Evaluation of ^{18}F -RO-948 PET for Quantitative Assessment of Tau Accumulation in the Human Brain

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The availability of tau PET radioligands enables quantitative assessment of tau density and distribution in the human brain. We evaluated the kinetics of a novel radioligand, ^{18}F -RO-948 (previously referred to as ^{18}F -RO6958948), and its ability to identify tau positivity in individual patients with mild Alzheimer disease (AD). **Methods:** Eleven subjects with amyloid-positive mild AD, 5 amyloid-negative older control subjects (OC), and 5 younger control subjects (YC) completed 1 or 2 (4 AD and 5 OC) PET scans with ^{18}F -RO-948 for 90, 120, or 200 min. The kinetics of the radioligand was evaluated with standard compartmental and noncompartmental models (with plasma data in 70% of cases), tissue-reference methods, and SUV ratio. These approaches were applied to assess the ability of ^{18}F -RO-948 to discriminate AD subjects from OC subjects. **Results:** The plasma reference graphical analysis appeared to be the optimal method of quantification for ^{18}F -RO-948, yielding strictly time-consistent values of distribution volume and distribution volume ratio at 90 min against the analyses at 120 and 200 min. The reference tissue graphical analysis and SUV ratio were cross-validated against plasma reference graphical analysis. Test-retest evaluation showed excellent reproducibility. A proposed novel index of tau load, the regional tau-positive fraction, showed high values in the medial and lateral temporal and parietal regions in AD and successfully separated AD subjects from OC and YC subjects with a significant margin. **Conclusion:** ^{18}F -RO-948 appears to be a promising radioligand for quantitative imaging of tau in the brain of AD patients.

Key Words: neurology; PET; radiotracer tissue kinetics; Alzheimer disease; ^{18}F -RO6958948 for tau imaging; radiotracer tissue kinetics

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Recently, interest in neuroimaging of tauopathies with PET has grown within the Alzheimer disease (AD)/dementia research

community, complementing the imaging of amyloid- β ($\text{A}\beta$) (1,2). Models of AD progression based on neuropathologic studies (3–5) are being examined with ^{18}F -AV1451 PET imaging (6,7). Independently of these models, distribution patterns of tau in early-AD subjects (8) and in subtypes of AD (9) are being evaluated with ^{18}F -AV1451. In addition, it has been reported that PET is able to detect progression of tau pathology in AD over intervals of as short as 6 mo (10).

We have developed ^{18}F -RO-948 (previously referred to as ^{18}F -RO6958948) for imaging tau aggregates in the human brain (11,12). The ligand has high affinity and specificity for tau aggregates in vitro while lacking affinity for $\text{A}\beta$ plaques (11). Autoradiographic studies indicate that ^{18}F -RO-948 binds with high affinity to tau aggregates in AD brain sections whereas lower reactivity is observed in non-AD tauopathies (11), suggesting that ^{18}F -RO-948 primarily recognizes a mixture of 3- and 4-repeat tau isoforms.

In this study, we evaluated the kinetics of ^{18}F -RO-948 in the brain of AD and younger and older control subjects (YC and OC subjects, respectively) to identify optimal modeling approaches, outcome variables, and tracer circulation time. We also evaluated reproducibility with test and retest scans. Imaging outcome measures with good reproducibility will be key to understanding the natural progression of tau deposition in patients and assessing the effects of tau-targeting therapies.

One of the primary purposes of neuroimaging of tauopathies is to accurately identify tau positivity in individual scans. Analogous schemes have been established for $\text{A}\beta$ imaging (13,14); however, SUV ratio (SUVR)-based attempts (15,16) have not yet been established for tau tracers. In this paper, we introduce a novel index of tau load, the regional tau-positive fraction (TPF). It was expected that the binary nature of TPF (positive or negative) could make it very sensitive in detecting initial increases of tau accumulation in AD compared with published quantitative approaches (15,16). In addition, after recent reports (17,18) suggesting the presence of off-target binding for ^{18}F -AV1451, we carefully examined the binding of ^{18}F -RO-948 in YC and OC subjects.

