

categorically, "that the steady-state  $^{15}\text{O}_2$  method cannot provide quantitative values of regional oxygen utilization" (5), but we did indeed state that such measurements may be possible to carry out with reasonable accuracy for tissues that exchange water slowly.

The theoretical model using the Fick principle (6-8) suggested by Jones et al. (9) for measurement of regional cerebral blood flow (rCBF) and oxygen extraction (OER), has been applied, in conjunction with quantitative positron emission tomography, to in vivo studies as well as extended to regional cerebral oxygen consumption rates (rCMRO<sub>2</sub>) (10) by means of their product, i.e.,  $\text{rCMRO}_2 = \text{rCBF} \times \text{OER} \times (\text{total blood oxygen content})$ . We use a compartmental-model approach (11) to analyze the possibility of the direct measurement of regional oxygen utilization by quantitative methods, e.g., our earlier two-dimensional (12,13) approach or now the more powerful three-dimensional approach provided by positron tomography. The clinical value of the steady-state technique will depend upon an understanding of the nature and magnitude of the uncertainties associated with the measurements and approximations used. In our opinion this understanding is facilitated by our work (1) and that of others (14-16), as well as by the material referred to by Jones et al. (5). One of us (REB) plans to carry out an error analysis similar to that for rCBF by SC Jones, JH Greenberg and M Reivich (15) in order to provide a more thorough understanding of the errors associated with the measurement of regional oxygen metabolism and OER in humans by the approach detailed by Frackowiak et al. (10).

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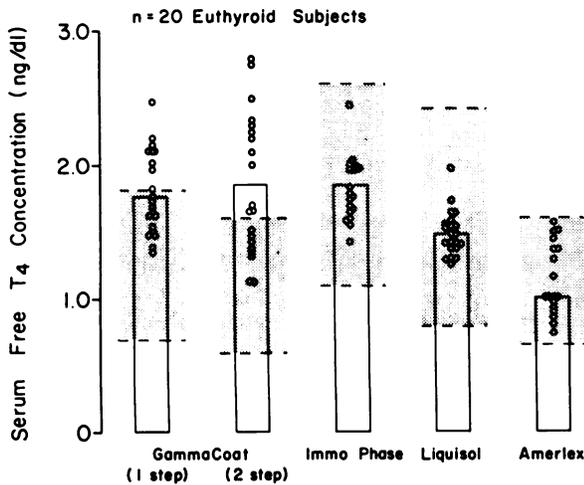
### Free Thyroxine RIA Concentration (GammaCoat) Is Spuriously Elevated in Blood Collected in Silicon-Coated Vacutainer Tubes

An unusual finding was observed when five radioimmunoassay (RIA) kits were used to measure the serum free thyroxine concentration (FT<sub>4</sub>) in euthyroid subjects during a study to evaluate the serum FT<sub>4</sub> in patients with familial dysalbuminemic hyperthyroxinemia (FDH) (1-3). This syndrome is characterized by elevations in the serum T<sub>4</sub> concentration and free thyroxine index due to the presence of an abnormal serum albumin that preferentially binds T<sub>4</sub>. However, these patients are euthyroid and the serum T<sub>3</sub> and FT<sub>4</sub> concentrations are normal when measured by equilibrium dialysis.

FT<sub>4</sub> concentrations in serum samples from 20 normal, euthyroid subjects were measured by RIA. The following methods were used: I-125-labeled T<sub>4</sub> analog, antibody-coated tube (GammaCoat, one-step)\*; antibody-coated tube (GammaCoat, two-step)\*; antibody-coated microfine silica (Immo Phase†); microencapsulated antibody (Liquisol)‡; and I-125-labeled T<sub>4</sub> analog (Amerlex). All blood samples had been collected in silicon-coated vacutainer tubes and the serum stored frozen in the silicon-coated tubes. All tests were performed in duplicate on the same day.

The results of the serum FT<sub>4</sub> measured by the five methods in 20 euthyroid subjects are shown in Fig. 1. All values were in the normal range when assayed by the Immo Phase, Liquisol, and Amerlex methods. In contrast, serum FT<sub>4</sub> values were above the normal range in nine of the 20 normal subjects when measured by the GammaCoat, one-step kit and in 12 subjects measured by the GammaCoat, two-step kit. The tests were repeated using other GammaCoat kits, and similar results were obtained. Therefore, these observations could not be explained on the basis of defective reagents or experimental error.

