeling, the need for complete radiochemical characterization of the arterial input function, and the constraints imposed by in vivo tumor biology-need not be re-learned during the evaluation of each new braintumor radiotracer. Claims of diagnostic and prognostic utility for novel PET tracers must be evaluated within the context of the bedside neurological examination and the results of CT/ MRI scanning and standard neurodiagnostic tests. Given the biologic and metabolic diversity of human brain tumors-which extends to individual deposits-multidimenmetastatic sional prospective clinical trials will be required to establish meaningful estimates of in vivo sensitivity and specificity.

D. A. Rottenberg

VA Medical Center and University of Minnesota Minneapolis, Minnesota

REFERENCES

- Warnick RE, Pietronigro DD, McBride DQ, et al. In vivo metabolism of radiolabeled putrescine in gliomas: implications for positron emission tomography of brain tumors. *Neurosur*gery 1988;23:464–469.
- 2. Volkow N, Goldman SS, Flamm ES, et al.

Labeled putrescine as a probe in brain tumors. *Science* 1983;221:673-675.

- Hiesiger E, Fowler JS, Wolf AP, et al. Serial PET studies of human cerebral malignancy with [1-¹¹C]putrescine and [1-¹¹C]2-deoxy-Dglucose. J Nucl Med 1987;28:1251-1261.
- Seiler N. Polyamines. In: Lajtha A, ed. Handbook of neurochemistry, volume 1. New York: Plenum Press; 1982:223-255.
- Goldman SS, Volkow ND, Brodie J, Flamm ES. Putrescine metabolism in human brain tumors. J Neuro Oncol 1986;4:23-291.
- Harik SI and Sutton CH. Putrescine as a biochemical marker or malignant brain tumors. *Cancer Res* 1979;39:5010-5015.
- Snyder SH, Kreus DS, Medina VJ. Polyamine synthesis and turnover in rapidly growing tissues. Ann NY Acad Sci 1970;171:749-771.
- Seiler N, Lamberty U, Al-Therib MJ. Acetylcoenzyme A: 1,4-diaminobutane N-acetyltransferase: activity in rat brain during development, in experimental brain tumours and in brains of fish of different metabolic activity. J Neurochem 1975;24:797-800.
- Marton LJ, Heby O. Polyamine metabolism in tumor, spleen and liver of tumor-bearing rats. Int J Cancer 1974;13:619-628.
- Kremzner LT. Polyamine metabolism in normal and neoplastic neural tissue. In: Russel DH, ed. Polyamines in normal and neoplastic growth. New York: Raven Press; 1973:27-40.
- Glikman P, Vegh I, Pollina MA, Mosto A, Levy C. Ornithine decarboxylase activity, prolactin blood levels, and estradiol and progesterone receptors in human breast cancer. *Cancer* 1987;60:2237-2243.
- Shin W-W, Fong W-F, Pang S-F, et al. Limited blood-brain barrier transport of polyamines. J Neurochem 1985;1056-1059.
- Bachrach U. Metabolism of polyamines by cultured glioma cells: effect of asparagine on gamma-aminobutyric acid concentrations. *Biochem J* 1980;188:387-392.
- Tyler JL, Diksic M, Villemure J-G, et al. Metabolic and hemodynamic evaluation of gliomas using positron emission tomography. J Nucl Med 1987;28:1123-1133.
- Lilja A, Bergstrom K, Hartvig, P, et al. Dynamic study of supratentorial gliomas with Lmethyl-¹¹C-methionine and positron emission tomography. Am J Neuroradiol 1985;6:505-514.
- Junck L, Olson JMM, Ciliax BJ, et al. PET imaging of human gliomas with ligands for the peripheral benzodiazepine binding site. Ann Neurol 1989;26:752-758.
- Wienhard K, Herholz K, Coenen HH, et al. Increased amino acid transport into brain tumors measured by PET of L-(2-¹⁸F)fluorotyrosine. J Nucl Med 1991;32:1338-1346.

Correction

In the November 1991 issue of the *Journal*, the author line for "Noninvasive Delineation of the Effects of Moderate Aging on Myocardial Perfusion", by Senneff et al (pages 2037–2042) was printed incorrectly. It should read: Martha J. Senneff, Edward M. Geltman, and Steven R. Bergmann, with the technical assistance of Judy Hartman.