MATERIALS AND METHODS

Subjects

After giving written informed consent, 11 subjects with AD, 5 YC subjects, and 5 OC subjects entered this study, which was approved by the institutional review board of the Johns Hopkins University, School of Medicine. Detailed inclusion and exclusion criteria are listed in a companion paper (12). Briefly, the YC subjects were less than 40 y old and had no neuropsychiatric disorders. The OC subjects were more

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TABLE 1
Demographic and Scan Information

Group	Age (y)	Sex	MMSE	SUVR*	Type	Scans (n) [†]			Injected-activity (MBq)	Mass (μg)
						90 min	120 min	200 min		
AD	69 ± 9	8 M, 3 F	16–25	¹¹ C-PIB: 1.8–2.8	Test	8/11	3/5	1/2	240 ± 86	0.08 ± 0.05
				¹⁸ F-AV45: 1.6–2.6	Retest	3/5	3/4	1/2		
OC	63 ± 9	5 M	>28	¹¹ C-PIB: 1.3–1.4	Test	4/5	4/5	1/2	235 ± 92	0.12 ± 0.20
					Retest	3/5	3/5	0/2		
YC	29 ± 5	4 M, 1 F	Not done	Not done	Test	4/5	—	—	252 ± 222	0.15 ± 0.05

*Ranges of 80 brain regions of individual subjects.

[†]Scans with plasma data/total scans, by circulation time.

MMSE: Mini-mental state examination scores; PIB = Pittsburgh compound B.

Data are ranges or mean ± SD.

than 50 y old, cognitively normal (mini-mental state examination score > 28), and Aβ-negative based on scans obtained with ¹¹C-Pittsburgh compound B or ¹⁸F-AV45. The AD subjects were more than 50 y old, had a clinical diagnosis of AD, scored between 16 and 26 on the mini-mental state examination, and were Aβ-positive based on scans obtained with ¹¹C-Pittsburgh compound B or ¹⁸F-AV45.

PET and MRI Procedures

PET studies were performed on a high-resolution research tomograph (Siemens). Preparation included insertion of an intravenous catheter for ligand injection and, in selected subjects, a radial arterial catheter for blood sampling. Each subject had a custom-fitted thermoplastic facemask to reduce head motion during the PET study. A 6-min attenuation scan was performed using a rotating ¹³⁷Cs point source. Starting with a slow-bolus injection of ¹⁸F-RO-948, the emission scan lasted for 90, 120, or 200 min in 3-dimensional list mode.

Short restroom breaks (<5 min) were allowed, and a 20-min break (between 120 and 140 min after injection) was scheduled for 200-min scans. In subjects with arterial catheters, blood samples were obtained throughout the scan, approximately every 5 s initially and increasing to every 5 min after 30 min. The radioactivity in plasma was measured with a cross-calibrated γ-counter. Radioactive species in plasma were determined with high-pressure liquid chromatography on selected plasma samples (2, 5, 10, 20, 30, 45, and 90 min and every 30 min thereafter). Each subject was carefully monitored for subjective symptoms together with vital signs and electrocardiography throughout the scan.

The emission scans were reconstructed (19), correcting for attenuation, scatter, and dead time. The frame schedules were four 15-s, four 30-s, three 1-min, two 2-min, and five 4-min frames up to 30 min, and 5-min frames thereafter to make totals of 30, 36, or 52 frames for the 90-, 120-, and 200-min scans, respectively. Each PET frame consisted of 256 by 256 by 207 (axial) voxels. The final spatial resolution was expected to be about 2.3 mm in full width at half maximum in 3 directions (19). In interrupted scans, separate transmission scans were used for pre- and postinterruption emission scans. PET frames were aligned to the frame with the highest non-decay-corrected count using the coregistration module of SPM12 (20) (<http://www.fil.ion.ucl.ac.uk/spm/>) to reduce the effects of head motion during the scan and head motion due to interruptions. Spoiled gradient sequence MR images were obtained with a 3-T Trio Tim system (Siemens) for each subject for anatomic identification of the structures of interest.

Volumes of Interest (VOIs) and Generation of Time-Activity Curves

VOIs of 40 brain regions (80 left and right VOIs) were generated with the software library tools of the Oxford Center for Functional MRI of the Brain (21,22) (subcortical) and with Freesurfer software (23). The VOIs were refined on the MR image using a locally developed VOI tool. VOIs for the cerebellar gray matter were defined manually, excluding the vermis and anterior lobules. Voxels within

FIGURE 1. Plots of time-activity curves (top) and time profiles of SUVR (bottom) of ¹⁸F-RO-948 in selected brain regions for representative subjects. Solid lines = left side of brain; dotted lines = right side of brain.