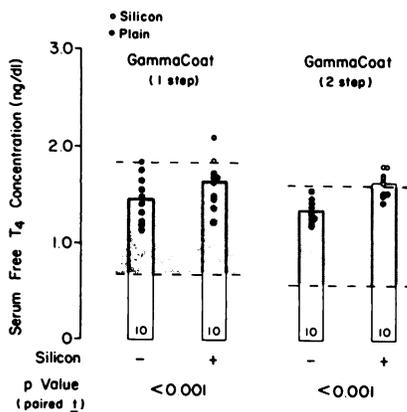
One explanation for the elevated values obtained by the GammaCoat methods could be that the silicon used to coat the vacutainer tubes might interfere with the assays. To test this hypothesis, blood samples were collected from ten more euthyroid subjects in vacutainer tubes both with and without silicon coating, and serum was stored frozen for 3 days in the respective tubes. Serum FT<sub>4</sub> was measured in duplicate by both GammaCoat kits in the sera



**FIG. 1.** Serum FT<sub>4</sub> (RIA) in 20 normal subjects measured by five commercial kits. Shaded areas indicate normal range for each test, and bars show mean values for each test.

collected and stored in both kinds of tubes. All samples were evaluated in one assay. The results are shown in Fig. 2. Values for FT<sub>4</sub> in serum samples collected in tubes without silicon coating were in the normal range in both kits. The FT<sub>4</sub> was always higher in both GammaCoat methods in serum collected and stored in silicon-coated tubes, as compared with values in serum collected and stored in tubes without silicon coating ( $p < 0.001$ , paired t-test). In the GammaCoat one-step assay, two of ten samples were above the normal range, and in the GammaCoat two-step assay, six of ten samples were.

These findings suggest that blood collected in at least some batches of silicon-coated vacutainer tubes result in spuriously elevated FT<sub>4</sub> values when measured by either the one-step or two-step GammaCoat methods. The explanation for these observations remains unknown. Since silicon-coated tubes do not appear to affect other methods for measuring FT<sub>4</sub> by RIA, it seems likely that silicon must specifically interfere with the assay involving the antibody-coated RIA tubes. It is suggested, therefore, that until



**FIG. 2.** Serum FT<sub>4</sub> (RIA) in ten normal subjects measured by GammaCoat one-step and GammaCoat two-step kits. Black dots represent blood samples collected in tubes not silicon coated; open dots represent blood samples collected in silicon-coated vacutainer tubes. Shaded areas indicate normal range of each test; bars show mean values for each test; numbers in the bars show numbers of subjects evaluated. Statistical analyses are performed by paired t-test.

further studies are carried out, tubes should not be silicon coated if used in collecting blood and storing serum for measurement of FT<sub>4</sub> with the GammaCoat kits.

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FOOTNOTES

- \* Clinical Assays, Division of Travenol Laboratories, Cambridge, MA.
- † Corning Medical, Medfield, MA.
- ‡ Damon Diagnostic, Needham Heights, MA.
- § Radiochemical Center, Amersham, U.K.

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**Automatic Edge Detection of Scintigraphic Images**

It may be appropriate now to reflect on the great efforts that are being made in automatic edge detection of scintigraphic images, especially in relationship to cardiac left ventricular wall motion studies. The academic exercise is noble, but we should consider what the thrust of the effort is. If the thrust is to automate the study so that minimal intervention of a trained physician is required, the experimental studies should include an indication of the time saved compared with manual methods for defining the left ventricular wall. They should be done in an average clinical setting that includes patients with tortuosity of the aorta, with abnormal contours of underlying left atria, and with activity of radiopharmaceuticals in other organs that may be included in the frame.

If the thrust is toward a more uniform computer treatment of the studies, the experiment must show us variance statistics demonstrating uniformity or nonuniformity of the technique in comparison with the manual method. Alternatively, if we are attempting to show greater accuracy in the technique, comparison of the edge detection algorithm, the manual technique, and a ventriculography gold standard should be made.

Unfortunately, in the effort to make progress in automated