FIGURE 2. Images of SUVR (60–90 min) of ^{18}F -RO-948 for representative YC, OC, and 3 levels (by mean SUVR) of AD subjects. Transaxial images (top) show medial and lateral temporal regions, and coronal images (bottom) were cut through uncus. Maximal SUVR for display was set at 6. MMSE = mini-mental state examination score.

6 mm dorsally were excluded to reduce contamination from supratentorial structures. The VOIs were spatially transferred to the PET space (20) and applied to successive PET frames to generate time–activity curves. Plasma data were decay-corrected to the injection time. Metabolite-corrected plasma time–activity curves were obtained by applying high-pressure liquid chromatography data to total plasma time–activity curves as previously reported (24).

Evaluation of Kinetic Approaches

Applicability of the 1- and 2-tissue-compartment models and plasma reference graphical analysis (25) (PRGA) was examined. The Akaike information criterion (26) was used to examine whether the 1-tissue-compartment model could be justified over the 2-tissue-

compartment model (27). The outcome variables were distribution volume (V_T) and distribution volume ratio (DVR). Tracer circulation times of 90, 120, and 200 min were used in the tracer evaluation.

Reference tissue graphical analysis (28) (RTGA) and the multilinear reference tissue method with 2 parameters (29) were also evaluated (outcome variable: DVR). SUVR was obtained as the target-to-cerebellar gray matter ratio at 60–90 min after injection.

Test–Retest Evaluation

Test–retest variability (given as a percentage) and interclass correlation coefficient were calculated in individual regions as the mean of [test – retest] over the means of test and retest values across subjects to evaluate the reproducibility of PET outcome variables. A level of 10% test–retest variability was considered acceptable (30). Test–retest variability was calculated separately for AD and OC subjects because of expected differences in value ranges.

Identification of Tau-Positive Individuals

TPF was defined as the fraction of voxels of a given VOI that exceeded the mean SUVR plus 2.5 SDs of the same VOI in YC and OC subjects. Mean, composite, and maximal TPFs, and conventional global mean SUVRs or mean SUVRs of composite regions (15), were evaluated for their ability to discriminate AD from OC and YC subjects.

Statistics

Values are expressed as mean and SD, when applicable. To compare groups, 2-way (group $\times$ region) ANOVA was performed first. When either effect was shown significant, subsequent t testing was performed with the significance level set at a P value of less than 0.05 and with Bonferroni adjustment for multiregion comparisons.

RESULTS

Demographic and scan information on the participants is shown in Table 1. There were no statistical differences in injected radioactivity or mass between groups. Three 120-min scans were terminated early (at 75 and 85 min) or interrupted (for <5 min) because of restroom breaks, resulting in varying numbers of scans for circulation times of 90, 120, and 200 min (Table 1).

Tissue time–activity curves peaked within 5 min of the tracer injection (Fig. 1). Thereafter, the time–activity curves of AD subjects showed varying degrees of retention across regions and subjects, whereas the time–activity curves of OC and YC subjects showed rapid clearance across regions. In YC and OC subjects, SUVR, or frame-by-frame SUVR, became stable after approximately 60 min, except for putamen, whose SUVR slightly declined at later times. In AD subjects, SUVR approached plateaus of varying heights by 150 min in the subject who showed the highest SUVR. YC and OC subjects showed a relatively homogeneous radioactivity distribution at 60–90 min (Fig. 2), whereas AD subjects showed a heterogeneous

FIGURE 3. Plots of observed and model-predicted tissue time–activity curves (TACs) of 2-tissue-compartmental model (A), PRGA (B), multilinear reference tissue method with 2 parameters (C), and RTGA (D) for AD subjects who had highest mean SUVRs with (A and B) and without (C and D) arterial plasma data. A(T), R(T), and C(T) stand for TACs in target and reference regions and in plasma, and k_2^B for the brain-to-blood clearance constant of the reference region.

FIGURE 4. (A) Scatterplot of regional DVR data for PRGA at 90 min (y) vs. 120 and 200 min. (B and C) Scatterplots of regional DVR data for RTGA (y) vs. PRGA at 90 and 120 min (B) and for RTGA at 90 min (y) vs. 120 and 200 min (C). (D) Scatterplot of SUVR data (y) vs. DVR data for RTGA at 90 and 120 min (D). Regression equations are shown in keys.

pattern of tracer retention in brain regions known to exhibit tau pathology in AD subjects. No off-target binding was noted in the basal ganglia, thalamus, choroid plexus, or white matter (pons, 0.98 ± 0.10 ; corpus callosum, 0.93 ± 0.08 [mean $\pm$ SD of SUVR of AD]); some retention was evident in the substantia nigra. In 1 control subject, tracer retention was observed in the cerebellar vermis. Extracerebral retention was occasionally observed in the meninges and often in the retina.

Total plasma time-activity curves peaked within 1 min of the tracer injection, cleared rapidly until about 5 min, remained stable toward 20 min, and slowly decreased thereafter (Supplemental Fig. 1A; supplemental materials are available at <http://jnm.snmjournals.org>). Parent fractions in plasma were $16.0\% \pm 5.2\%$ at 30 min and $9.9\% \pm 3.5\%$ at 90 min (Supplemental Fig. 1B). In longer scans, parent fractions were $7.3\% \pm 2.9\%$ at 120 min and $2.1\% \pm 0.9\%$ at 200 min. Because of the metabolism, plasma time-activity curves of the parent compound decreased monoexponentially after the peaks.

Plasma Input Methods

On visual inspection, model-predicted time-activity curves aligned well with observed time-activity curves across frames for the 2-tissue-compartment model (Fig. 3A) but not for the 1-tissue-compartment model (not shown). In agreement, the 2-tissue-compartment model showed lower Akaike information criterion values in 95% of regions. These findings support the use of the 2-tissue-compartment model over the 1-tissue-compartment model for ^{18}F -RO-948. Plots of PRGA approached asymptotes by 20 min (t^*) in AD and OC subjects and showed excellent linearity ($R^2 > 0.98$) in more than 98% of regions for circulation times of 90, 120, and 200 min (Fig. 3B). Although PRGA (y) yielded tightly correlated V_T values to the 2-tissue-compartment model at 90 min ($y = 0.87 \cdot x + 0.60$; $R^2 = 0.942$), the

2-tissue-compartment model yielded sporadic outlier values (incidence: 0.7% of regions; outliers: 1 range below or above the observed range of RTGA values). Regional V_T and DVR data at 90 min were tightly proportional (i.e., regression lines passed through the theoretic origin) to respective data at 120 and 200 min (Fig. 4A; Supplemental Fig. 2A), although slopes minimally decreased as a function of circulation time. Very similar regression results were obtained when data of the OC, YC, and AD groups were pooled (Supplemental Figs. 2B and 2C). These findings establish the timewise consistency of V_T and DVR across the 90-, 120-, and 200-min analyses, which validated the 90-min dynamic scans for derivation of V_T and DVR for ^{18}F -RO-948.

Reference Tissue Methods

The brain-to-blood transfer rate constant of the reference region was estimated from the intercepts of PRGA (25) and set at 0.05 min^{-1} , and t^* was set at 15 min after R^2 was confirmed to be more than 0.98 in over 95% of regions. RTGA yielded DVRs essentially identical to PRGA at 90 and

120 min for DVRs of less than 3.5 and showed a trend to slightly overestimate for DVRs above 3.5 (Fig. 4B). These results validated the RTGA by agreement with PRGA. RTGA also yielded strictly proportional DVRs at 90 min against the 120- and 200-min analyses (C). The multilinear reference tissue method with 2 parameters showed statistically indistinguishable correlations to PRGA and timewise consistency with RTGA (data not shown).

FIGURE 5. Box plot of regional test-retest variability estimates for PRGA and RTGA (variable: DVR) and SUVR for AD subjects in regions where mean SUVR exceeded 1.5 (56 left and right regions). Boxes indicate median value and upper 25th and 75th percentiles, and whiskers indicate highest and lowest values, excluding outliers (black dots; data points exceeding ± 2.7 SDs). Dotted line indicates desired test-retest variability level of 10%.

TABLE 2
Forty Brain Regions in Descending Order of SUVR of ^{18}F -RO-948

Region	Abbreviation	SUVR rank of ^{18}F -AV1451 (18)	Composite region*
Inferior parietal lobe	iPa	1, 2	X
Inferior temporal lobe	iTp	6	X
Middle temporal lobe	mTp	5	X
Supramarginal region	SM	10	
Fusiform gyrus	Fs	16	X
Lateral occipital lobe	IOc	7, 9, 12	
Precuneus	Pr	4	
Middle frontal lobe	mFr	11	
Superior parietal lobe	sPa	3	X
Rostral frontal lobe	rFr		
Frontal operculum	FO	19	
Pars triangularis	PT		
Isthmus/cingulate	IC	20	
Orbital operculum	OO		
Posterior cingulate	pCg	8	
Parahippocampus	PH		X
Entorhinal area	ER	13	X
Cuneus	Cu	15	
Lingular gyrus	LG		
Lateral orbital gyrus	IOG		
Superior temporal lobe	sTp	14	X
Amygdala	Am		X
Anterior temporal lobe	aTp	17, 18	
Superior frontal lobe	sFr	16	
Postcentral gyrus	PS		
Anterior frontal lobe	aFr		
Precentral gyrus	Pc		
Medial orbital gyrus	mdOG		
Insula	In		
Pericalcarine cortex	pC		
Paracentral region	pC		
Caudal anterior cingulate	cAC		
Rostral anterior cingulate	rAC		
Putamen	Pu		
Hippocampus	Hp		X
Ventral striatum	vS		
Globus pallidus	GP		
Transverse temporal	tT		
Thalamus	Th		
Caudate nucleus	CN		

*Regions for composite SUVR and TPF calculation.

shown). Binding to melanin has been reported also for other tau tracers (15).

Among quantitative approaches with ^{18}F -AV1451, the mean SUVR of composite regions (15,16) failed to discriminate individual AD subjects from control subjects as accurately as other

regionally based methods. The composite SUVR of ^{18}F -RO-948 showed similar, suboptimal separation capabilities. Maximal TPF appeared to be promising for identification of tau positivity in individual scans, showing a significant margin between AD and control subjects. The progressive improvement could be explained

FIGURE 7. (A) Scatterplots of composite SUVR and TPFs, and maximal tau-positive fractions of individual subjects in baseline (dots) and retest (x) scans of AD and OC + YC subjects. (B) Line plots of TPRs and maximal SUVRs of individual regions (showing highest 23 regions alone) for 5 subjects who showed lowest composite SUVR and TPRs. Three regions (per side of brain) that had highest TPF or relative SUVRs are separately shown by circles and denoted in key as Top 3. (C) SUVR images of 3 mildest cases showing locations where maximal SUVRs were observed. Key to abbreviated brain regions is in Table 2.

by the progressively narrower AD distributions along the total observed ranges from composite SUVR to composite FTP to maximal TPF. The analysis of regional TPF and relative maximal SUVR suggests that initial tau deposition in AD could start not only in the region of the mesial (amygdala, entorhinal area, fusiform gyrus, and parahippocampus) and lateral inferior temporal lobe but also in the lateral parietal areas (superior parietal lobe and supramarginal), thus supporting the notion of anatomic variability in the pattern of tau deposition (9). Similar patterns of regional involvement in cases of very mild AD were reported with ^{18}F -AV1451 (15) by means of visual inspection alone. Therefore, analysis of regional TPF and relative maximal SUVR could be useful for improving our understanding of the progression of tau pathology in various subtypes of AD.

This study had some limitations. First, the findings for group comparisons should be considered preliminary because of the limited numbers of subjects. Accordingly, the usefulness of TPF should be examined further with larger numbers of AD subjects at different stages of the disease. Second, the current scan duration

(200 min) included DVRs of up to 3.5 and SUVRs of up to 4.5. Although these ranges are expected to be sufficient for most applications, the effect of scan duration on quantification may need to be further validated using subjects with more advanced tau accumulations.

CONCLUSION

^{18}F -RO-948 is a promising radioligand for imaging tau pathology in AD, showing excellent kinetic properties and displaying little off-target binding. ^{18}F -RO-948 SUVRs showed excellent reproducibility, and SUVR-derived variables appeared to be promising for identification of tau positivity in individual scans and for characterization of progression of tauopathies in AD.

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REFERENCES

1. Villemagne VL, Fodero-Tavoletti MT, Masters CL, Rowe CC. Tau imaging: early progress and future directions. *Lancet Neurol.* 2015;14:114–124.
2. Iqbal K, Liu F, Gong CX. Tau and neurodegenerative disease: the story so far. *Nat Rev Neurol.* 2016;12:15–27.
3. Braak H, Braak E. Diagnostic criteria for neuropathologic assessment of Alzheimer's disease. *Neurobiol Aging.* 1997;18(suppl):S85–S88.
4. Delacourte A, David JP, Sergeant N, et al. The biochemical pathway of neurofibrillary degeneration in aging and Alzheimer's disease. *Neurology.* 1999; 52:1158–1165.
5. Braak H, Alafuzoff I, Arzberger T, Kretschmar H, Del Tredici K. Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathol (Berl).* 2006;112:389–404.
6. Schwarz AJ, Yu P, Miller BB, et al. Regional profiles of the candidate tau PET ligand ¹⁸F-AV-1451 recapitulate key features of Braak histopathological stages. *Brain.* 2016;139:1539–1550.
7. Schöll M, Lockhart SN, Schonhaut DR, et al. PET imaging of tau deposition in the aging human brain. *Neuron.* 2016;89:971–982.
8. Johnson KA, Schultz A, Betensky RA, et al. Tau positron emission tomographic imaging in aging and early Alzheimer disease. *Ann Neurol.* 2016;79:110–119.
9. Ossenkoppele R, Schonhaut DR, Scholl M, et al. Tau PET patterns mirror clinical and neuroanatomical variability in Alzheimer's disease. *Brain.* 2016;139: 1551–1567.
10. Ishiki A, Okamura N, Furukawa K, et al. Longitudinal assessment of tau pathology in patients with Alzheimer's disease using [¹⁸F]THK-5117 positron emission tomography. *PLoS One.* 2015;10:e0140311.
11. Honer H, Gobbi L, Knust H, et al. Preclinical evaluation of ¹⁸F-RO6958948, ¹¹C-RO6924963 and ¹¹C-RO6931643 as novel radiotracers for imaging aggregated tau in AD with positron emission tomography. *J Nucl Med.* 2018;59:675–681.
12. Wong DF, Comley RA, Kuwabara H, et al. First in-human PET study of 3 novel tau radiopharmaceuticals: [¹¹C]RO6924963, [¹¹C]RO6931643, and [¹⁸F]RO6958948. *J Nucl Med.* May 4, 2018 [Epub ahead of print].
13. Villemagne VL, Burnham S, Bourgeat P, et al. Amyloid beta deposition, neurodegeneration, and cognitive decline in sporadic Alzheimer's disease: a prospective cohort study. *Lancet Neurol.* 2013;12:357–367.
14. Grundman M, Pontecorvo MJ, Salloway SP, et al. Potential impact of amyloid imaging on diagnosis and intended management in patients with progressive cognitive decline. *Alzheimer Dis Assoc Disord.* 2013;27:4–15.
15. Pontecorvo MJ, Devous MDS, Navitsky M, et al. Relationships between flortaucipir PET tau binding and amyloid burden, clinical diagnosis, age and cognition. *Brain.* 2017;140:748–763.
16. Jack CR Jr, Wiste HJ, Weigand SD, et al. Defining imaging biomarker cut points for brain aging and Alzheimer's disease. *Alzheimers Dement.* 2017;13:205–216.
17. Vemuri P, Lowe VJ, Knopman DS, et al. Tau-PET uptake: regional variation in average SUVR and impact of amyloid deposition. *Alzheimers Dement (Amst).* 2016;6:21–30.
18. Kantarci K, Lowe VJ, Boeve BF, et al. AV-1451 tau and beta-amyloid positron emission tomography imaging in dementia with Lewy bodies. *Ann Neurol.* 2017;81: 58–67.
19. Rahmim A, Cheng JC, Blinder S, Camborde ML, Sossi V. Statistical dynamic image reconstruction in state-of-the-art high-resolution PET. *Phys Med Biol.* 2005;50:4887–4912.
20. Ashburner JFK. Rigid body registration. In: Frackowiak RSJ, Ashburner J, Penny WD, et al, eds. *Human Brain Function*. Cambridge, MA: Academic Press; 2004:635–654.
21. Smith SM, Jenkinson M, Woolrich MW, et al. Advances in functional and structural MR image analysis and implementation as FSL. *Neuroimage.* 2004;23(suppl): S208–S219.
22. Patenaude B, Smith SM, Kennedy DN, Jenkinson M. A bayesian model of shape and appearance for subcortical brain segmentation. *Neuroimage.* 2011;56:907–922.
23. Fischl B, van der Kouwe A, Destrieux C, et al. Automatically parcellating the human cerebral cortex. *Cereb Cortex.* 2004;14:11–22.
24. Kuwabara H, Gao Y, Stabin M, et al. Imaging alpha4beta2 nicotinic acetylcholine receptors (nAChRs) in baboons with [¹⁸F]XTRA, a radioligand with improved specific binding in extra-thalamic regions. *Mol Imaging Biol.* 2017;19:280–288.
25. Logan J, Fowler JS, Volkow ND, et al. Graphical analysis of reversible radioligand binding from time-activity measurements applied to [¹¹C-methyl]-(-)-cocaine PET studies in human subjects. *J Cereb Blood Flow Metab.* 1990; 10:740–747.
26. Akaike H. A new look at the statistical model identification. *IEEE Trans Automatic Control.* 1974;19:716–723.
27. Koeppe RA, Holthoff VA, Frey KA, Kilbourn MR, Kuhl DE. Compartmental analysis of [¹¹C]flumazenil kinetics for the estimation of ligand transport rate and receptor distribution using positron emission tomography. *J Cereb Blood Flow Metab.* 1991;11:735–744.
28. Logan J, Fowler JS, Volkow ND, Wang GJ, Ding YS, Alexoff DL. Distribution volume ratios without blood sampling from graphical analysis of PET data. *J Cereb Blood Flow Metab.* 1996;16:834–840.
29. Ichise M, Liow JS, Lu JQ, et al. Linearized reference tissue parametric imaging methods: application to [¹¹C]DASB positron emission tomography studies of the serotonin transporter in human brain. *J Cereb Blood Flow Metab.* 2003;23:1096–1112.
30. Tolboom N, Yaqub M, Boellaard R, et al. Test-retest variability of quantitative [¹¹C]PIB studies in Alzheimer's disease. *Eur J Nucl Med Mol Imaging.* 2009; 36:1629–1638.
31. Barret O, Alagille D, Sanabria S, et al. Kinetic modeling of the tau PET tracer ¹⁸F-AV-1451 in human healthy volunteers and Alzheimer's disease subjects. *J Nucl Med.* 2017;58:1124–1131.
32. Hahn A, Schain M, Erlandsson M, et al. Modeling strategies for quantification of in vivo ¹⁸F-AV-1451 binding in patients with tau pathology. *J Nucl Med.* 2017;58: 623–631.
33. Ng KP, Pascoal TA, Mathotaarachchi S, et al. Monoamine oxidase B inhibitor, selegiline, reduces ¹⁸F-THK5351 uptake in the human brain. *Alzheimers Res Ther.* 2017;9:25.
34. Jonasson M, Wall A, Chiotis K, et al. Tracer kinetic analysis of (S)-¹⁸F-THK5117 as a PET tracer for assessing tau pathology. *J Nucl Med.* 2016;57:574–581.
35. Harada R, Okamura N, Furumoto S, et al. Characteristics of tau and its ligands in PET imaging. *Biomolecules.* 2016;6:7.
36. Maruyama M, Shimada H, Suhara T, et al. Imaging of tau pathology in a tauopathy mouse model and in Alzheimer patients compared to normal controls. *Neuron.* 2013;79:1094–1108.
37. Kimura Y, Ichise M, Ito H, et al. PET quantification of tau pathology in human brain with ¹¹C-PBB3. *J Nucl Med.* 2015;56:1359–1365.
38. Passamonti L, Vazquez Rodriguez P, Hong YT, et al. ¹⁸F-AV-1451 positron emission tomography in Alzheimer's disease and progressive supranuclear palsy. *Brain.* 2017;140:781–791.
39. Vermeiren C, Motte P, Viot D, et al. The tau positron-emission tomography tracer AV-1451 binds with similar affinities to tau fibrils and monoamine oxidases. *Mov Disord.* 2018;33:273–281.